

Effect of different crate material types for transit on production, physiological characteristics, and welfare of broilers during the summer season

Myunghwan Yu[#], Nuwan Chamara Chathuranga[#], Elijah Ogola Oketch, Jun Seon Hong, Haeun Park, Jung Min Heo^{*}

Department of Animal Science and Biotechnology, Chungnam National University, Daejeon 34134, Korea



Received: Apr 4, 2024
Revised: Apr 20, 2024
Accepted: Apr 30, 2024

[#]These authors contributed equally to this work.

*Corresponding author

Jung Min Heo
Department of Animal Science and Biotechnology, Chungnam National University, Daejeon 34134, Korea.
Tel: +82-42-821-5777
E-mail: jmheo@cnu.ac.kr

Copyright © 2024 Korean Society of Animal Sciences and Technology. This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

ORCID

Myunghwan Yu
<https://orcid.org/0000-0003-4479-4677>
Nuwan Chamara Chathuranga
<https://orcid.org/0000-0003-1002-4068>
Elijah Ogola Oketch
<https://orcid.org/0000-0003-4364-460X>
Jun Seon Hong
<https://orcid.org/0000-0003-2142-9888>
Haeun Park
<https://orcid.org/0000-0003-3244-0716>
Jung Min Heo
<https://orcid.org/0000-0002-3693-1320>

Abstract

The current study investigated the impact of using iron and plastic crates during summer transportation on production, physiological characteristics, and welfare of broiler chickens. A total of 160 Ross 308 male broilers were randomly selected from a battery-caged house at 35 days of age. Their average body weight was $1,866.62 \pm 36.048$ g (mean \pm SEM). Broilers were crated into fixed iron crates with 1.00 m (length) \times 0.78 m (width) \times 0.26 m (height) and plastic crates with 0.82 m (length) \times 0.57 m (width) \times 0.29 m (height) dimensions at 173 cm²/kg densities. Afterward, they were transported in the early morning at an average speed of 30–50 km/h for 40 minutes under 30°C and 40% relative humidity, completing a total distance of 20 km. Body weights were recorded before and after completing the journey. Following the weighing of birds, blood samples were collected for blood metabolite (cortisol, glucose, and lactate) analysis. Cervical dislocation was performed to euthanize broilers followed by breast and drumstick collection. Dressing, drumstick, and breast meat were calculated as percentages whereas respiratory frequencies were measured as the number of breaths per minute. Collected breast meat samples were utilized to analyze physiochemical parameters such as pH, color (CIE L*, a*, b*), water holding capacity, and cooking loss. Results from skin temperature assessments showed higher temperatures ($p < 0.05$) in broilers that were loaded into iron crates, both before (iron, $41.23 \pm 0.610^\circ\text{C}$; plastic, $39.25 \pm 0.065^\circ\text{C}$) and after (iron, $43.53 \pm 0.723^\circ\text{C}$, and plastic, $41.63 \pm 0.132^\circ\text{C}$) completing the journey. However, total skin temperature change was not significantly affected. Importantly, stress-indicating blood metabolite analysis revealed that glucose and lactate levels were lower ($p < 0.05$) in broilers transported in plastic crates. Nevertheless, cortisol levels remained unaffected by crate materials. Furthermore, transit losses, carcass characteristics, and physiochemical properties were also unaffected despite the dissimilar crate types. In conclusion, the study revealed that plastic is the more advantageous crating material compared to iron. Besides, plastic crates ensure meat quality and animal welfare, as evidenced by blood metabolite levels and skin temperature after transit.

Keywords: Broiler, Crate, Meat quality, Stress, Transportation, Welfare

Competing interests

No potential conflict of interest relevant to this article was reported.

Funding sources

This research was carried out with the support of “Cooperative Research Program for Agriculture Science and Technology Development (Project No. RS-2021-RD010100 [PJ016214])” Rural Development Administration, Korea.

Acknowledgements

Not applicable.

Availability of data and material

Upon a reasonable request, the datasets of this study can be available from the corresponding author.

Authors' contributions

Conceptualization: Yu M, Heo JM.

Data curation: Yu M.

Formal analysis: Yu M, Chathuranga NC.

Methodology: Yu M.

Software: Yu M, Chathuranga NC.

