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ARTICLE INFORMATION	Fill in information in each box below
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Article Title (within 20 words without abbreviations)	Increasing arginine supplementation alleviated heat stress and citrulline can effectively substitute arginine in broilers
Running Title (within 10 words)	Optimal dietary arginine ratio and citrulline substitution
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9 Abstract

10 This study was conducted to determine the optimal standard ileal digestible (SID) arginine (Arg) to 11 SID lysine (Lys) ratio in broilers under cyclic heat stress. Additionally this study tested whether 12 citrulline (Cit) can replace Arg under cyclic heat stress, based on the report that a large amount of Arg 13 is metabolized in the liver while Cit can by-pass metabolism in the liver. A total of 360, one-day-old 14 Arbor Acres broiler chickens with initial body weight of 34.50 ± 0.87 g were placed in 24 pens. The 15 24 pens were randomly assigned to four dietary treatments with six replicates of fifteen broiler 16 chickens. Treatments were as follows: 1) NC (SID Arg : Lys =0.95), 2) PC (SID Arg : Lys=1.05), 3) 17 Arg1.15 (SID Arg : Lys =1.15), 4) Arg1.25 (SID Arg : Lys =1.25), 5) Cit33 (supplementation of Cit 18 at 33% of Arg supplementation in Arg1.15, 6) Cit50 (supplementation of Cit at 50% of Arg in 19 Arg1.15). The Arg1.25 group had the highest BW on 32 days and BWG during the overall period 20 (p<0.05) than the NC groups. However, there was no significant difference (p>0.05) on day 32 BW 21 and BWG during the overall period in Arg supplemented groups (Arg1.15 and Arg1.25) and Arg replacement with Cit groups (Cit33 and Cit50). Arg1,25 and Cit33 groups had higher villus height 22 23 (VH) in the duodenum, jejunum and ileum than the NC groups. Moreover, the Arg1.25 group had the lowest crypt depth (CD) in the jejunum and ileum than the NC group, while there was no significant 24 25 difference (p>0.05) between Arg supplementation and Arg replacement with Cit groups. Arg1.25 26 group had the highest arginase activity in the liver and total nitric oxide synthase (NOS) and arginase 27 activity in the kidney than other treatments, but no statistical difference was observed (p>0.05) in 28 arginase in the liver among treatments. Collectively the results ascertain that Cit can effectively 29 replace a certain part of dietary arginine in broiler diets.

30

31 Keywords (3 to 6): Arginine, Broiler chickens, Citrulline, Growth performance, Heat stress

32

33

35 Introduction

36 Broilers are susceptible to high temperatures since they have feathers on the body, the absence of 37 sweat glands, and a high metabolic rate, and thus heat stress (HS) significantly affects well-being and 38 production [1]. Actually, broilers exposed to HS exhibit reduced growth performance, lower nutrient 39 digestibility, increased production of free radicals in the body, and higher mortality rates compared to 40 those rared in thermoneutral temperatures [2-3]. Morevoer, climate change have been increasing and 41 globally is contributing to a rise in HS in poultry production, which could increase impaired 42 production of broilers around the world [4]. Certain functional AAs can influence metabolic pathways, 43 thereby promoting overall health, including growth and survival in animals [5].

44 Arginine (Arg) among them is the fifth limiting AA in broiler chickens, and a precursor of Citrulline 45 (Cit), ornithine, and nitric oxide (NO) [6]. Arg plays a role in immune and metabolic pathways, and supplementing the diet with Arg has been shown to improve feed efficiency, growth performance, and 46 47 the immune system [7]. Arg has also been given attention in relation to HS. NO is an active effector 48 of vasodilation [6]. Increased NO production can decrease body temperature by improved blood flow and oxidative stress by elevating the activity of superoxide dismutase (SOD) [8-10]. Ornithine, along 49 with its downstream metabolites such as polyamines, has been shown to be critical in the process of 50 51 tissue recovery [11-12]. These mechanisms could help to minimize the negative impacts of HS. 52 Various conditions such as diets and environments considerably affect arginine requirement of 53 broilers [13]. Moreover, the dietary Arg requirements for modern broilers need to be optimized due to 54 superior genetic potential for protein deposition and the antagonism between Arg and lysine (Lys) 55 [14].

56 Cit and Arg are both metabolic intermediates in the urea cycle. Cit is a non-protein AA and acts as a 57 catalyst in the formation of Arg, which in turn leads to the production of NO [15-17]. It is important 58 to note that Cit can be recycled to Arg and have Arg sparing effects [18]. Actually, previous studies 59 have reported that dietary Cit supplementation to Arg-deficient feed improved gut health by 60 increasing the circulation rate of Arg and NO in tissues [19-20]. Likewise, in recent studies related to 61 mono-gastric animals, dietary Cit supplementation decreased pre-weaning mortality rate of piglets, 62 and increased intestinal morphology villus height (VH) and improved feed efficiency of broilers under HS condition (about increased 8°C and 9°C compared to normal condition) [17, 21]. Chowdhury et al. 63

64 [22] additionally reported that supplementing Cit lowered body temperature in HS conditions (about 65 increased 5°C compared to normal condition). Therefore, the objective of this study was to determine 66 the optimum requirement of Arg and Cit for broilers under cyclic HS. The hypotheses tested in this 67 experiment were (1) optimum SID Arg:Lys ratio for broilers will be greater than the current 68 recommendations of 105% under cyclic HS; (2) Cit can replace Arg when supplemented in Arg 69 deficient broiler diet; and (3) Cit inclusion at 33% or 50% of Arg will be sufficient to maintain 70 optimum growth of broilers, as Cit is not metabolized in the liver as Arg does.

71

72 Materials and Methods

73 Experimental animal and design

74 A total of 360, one-day-old Arbor Acres broiler chickens (Cherrybro Co., Eumseong, Korea) with an 75 initial body weight of 34.50 ± 0.87 g were placed in 24 pens (173 cm width, 63 cm depth and 55 cm 76 height). The 24 pens were randomly assigned to four dietary treatments with six replicates of fifteen broiler chickens. Treatments were as follows: 1) NC (SID Arg : Lys =0.95), 2) PC (SID Arg : 77 Lys=1.05), 3) Arg1.15 (SID Arg : Lys =1.15), 4) Arg1.25 (SID Arg : Lys =1.25), 5) Cit33 78 79 (supplementation of Cit at 33% of Arg in Arg1.15), Cit50 (supplementation of Cit at 50% of Arg in Arg1.15). All diets were formulated to meet or exceed the National Research Council [23] (Table 1 -80 81 4). All broiler chickens were allowed to consume feed and water ad libitum. Each pen was equipped 82 with two nipple drinkers connected to a common water supply line. The experiment period was 83 divided into three phases: the starter phase (0 to 7 days of age), the grower phase (8 to 21 days of age) 84 and the finishing phase (22 to 32 days of age). The lighting period was a continuous schedule with 85 lightning intensities of 50 lux from 0 to 7 d of age, and 20 lux from 8 to 32 d of age for broiler. The 86 experiment environment was controlled under at 33 ± 1 °C and 50 % relative humidity, and then 87 temperature was reduced by 2 °C every week until 24 °C on day 25. The broilers were challenged 88 with cyclic HS between 22 to 32 days of age. The room temperature was performed every day during 89 the same period (from 9:00 to 18:00 h) to ensure consistency in the design. The temperature was gradually increased from 24 ± 1 °C to 30 ± 1 °C over 30 min and this temperature was maintained for 90 91 the next 8 h before returning to 24 ± 1 °C.

93 Growth performance

94 All birds and leftover feed in the cages were weighed at each time point to determine the body weight 95 (BW), body weight gain (BWG), feed intake (FI), and feed conversion ratio (FCR) on 0, 7, 21 and 32 96 days. Mortality was recorded as it occurred. The BWG was calculated as the BW of the previous time 97 point was subtracted from the BW of the current time point. FI was calculated by subtracting the 98 remaining feed amount from the initial feed amount, and FCR was calculated by dividing FI by BWG. 99 Adjusted FCR at 1.5 kg BW was calculated as follows: FCR at 1.5 kg BW = FCR - (average BW-1.5)100 \times 0.3. Production index was calculated depending on the following formula: Production index = 101 {(Average body weight (kg) \times livability) / (Duration of the period (day) \times FCR)} \times 100.