Validation: Yu M, Heo JM.

Investigation: Yu M, Chathuranga NC, Oketch EO, Hong JS, Park H.

Writing - original draft: Yu M, Chathuranga NC.

Writing - review & editing: Yu M, Chathuranga NC, Oketch EO, Hong JS, Park H, Heo JM.

Ethics approval and consent to participate

This study was approved by the Animal Ethics Committee of Chungnam National University, Daejeon, Korea (approval number: 202206A-CNU-081).

INTRODUCTION

The global poultry meat industry, representing approximately 40% of total meat production heavily relies on transportation. Notably, chickens have constituted a staggering 87% of poultry that transit upon reaching market weight [1]. However, concerns have arisen regarding broiler transportation and its impact on production, meat quality, food safety, and animal welfare [2]. This process involves pre-transit, transit, and post-transit phases, during which broilers encounter various stressors disrupting their physiological equilibrium. These stressors include health conditions, microclimate, crating densities, trailer design, vibration, noise, and feed and water withdrawal periods [3–5]. Such stressors can compromise productivity and meat quality [6], with seasonal changes further exacerbating challenges, particularly in coping with temperature fluctuations [7,8].

Notwithstanding, producers adopt different strategies to mitigate economic and quality losses during different phases of broiler transit [9], including temperature regulation in microclimates [10], optimizing journey length [7], crating densities [11], ventilation [12], and managing feed and water withdrawal periods [11] as crucial factors. However, these strategies must prioritize animal welfare, encompassing physical and mental states, emotional and cognitive aspects, avoidance behavior, biochemistry, and responses to stimuli [13,14]. Stressful environments can significantly impact broiler productivity and health by activating the hypothalamo-pituitary-adrenocortical cascade and triggering hormonal secretions [15,16]. Hence, plasma or serum corticosteroid levels serve as acute stress indicators when proper blood drawing methods are followed [17].

Stress hormones such as corticosterone and cortisol trigger rapid energy production in animals [18], increasing the demand for glucose and elevating blood glucose levels [19]. In situations where oxygen supply is inadequate for aerobic metabolism, muscles undergo anaerobic respiration, converting glucose into lactate. This accumulation of lactate in the bloodstream indicates stress in birds [20]. Stressors also elevate plasma corticosteroid levels due to heightened secretion of adrenocorticotrophic hormone and corticotropin-releasing hormone. However, cortical cell function and numbers may also be altered by dietary protein restrictions as well [21].

The World Organization for Animal Health acknowledges the crucial role of maintaining appropriate animal welfare measures during transportation and recognizing its significant impact on meat quality [2]. European Council regulation (EC No 1/2005) complements this stance by aiming to prevent animal suffering and promote welfare regulations for European operators. This regulation covers various aspects, including transport preparation, responsibilities, competence, equipment standards, and documentation requirements [22]. Additionally, countries such as the USA, Australia, and the Republic of Korea have implemented regulations prioritizing crate design and construction within the respective regions [23,24]. However, Duval et al. [25] identified four major risk factors associated with animal transportation across different jurisdictions: climatic conditions, animal fitness, stocking densities, and transit durations.

The selection of crate materials for broiler transportation is diverse globally due to factors such as operational scale, cost, technology, durability, and environmental impact. Recently, plastic crates have been among commonly used, while balancing convenience with ecological considerations. However, researchers have traditionally overlooked crate materials as a major concern. Emerging factors like environmental impact, hygiene, and local regulations should now shape these decisions. Thus, the current study aims to explore the effects of different crate materials on broiler production, stress levels, meat quality, and welfare, providing insights to improve transportation practices, especially during the summer season.

MATERIALS AND METHODS

The Animal Ethics Committee of Chungnam National University, Daejeon, Korea, approved the protocols used in this experiment (approval number: 202206-CNU-081).