102

103 Nutrient digestibility

104 Four broilers per pen were randomly selected to collect ileal digesta from Merkel's diverticulum to 105 the ileocecal junction). The digesta samples were pooled by pen in plastic bags and stored in a freeze 106 dryer for further analysis. Diets and freeze-dried ileal digesta were ground using a coffee grinder 107 before further analysis. A 0.2% chromium dioxide was included in the diets as an indigestible marker 108 and analyzed to estimate the apparent ileal digestibility (AID) of dry matter (DM), crude protein (CP), 109 gross energy (GE) and AA. The GE was determined using a calorimeter (model 6400, Parr Instrument 110 Company, Moline, IL, USA). DM and CP were analyzed according to the methods described in 111 AOAC Method 930.15 and Method 990.03 [24] and AA was analyzed using high performance liquid 112 chromatography (HPLC) (SHIMADZU, Model LC-10AT, Shimadzu Corp., Kyoto, Japan) followed 113 with AOAC Method 982.30E (a, b, c) [24]. In order to calculate AID, the concentrations of the maker, 114 and AA, DM and CP in diets and digesta were used, as shown below.

- 115 AID % = $100 [100 \times (Cr_2O_3 \text{ in diet}/ Cr_2O_3 \text{ in ileal digesta}) \times (nutrient in ileal digesta/nutrient in 116 diet)].$
- 117

118 Relative organ weight and carcass trait

Six broilers per each treatment were euthanized on 32 days of age by an intravenous injection of pentobarbital, with cervical dislocation to confirm death. After broilers were euthanized, the abdomen area was opened to excise and weigh the carcass, gastrointestinal tract (gizzard, stomach, duodenum,

- jejunum, and ileum), liver, spleen, and heart. Carcass yields were calculated relative to the live BW.
 The weights of the gastrointestinal tract and other organs were recorded and then calculated using the
- 124 following formula: Relative organ weight (g/kg) = organ weight (g)/live BW (kg).
- 125

126 Intestinal morphology

127 The intestinal segments ileum (midpoint from Meckel's diverticulum to the ileocecal junction), 128 jejunum (midpoint from the pancreatic duct to Meckel's diverticulum), and duodenum (the midsection 129 of the ascendant loop) were collected on 32 days and were rinsed with cold PBS and fixed with 4% 130 paraformaldehyde. Then, the tissues were embedded with paraffin and sectioned at a thickness of 3 to 131 5 µm. Next, the slides were stained with hematoxylin and eosin (H&E). Five slides were prepared for 132 each sample (from the central region of the sample), and images were captured using a light 133 microscope (OLYMPUS DP71, BX50F-3, Olympus Optical Co. Ltd., Tokyo, Japan). The distance 134 between the top of the villus to the villus-crypt junction was measured as VH, while the distance from the villus-crypt junction down to the bottom of the crypt was measured as CD. Three measurements 135 136 were taken per slide and the average was obtained for analysis. The VH to CD ratio (VCR) was 137 computed per observation.

138

139 **Blood profile**

Blood samples (2 mL each) were collected from the wing vein of broilers (one broiler per cage) using
vacuum tubes containing K₃EDTA (Becton, Dickinson and Co., Franklin Lakes, NJ, USA) at 32 days
of age. Concentrations of total protein (TP), blood urea nitrogen (BUN), and cortisol were determined
using an automatic biochemical analyzer (Hitachi, Tokyo, Japan). Concentration of NO
(MyBioSource: MBS263050), Cit (MyBioSource: MBS2601045) and Arg (Assay Genie: CHEB0588)
were determined using commercial ELISA kits (MyBioSource, San Diego, CA, USA; Assay Genie,
Dublin, Ireland).

147

148 **Enzyme activity**

149 Six broilers per treatment were selected to analyze the enzyme activity of Arg and NO in the liver and

150 kidney at 32 days of age. Enzymes involved in Arg metabolism including endothelial nitric oxide

151 synthase (Intron Biotechnology, Korea: 21023) and arginase synthase (MyBioSource: MBS746934)

152 were determined using chicken ELISA kits. The assay was measured at 450 nm using a microplate

153 reader (Elx808, Bio-Tek, Winooski, Vermont, USA) and the standard curve was used to compute the

154 sample concentration.

155

156 Statistical analysis

157 Growth performance, nutrient digestibility, blood profiles, intestinal morphology, relative organ 158 weight and carcass trait, and enzyme activity were statistically analyzed using JMP 16.0 (SAS 159 Institute Inc., Cary, NC). One-way ANOVA was conducted and Tukey HSD test was used to separate 160 the means. Variability in the data was expressed as the pooled standard error, P<0.05 was considered 161 statistically significant, and $0.05 \le P \le 0.10$ was considered statistically tendency.

162

163 **Results**

164 **Growth performance**

165 There was no significant difference (p>0.05) on BW on 7 and 21 days, and FI among treatments (Table 5). Arg deficiency (NC) caused (p < 0.05) a decrease in BW at 32 days of age, and BWG 166 167 during the overall period compared to other treatments. The Arg1.25 group had the highest BW at 32 days of age and BWG during the overall period (p < 0.05) than the NC group. However, there was no 168 169 significant difference (p>0.05) in BW at 32 days of age and BWG during the overall period in Arg-170 supplemented groups (Arg1.15 and Arg1.25) and Arg replacement with Cit groups (Cit33 and Cit50). 171 As for FCR and PI, there was no significant difference (p>0.05) in FCR in each period except for the 172 overall period among treatments. Arg deficiency increased (p < 0.05) FCR during the overall period 173 and FCR at 1.5kg, while Arg deficiency decreased (p < 0.05) PI compared to other treatments. The 174 ARG1.25 group had the lowest FCR during the overall period and FCR at 1.5 kg (p < 0.05) while Arg 175 1.25 had the highest PI (p < 0.05) compared to NC and PC groups. However, there was no significant 176 difference (p>0.05) in FCR during the overall period, FCR at 1.5 kg and PI in Arg supplemented 177 groups (Arg1.15 and Arg1.25) and Arg replacement with Cit groups (Cit33 and Cit50).

178

179 Nutrient digestibility

180 There was no significant difference (p > 0.05) in DM and GE digestibility among treatments (Table 181 6). Arg deficiency caused (p < 0.05) the reduction of CP digestibility compared to other treatments. 182 Direct Arg supplementation (Arg1.15 and Arg1.25) and Arg replacement with Cit (Cit33 and Cit50) 183 improved (p < 0.05) CP digestibility compared to the NC group, while there was no significant 184 difference among Arg and Cit supplemented groups. As for SID of AAs, there was no significant 185 difference (p > 0.05) except for Arg, Lys, Met and Cys among treatments (Table 7). Arg deficiency 186 decreased (p < 0.05) the digestibility of Arg, Lys, Met and Cys compared to other treatments. Direct 187 Arg supplementation (Arg1.15 and Arg1.25) and Arg replacement with Cit (Cit33 and Cit50) 188 improved (p < 0.05) the digestibility of Arg and Lys compared to the NC group.

189

190 **Blood profile**

191 There was no significant difference (p>0.05) in TP, BUN, cortisol and Arg concentrations in the 192 blood among treatments (Table 8). Arg deficiency decreased (p < 0.05) NO and Cit concentration in 193 the blood, while Arg supplementation and Arg replacement with Cit improved (p < 0.05) the 194 production of NO and Cit in the blood.

195

196 Intestinal morphology

As for duodenal morphology, Arg deficiency reduced (p < 0.05) VH and VCR compared to other treatments (Table 9). Furthermore, Arg deficiency induced (p < 0.05) poor morphology such as a decrease in VH and VCR, and an increase in CD of the jejunum and ileum compared to other treatments. Arg1.25 and Cit33 groups had higher VH and VCR in the duodenum, jejunum and ileum than the NC group. Moreover, Arg1.25 group had the lowest CD in the jejunum and ileum than the NC group, while there was no significant difference (p>0.05) between Arg supplementation and Arg replacement with Cit groups.

204

205 Organ weight

There was no significant difference in the relative weight of the gizzard, bursa of Fabricius and small intestine among treatments (Table 10). Although there was a significant difference (p < 0.05) in the

208	relative weight of the liver and spleen among treatments, no statistical difference was observed (p
209	>0.05) among Arg deficiency, and Arg and Cit supplementation groups.

211 Carcass weight

212 There was no significant difference (*p*>0.05) in carcass weight among treatments (Table 11).

213

214 **Enzyme activity**

There was no significant difference (p>0.05) in total NOS in the liver among treatments (Table 12). Arg1.25 group had the highest arginase in the liver and total NOS and arginase in the kidney than other treatments, but no statistical difference was observed (p>0.05) in arginase in the liver among Arg supplementation and Arg replacement with Cit.