Birds, experimental design, and treatments

Before transportation, all birds were housed in Chungnam National University's experiment farm which had 48 battery cages ($76 \times 61 \times 46 \text{ cm}^3$) that housed six birds until transportation and were managed according to the Ross 308 broiler management guideline [26]. A total of 160 Ross 308 male broilers were used at 35 days of age with an average body weight of $1,866.62 \pm 36.048 \text{ g}$ (mean \pm SEM). They were randomly selected after 4 h of feed withdrawal before catching. Afterward, birds were taken out from the cages and transported securely by holding their wings against the handler's body using both hands (Japanese method) [27]. The birds were transported in different types of crates as follows: an iron crate having dimensions of 1.00 m (length) \times 0.78 m (width) \times 0.26 m (height); a plastic crate having dimensions of 0.82 m (length) \times 0.57 m (width) \times 0.29 m (height) with 4 replicates per crate type. The birds were placed in crates based on optimal crating density suggested by the Korean transportation standards [28] of an average of $173 \text{ cm}^2/\text{kg}$. The transportation's distance was 20 km for 40 min at an average speed of 30–50 km/h during the early morning from 8:00 a.m. The weather was bright and sunny, the average temperature was 30°C , and the relative humidity was 40%. Temperature and humidity data were obtained from the Meteorological Agency records.

Transportation losses

Body weight loss (g) in transit was measured as the difference between all broilers' weight before transportation and the final body weight (g) from all crates upon arrival at the destination after transportation [29].

Carcass traits and sample collection

Once birds arrived at the destination, carcass trait measurements and sample collection were taken place. Two birds were selected based on closeness to the mean body weight of the birds in the respective crate, and the resulting weight was recorded as the live body weight. Blood samples were collected from the brachial vein into a vacutainer coated with lithium heparin (BD Vacutainer, BD, Franklin Lakes, NJ, USA) before euthanizing the birds. The birds were then euthanized by cervical dislocation for the evaluation of some carcass characteristics. The dressing percentage with giblets (heart, gizzard, and liver) was determined as a function of the live weight of the birds. The breast meat was then separated and weighed to measure its relative to the total carcass weight. The breast meat of broilers was then collected for meat quality analyses [29].

Physiological responses

Collected blood samples were centrifuged (LABOGENE 1248R, Gyrozen, Daejeon, Korea) at $3,000\times g$ for 10 min at 4°C and the plasma was separated and stored at -80°C (UniFreez U 400, DAIHAN Scientific, Wonju, Korea) until analysis. Cortisol concentrations were determined from the plasma with a cortisol ELISA kit (CUSABIO, Wuhan Huamei lotech, Wuhan, China) used in accordance with the manufacturer's instructions. Lactate concentration was determined by lactate assay kit (Sigma Aldrich, Burlington, NJ, USA) using the manufacturer's instructions. Briefly, glucose was determined from the collected plasma using a glucose assay kit (Asan Pharmaceutical, Seoul, Korea), following the manufacturer's instructions.

After finishing transportation, the respiratory frequency was measured as the number of breaths per minute using three randomly selected broilers per crate observed by the camera (GoPro Hero 8, GoPro, San Mateo, CA, USA) for 1 minute [30].

Physicochemical traits

The pH values of the breast meat were monitored immediately after sample collection. An aliquot (9 mL) of distilled water was added to 1 g of muscle, followed by homogenization (T25 basis, IKA-Werke GmbH & Co. KG, Staufen, Germany) for 30 seconds. The homogenate was centrifuged at $2,090 \times g$ (ScanSpeed 1580R, Labogene ApS, Lillerød, Denmark) for 10 min and the supernatant was filtered through filter paper (No. 4, Whatman, Maidstone, UK). The pH of the filtrate was measured using a pH meter (SevenEasy, Mettler-Toledo International, Schwerzenbach, Switzerland).

The CIE (Commission Internationale de l'Eclairage) L^* , a^* , and b^* of broiler breast meat were determined using a spectrophotometer (CM-3500d, Minolta, Tokyo, Japan). Measurements were taken perpendicularly to the surface of the broiler breast meat with a 30 mm diameter illumination area at two different locations per sample. The results were analyzed in the SpectraMagic software (Spectramagic™ NX, Konica Minolta, Tokyo, Japan).

For the water holding capacity (WHC) measurements, a 2 g sample of raw broiler breast meat was precisely weighed, placed on cotton wool, and then added to a centrifuge tube. The weight of the meat after centrifugation at $2,090 \times g$ (ScanSpeed 1580R, Labogene ApS) for 10 min was measured and compared to the initial meat weight. The moisture content of meat was determined by drying 2 g of samples placed in aluminum dishes for 3 h at 110°C . The remaining moisture (%) present in the meat after centrifugation was expressed as the WHC [31].

To measure the cooking loss, the breast meat of the broiler was weighed vacuum packaged and cooked for 20 min in a water bath at 80°C until the internal temperature reached 70°C . The cooked breast meat of broilers was cooled at room temperature (20°C) for 30 min. After removal of the vacuum bag, the surface moisture of the breast meat of the broiler was removed with paper towels, and the cooked breast meat of the broiler was weighed. The cooking loss was calculated as the difference between the weight of raw breast meat and cooked breast meat.

Skin temperature measurements

The body surface temperature of broilers within individual broiler cages was assessed utilizing a portable thermal imaging camera (Infrared, IRay T3PRO, Shandong, China) both before and after transportation. The measurements were then averaged to derive a representative value for each cage. Furthermore, temperature differentials pre- and post-arrival were quantified by converting them into absolute values, facilitating rigorous analysis of the thermal dynamics experienced by the broilers during transport [32].

Statistical analysis

The statistical analysis of the data was performed using SPSS (Version 26, IBM, Armonk, NY, USA). The data obtained from the experiment except for the skin temperature were analyzed by independent sample t-test. The data of skin temperature were analyzed by a two-way ANOVA model to evaluate the main effects (types of crate and transportation) followed by Tukey's multiple range analysis with each crate as the experimental unit. In terms of transportation loss and respiratory frequency measurements, the experimental unit was defined as the crate. For carcass traits, meat quality, and blood metabolites, selected individual birds were considered as the experimental unit. Statistical significance was determined at a significance level of $p < 0.05$.

RESULTS

The broilers used in the experiment were transported according to the appropriate transport density specified in Korean Law [28], so the mortality did not occur regardless of the two crate material types.

Transportation losses and carcass traits

The results of the transportation losses and carcass traits of the broiler using different types of crates during transportation are shown in Table 1. There was no significant difference ($p > 0.05$) in body weight loss and carcass traits such as dressing ratios, relative breast meat, and drumstick weights between the treatments.

Physiological responses

The results of broiler blood metabolites, after transporting with different crate types are shown in Fig. 1. Broilers transported in plastic crates had reduced glucose and lactate contents ($p < 0.05$) compared with those in iron crates. However, plasma cortisol levels remained unaffected ($p > 0.05$) despite the different types of crates used during transportation. The impact of different

Table 1. Live weight loss and carcass traits as affected by different materials of crates during transportation in summer.

Items	Iron	Plastic	SEM	p-value
Body weight loss (g)	32.15	26.46	1.737	0.102
Dressing ratio ¹ (%)	89.58	89.72	0.238	0.788
Relative breast meat weight ² (%)	28.03	28.23	0.604	0.878
Relative drumstick weight ³ (%)	10.53	10.41	0.155	0.713

¹(Carcass weight/Live body weight) × 100.

²(Breast meat weight/carcass weight) × 100.

³(Drumstick weight/carcass weight) × 100.

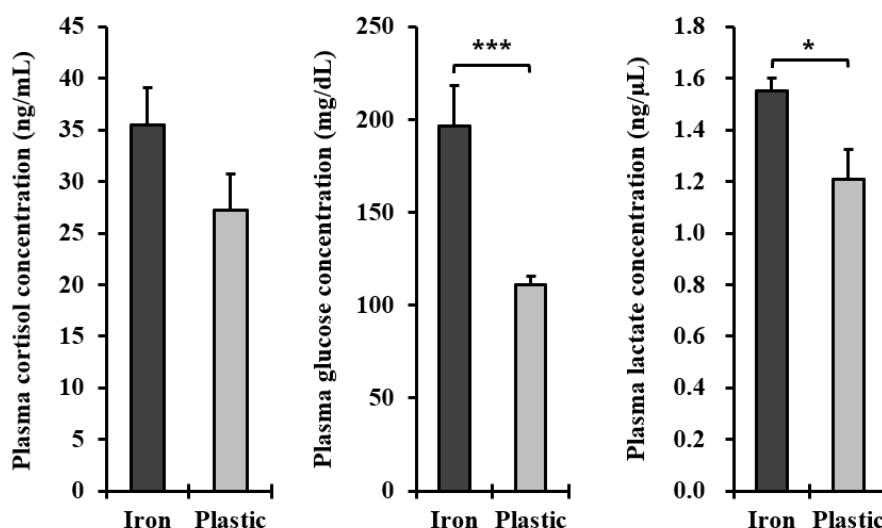


Fig. 1. Blood metabolites of transported broilers under different materials of crates during transportation in summer. The values in the histogram are the means ± SEM (n = 8). Significant difference of * $p < 0.05$ and *** $p < 0.01$.