- 219
- 220

221 **Discussion**

222 This study was conducted to investigate the effects of Arg supplementation and replacing Arg with 223 Cit in broiler chickens under cyclic heat stress. The results demonstrated that Arg deficiency (NC) 224 negatively impacted growth performance. However, the groups that received Arg and Cit 225 supplementation showed an improvement in growth performance counteracting the negative effects 226 caused by Arg deficiency. These findings are consistent with previous studies that have shown 227 impaired growth in birds fed Arg-deficient diets, which can be alleviated by supplementing with Arg 228 [25-28]. Additionally, Abdulkarimi et al. [29] reported that dietary supplementation of Arg beyond 229 20% of the NRC recommendation (Arg: Lys=1.26) resulted in improved BWG and FCR. The authors 230 suggested that polyamines, which are derived from Arg, may possess anabolic properties that enhance 231 protein synthesis and uptake of AA by cells [30]. One of our hypotheses was that the Arg requirement 232 for optimum growth would be higher than the current recommendation of 105% of Lys under HS. The 233 result of the present study supports the hypothesis as the optimum growth of brids was achieved at 234 125% of Lys, which is around 20% higher than the recommendation. It has been well documented 235 that under HS birds use part of arginine for production of NO that increases blood flow for radiation

of internal heat [31]. Given that the part of arginine is used for response to HS the requirement of Arg

237 for optimal protein deposition is increased.

238 Cit is a metabolite of Arg [32]. Previous studies have shown that Cit can be transported to the kidney 239 and other tissues where conditions for arginine synthesis are favorable without passing through the 240 liver and intestine [19-20]. On the other hand, Arg after absorption is transported to the liver where 241 significant amounts are metabolized, partly through increased arginase activity. Our study found no 242 significant difference in growth performance between groups that received direct Arg 243 supplementation and those that had partial Arg replacement with Cit groups. These results are 244 consistent with those reported by Uyanga et al. [33], who found no significant difference in growth 245 performance between the group supplemented with Arg and a group that had full Arg replacement 246 with Cit. While the partial replacement of Arg with Cit did not significantly improve growth performance compared to the Arg supplemented group in our study, Cit supplementation at 33% of 247 supplemented Arg did maintain growth similar to that of the group receiving direct Arg 248 supplementation. These findings could be indirectly support our hypothesis that Cit inclusion at 33% 249 or 50% of Arg will be sufficient to maintain optimum growth of broilers, as Cit is not metabolized in 250 251 the liver as Arg is.

252 The small intestine plays a vital role in digestion and nutrient absorption, making it one of the most 253 important digestive organs. Intestinal morphology is particularly important for assessing intestinal 254 health as it affects nutrient absorption, gut immunity, and gut barrier function [34]. In this study, the 255 groups supplemented with direct Arg showed improved VH and CD in the duodenum, jejunum, and 256 ileum. However, the relative weight of the small intestine was not affected. These findings are 257 consistent with a previous study by Murakami et al. [35], which found that Arg supplementation did 258 not impact the weight and length of the small intestine, but did improve intestinal morphology by 259 increasing VH and decreasing CD in broilers. Similarly, Castro et al. [36] and Zhang et al. [34] 260 reported that VH and CD in the small intestine increased with additional Arg supplementation in diets. Arg supplementation has also been shown to improve intestinal morphology after an inflammatory 261 262 injury caused by *Clostridium perfringens* and *Eimeria* spp. [37-39].

This tendency was also observed in nutrient digestibility during this study. We found that adding additional Arg improved the digestibility of CP and amino acids, particularly Lys, Met, Arg, and Cys, when compared to Arg-deficient diets. Previous research has reported a positive correlation betweenincreased nutrient absorption and gut morphology in relation to Arg [29, 40].

Arg plays an essential role in intestinal physiology [41]. It serves as a precursor of polyamines and can be considered a nutritional agent in promoting intestinal mucosal growth. Arg speeds up the mitotic process in the villus-crypt area, consequently increasing the number of villus cells [42]. Moreover, Arg can enhance the growth of intestinal cells by activating the mechanistic target of rapamycin (mTOR), toll-like receptor 4 (TLR 4), and promoting nitric oxide (NO) production [29, 42-43].

Arginase is an important enzyme that breaks down Arginine into urea and ornithine [7]. In broilers, the activity of kidney and liver arginase plays a crucial role in controlling the metabolism of Arg because it is necessary for the degradation of Arg [33]. NOS, also known as catalyzing enzymes, is responsible for converting Arg into NO. Previous studies have shown that supplementation of Arg increased the activity of arginase in the kidney and liver [33, 44-45].