crate materials on broiler chicken respiration after transportation is shown in Fig. 2. There was no significant difference ($p > 0.05$) in respiratory rate between the two treatments.

Physicochemical traits

The results of the physicochemical characteristics of broilers related to different crate types during transportation are shown in Table 2. The meat quality parameters such as pH, WHC, cooking loss, and meat color (L^* , a^* , b^*) were not significantly different ($p > 0.05$) related to the two treatments.

Skin temperature evaluation

The skin temperature of transported broilers in crates made of different materials during transportation in summer is presented in Table 3. The interaction between transportation and crate type did not show any significant effect in the current study ($p > 0.05$). However, transportation did affect the increase in total shipment temperature after transportation ($p < 0.001$), while crate type demonstrated higher ($p < 0.01$) temperatures in fixed iron crates compared to plastic crates.

DISCUSSION

The current study intended to evaluate the impact of different crating materials used during

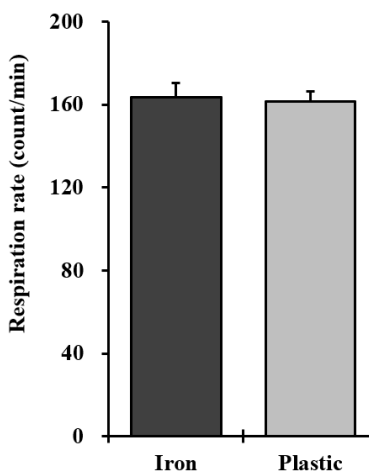


Fig. 2. Respiration rate of transported broilers under different materials of crates during transportation in summer. The values in the histogram are the means \pm SEM ($n = 4$).

Table 2. Physicochemical traits of chicken breast meat under different materials of crates during transportation in summer

Items	Iron	Plastic	SEM	<i>p</i> -value
pH	5.92	5.96	0.035	0.588
WHC (%)	63.13	67.22	2.019	0.335
Cooking loss (%)	23.13	21.42	0.966	0.402
L^*	53.45	52.85	0.411	0.476
a^*	6.74	7.20	0.287	0.440
b^*	16.34	17.19	0.250	0.095

WHC, water holding capacity.

Table 3. Skin temperature of broilers under different materials of crates and transportation in summer

Transportation	Type of crate	Skin temperature (°C)
X	Iron	41.23
	Plastic	39.25
O	Iron	43.53
	Plastic	41.63
SEM		0.239
Main effect		
Transportation		
X		40.24
O		42.58
SEM		0.341
Type of crate		
Iron		42.38
Plastic		40.44
SEM		0.383
<i>p</i> -value		
Transportation		< 0.001
Type of crate		0.002
Transportation × type of crate		0.939

transportation on the production, physiological characteristics, and welfare of broilers in the summer season. The broilers used in the current study were transported in compliance with the transport density regulations stipulated by Korean Law [28], which could result in non-mortality during transportation.

Iron possesses high heat conducting and heat transferring properties as a structural material. These characteristics enhance the heat absorption from the outer environment and convey it from highly heated areas to low-heated areas through conductivity. Ultimately, this process increases the total surface temperatures in particular areas [33,34]. In comparison, plastic has excellent insulating capabilities with inherited heat resistance properties which can be used as a structural material in many industries of the world [35]. In our study, crates made of iron have been observed to elevate the microenvironment temperature, as evidenced by the rise in the skin temperatures of broilers. Conversely, broilers housed in plastic crates, under similar conditions, exhibited lower skin temperatures. Since body surface temperature serves as a common indicator of thermal comfort or stress in broiler chickens [36,37], it is possible that iron crates contribute to increased temperatures within the crate space due to radiation reflection and high thermal absorptivity. In contrast, plastic crates exhibit lower temperatures, due to their better thermal insulation properties against heat.

Typically, the plasma glucose levels in an avian species are 150% to 200% greater than those found in mammals with comparable body mass [38]. Hence, its concentration directly affects the metabolism of poultry. According to Liu and Zhao [39], there is a significant impact on glucose metabolism in chickens related to their stress regulations. Particularly, glucose and triglycerides are recognized as major metabolites that play crucial roles in biochemical and physiological functions, providing the energy needed to fuel these processes [40]. This glucose concentration can vary based on several factors in poultry including, physiological status, age, gender, energy intake, and ambient temperature [39]. In relevance, the glucose level increment in broilers transported in iron crates could be due to the stress, caused by the type of crating material and macroenvironment (in

summer) temperatures.

While transportation, broilers experience acute stress conditions (such as bruising or broken wings) from hard surfaces like iron crates and they could lead to increased gluconeogenesis and glucose mobilization within the animal body. Additionally, this sudden introduction of broilers into an excessive heat and humidity environment (in summer) from a relatively constant environment can exhibit a detrimental effect on them [27]. Consequently, elevated microclimate temperatures can further exacerbate these physiological processes in broilers, potentially resulting in augmented glucose levels in broiler plasma [41]. Similarly, lactate level increment in metal-crated broilers could also be due to the aforementioned stress factors similarly glucose level fluctuations.

The current results on broiler transport and welfare, emphasize the importance of further research into the effects of crate materials on broiler health and the economic and practical benefits for the industry. In conclusion, the study underscores the importance of crate material choice in broiler transportation while revealing the plastic crate as the advantageous crate material type compared to the iron that can ensure meat quality and animal welfare in terms of blood metabolite changes and skin temperature changes after loading.

REFERENCES

1. Weeks CA. Poultry handling and transport. In: Grandin T, editors. *Livestock handling and transport*. 4th ed. Wallingford: CAB International; 2014. p. 378-98.
2. Schwartzkopf-Genswein KS, Faucitano L, Dadgar S, Shand P, González LA, Crowe TG. Road transport of cattle, swine and poultry in North America and its impact on animal welfare, carcass and meat quality: a review. *Meat Sci.* 2012;92:227-43. <https://doi.org/10.1016/j.meatsci.2012.04.010>
3. Denadai JC, Mendes AA, Garcia RG, Almeida ICL, Moreira J, Takita TS, et al. Effect of feed and water withdrawal on carcass yield and breast meat quality of broilers. *Rev Bras Cienc Avic.* 2002;4:101-9. <https://doi.org/10.1590/S1516-635X2002000200002>
4. Nicol CJ, Scott GB. Pre-slaughter handling and transport of broiler chickens. *Appl Anim Behav Sci.* 1990;28:57-73. [https://doi.org/10.1016/0168-1591\(90\)90046-G](https://doi.org/10.1016/0168-1591(90)90046-G)
5. Saraiva S, Esteves A, Oliveira I, Mitchell M, Stilwell G. Impact of pre-slaughter factors on welfare of broilers. *Vet Anim Sci.* 2020;10:100146. <https://doi.org/10.1016/j.vas.2020.100146>
6. Mitchell MA, Kettlewell PJ. Physiological stress and welfare of broiler chickens in transit: solutions not problems! *Poult Sci.* 1998;77:1803-14. <https://doi.org/10.1093/ps/77.12.1803>
7. Dos Santos VM, Dallago BSL, Racanicci AMC, Santana ÂP, Cue RI, Bernal FEM. Effect of transportation distances, seasons and crate microclimate on broiler chicken production losses. *PLOS ONE.* 2020;15:e0232004. <https://doi.org/10.1371/journal.pone.0232004>
8. Kpomasse CC, Oke OE, Houndonougbo FM, Tona K. Broiler production challenges in the tropics: a review. *Vet Med Sci.* 2021;7:831-42. <https://doi.org/10.1002/vms3.435>
9. Kettlewell PJ, Mitchell MA. Catching, handling and loading of poultry for road transportation. *Worlds Poult Sci J.* 1994;50:54-6. <https://doi.org/10.1079/WPS19940005>
10. Mitchell MA, Kettlewell PJ. Welfare of poultry during transport - a review. In: *Proceedings of the 8th European Symposium on Poultry Welfare*; 2009; Cervia, Italy. p. 90-100.
11. Yu M, Oketch EO, Hong JS, Nawarathne SR, Vohobjonov Y, Heo JM. Evaluation of preslaughter losses, meat quality, and physiological characteristics of broilers in response to crating density for the standard of animal welfare and to seasonal differences. *Korean J Agric Sci.* 2022;49:927-36. <https://doi.org/10.7744/kjoas.2022084>
12. Aldridge DJ, Luthra K, Liang Y, Christensen K, Watkins SE, Scanes CG. Thermal micro-

- environment during poultry transportation in South Central United States. *Animals*. 2019;9:31. <https://doi.org/10.3390/ani9010031>
13. OIE [World Organization for Animal Health]. Introduction to the recommendations for animal welfare. In: *Terrestrial Animal Health Code*. 28th ed. Paris: OIE; 2019. p.333-5.
 14. Kumar P, Ahmed MA, Abubakar AA, Hayat MN, Kaka U, Ajat M, et al. Improving animal welfare status and meat quality through assessment of stress biomarkers: a critical review. *Meat Sci*. 2023;197:109048. <https://doi.org/10.1016/j.meatsci.2022.109048>
 15. Scanes CG, Dridi S. *Sturkie's avian physiology*. 7th ed. Amsterdam: Academic Press; 2021.
 16. Blas J. Stress in birds. In: Scanes CG, editor. *Sturkie's avian physiology*. 6th ed. Amsterdam: Academic Press; 2015. p. 769-810.
 17. Weimer SL, Wideman RF, Scanes CG, Mauromoustakos A, Christensen KD, Vizzier-Thaxton Y. An evaluation of methods for measuring stress in broiler chickens. *Poult Sci*. 2018;97:3381-9. <https://doi.org/10.3382/ps/pey204>
 18. Ghassemi Nejad J, Ghaffari MH, Ataollahi M, Jo JH, Lee HG. Stress concepts and applications in various matrices with a focus on hair cortisol and analytical methods. *Animals*. 2022;12:3096. <https://doi.org/10.3390/ani12223096>
 19. Sapolsky RM, Romero LM, Munck AU. How do glucocorticoids influence stress responses? Integrating permissive, suppressive, stimulatory, and preparative actions. *Endocr Rev*. 2000;21:55-89. <https://doi.org/10.1210/edrv.21.1.0389>
 20. Scanes CG. Hormones and metabolism in poultry. In: Aimaretti G, Marzullo P, Prodham F, editors. *Update on mechanisms of hormone action: focus on metabolism, growth and reproduction*. Rijeka: InTech; 2011. p.111-32.
 21. Scanes CG. Biology of stress in poultry with emphasis on glucocorticoids and the heterophil to lymphocyte ratio. *Poult Sci*. 2016;95:2208-15. <https://doi.org/10.3382/ps/pew137>
 22. Warin L, Mindus C, Sossidou E, Spoolder H, Bignon L. European guide of good practices for poultry transport. In: *13èmes Journées de la Recherche Avicole et Palmipèdes à Foie Gras*; 2019; Tours. p. 811-4.
 23. Delezie E, Swennen Q, Buyse J, Decuypere E. The effect of feed withdrawal and crating density in transit on metabolism and meat quality of broilers at slaughter weight. *Poult Sci*. 2007;86:1414-23. <https://doi.org/10.1093/ps/86.7.1414>
 24. Hussnain F, Mahmud A, Mehmood S, Jaspal MH. Influence of long-distance transportation under various crating densities on broiler meat quality during hot and humid weather. *J Poult Sci*. 2020;57:246-52. <https://doi.org/10.2141/jpsa.0190087>
 25. Duval E, Lecorps B, von Keyserlingk MAG. Are regulations addressing farm animal welfare issues during live transportation fit for purpose? A multi-country jurisdictional check. *R Soc Open Sci*. 2024;11:231072. <https://doi.org/10.1098/rsos.231072>
 26. Aviagen. *Ross broiler management handbook: preface* [Internet]. Aviagen. 2018 [cited 2024 Apr 4]. https://en.aviagen.com/assets/Tech_Center/Ross_Broiler/Ross-Broiler_Handbook2018-EN.pdf
 27. Dos Santos VM, Dallago BSL, Racanicci AMC, Santana ÂP, Bernal FEM. Effects of season and distance during transport on broiler chicken meat. *Poult Sci*. 2017;96:4270-9. <https://doi.org/10.3382/ps/pex282>
 28. Animal and Plant Quarantine Agency. *Animal Transport Detailed Regulations*, Notice No. 2013-3 (Mar. 5, 2013).
 29. Yu M, Jeon JO, Cho HM, Hong JS, Kim YB, Nawarathne SR, et al. Broiler responses to dietary 3,4,5-trihydroxybenzoic acid and oregano extracts under *Eimeria* challenge conditions. *J Anim Sci Technol*. 2021;63:1362-75. <https://doi.org/10.5187/jast.2021.e121>

30. Yu M, Oketch EO, Chaturanga NC, Nawarathne SR, Hong JS, Maniraguha V, et al. Effect of crating density and weather conditions during transit on preslaughter losses, physiological characteristics, and meat quality in broilers. *J Anim Sci Technol*. Forthcoming 2023. <https://doi.org/10.5187/jast.2023.e132>
31. Jeong HG, Jo K, Lee S, Yong HI, Choi YS, Jung S. Characteristics of pork emulsion gel manufactured with hot-boned pork and winter mushroom powder without phosphate. *Meat Sci*. 2023;197:109070. <https://doi.org/10.1016/j.meatsci.2022.109070>
32. Lee J, Yu M, Nawarathne SR, Oketch EO, Heo JM. Effect of crating density and weather in transit on behavior, surface temperature, and respiration rate in broilers considering animal welfare. *Korean J Poult Sci*. 2023;50:293-301. <https://doi.org/10.5536/KJPS.2023.50.4.293>
33. ElJersifi A, Aouadi K, Ben Ali M, Chbihi A, Semlal N, Bouaouine H, et al. The effect of heat treatments on the properties of a ferritic high-chromium cast iron. *J Mater Eng Perform*. 2023;32:8262-73. <https://doi.org/10.1007/s11665-022-07700-9>
34. Wasserman E, Stixrude L, Cohen RE. Thermal properties of iron at high pressures and temperatures. *Phys Rev B Condens Matter*. 1996;53:8296-309. <https://doi.org/10.1103/PhysRevB.53.8296>
35. Agarwal S, Gupta RK. Plastics in buildings and construction. In: Kutz M, editor. *Applied plastics engineering handbook: processing, materials, and applications*. William Andrew: Oxford; 2017. p. 635-49.
36. Cangar Ö, Aerts JM, Buyse J, Berckmans D. Quantification of the spatial distribution of surface temperatures of broilers. *Poult Sci*. 2008;87:2493-9. <https://doi.org/10.3382/ps.2007-00326>
37. Moghbeli Damane M, Barazandeh A, Sattaei Mokhtari M, Esmailipour O, Badakhshan Y. Evaluation of body surface temperature in broiler chickens during the rearing period based on age, air temperature and feather condition. *Iran J Appl Anim Sci*. 2018;8:499-504.
38. Hu X, Liu X, Guo Y, Li Y, Cao Z, Zhang Y, et al. Effects of chicken serum metabolite treatment on the blood glucose control and inflammatory response in streptozotocin-induced type 2 diabetes mellitus rats. *Int J Mol Sci*. 2023;24:523. <https://doi.org/10.3390/ijms24010523>
39. Liu W, Zhao J. Insights into the molecular mechanism of glucose metabolism regulation under stress in chicken skeletal muscle tissues. *Saudi J Biol Sci*. 2014;21:197-203. <https://doi.org/10.1016/j.sjbs.2014.01.005>
40. Hernawan E, Wahyuni S, Suprpti H. The levels of blood glucose, triglyceride, final body weight and abdominal fat percentage of broiler under sex-separated and straight run rearing system. *Lucrări Științifice-Seria Zootehnie*. 2012;57:28-33.
41. Virden WS, Kidd MT. Physiological stress in broilers: ramifications on nutrient digestibility and responses. *J Appl Poult Res*. 2009;18:338-47. <https://doi.org/10.3382/japr.2007-00093>