278 In line with our objectives, our study has shown that supplementation with Arg increased the 279 activity of arginase and NOS in the liver and kidney. Arg is converted by NOS into Cit and NO, and 280 by arginases into L-ornithine for polyamine biosynthesis [46]. Interestingly, we also observed 281 increased production of NO in the blood when Arg and Cit were added to the diets. It is worth noting 282 that replacing Arg with Cit did not result in any changes in intestinal morphology, nutrient 283 digestibility, or NO production in the blood, compared to direct Arg supplementation in this study. 284 This finding is consistent with a previous study which found that a complete replacement of Arg with 285 Cit did not affect the concentrations of Arg and NO in the plasma [33]. These positive results from Cit 286 supplementation may be attributed to the improved availability of Arg through Cit supplementation. 287 HS has negative effects on poultry performance by causing physiological changes such as 288 hyperthermia, oxidative stress, and systemic inflammation [31, 47-48]. In this study, we exposed 289 broiler chickens to HS conditions and observed significant differences in growth performance, 290 intestinal morphology, and nutrient digestibility among the treatment groups. Chowdhury [49] 291 reported that an increase in blood Cit concentration is considered a biomarker of HS, as it is elevated 292 during short periods of HS. Additionally, Luiking et al. [50] found that NO concentration in the blood 293 can alleviate HS damage by improving vascular tone and blood flow in the smooth muscle.

We observed a significant difference in NO and Cit concentrations in the blood between the NC group and the Arg and Cit supplemented groups in this study. However, there was no significant difference in NO and Cit concentrations in the blood between the Arg and Cit supplemented groups in this study. These findings are in agreement with a previous study that reported that full Arg replacement with Cit did not affect NO concentration compared to the Arg supplementation group [33].

300

301 CONCLUSION

All things taken together, additional Arg supplementation improved NO production in the blood, which enhanced growth performance, gut morphology and nutrient digestibility. Also, Cit supplementation maintain growth, gut morphology, nutrient digestibility and even NO production in the blood similar with Arg supplementation, and thus Cit can replace with arginine in broiler's diet. Therefore, this study would be useful to prove the requirement of optimal Arg and Cit levels in poultry feeding programs.

308

309

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Tables

Table 1. Ingredient composition	-					
TRT	NC	PC	Arg1.15	Arg1.25	Cit33	Cit50
CORN	57.50	57.50	57.50	57.50	57.50	57.50
SBM (46%CP)	25.15	25.15	25.15	25.15	25.15	25.15
DDGS	6.00	6.00	6.00	6.00	6.00	6.00
Animal Fats	2.20	2.20	2.20	2.20	2.20	2.20
Animal Protein	6.25	6.25	6.25	6.25	6.25	6.25
Limestone	0.60	0.60	0.60	0.60	0.60	0.60
MDCP	0.40	0.40	0.40	0.40	0.40	0.40
SALT-Proc	0.20	0.20	0.20	0.20	0.20	0.20
Vit premix ^a	0.12	0.12	0.12	0.12	0.12	0.12
Min premix ^a	0.20	0.20	0.20	0.20	0.20	0.20
Liq-Choline	0.27	0.27	0.27	0.27	0.27	0.27
Sand	0.05	0.11	0.11		0.11	0.11
Glycine	0.36	0.16	0.02		0.21	0.16
DL-MET 99%	0.30	0.30	0.30	0.30	0.30	0.30
L-Lys-SO ₄ (55%)	0.40	0.40	0.40	0.40	0.40	0.40
Arginine 98%		0.14	0.28	0.41		
Citrulline					0.09	0.14
Calculated nutrient value						
ME, Kcal/kg	3030	3030	3030	3030	3030	3030
C protein	22.36	22.41	22.52	22.76	22.39	22.39
Total Ca	0.92	0.92	0.92	0.92	0.92	0.92
Total P	0.69	0.69	0.69	0.69	0.69	0.69
Total Na	0.20	0.20	0.20	0.20	0.20	0.20
Total Cl	0.30	0.30	0.30	0.30	0.30	0.30
SID Lys	1.35	1.35	1.35	1.35	1.35	1.35
SID TSAA	0.85	0.85	0.85	0.85	0.85	0.85
SID Arg	1.28	1.41	1.55	1.68	1.28	1.28

^aSupplied per kilogram diet: vitamin A (retinyl acetate), 9,000 IU; vitamin D3, 3,000 IU; vitamin E (DL-αtocopheryl acetate), 48 mg; vitamin K, 3 mg; thiamin, 1.8 mg; riboflavin, 6 mg; pyridoxine, 3 mg; vitamin B12, 0.012 mg; niacin, 42 mg; folic acid, 1.2 mg; biotin, 0.24 mg; pantothenic acid, 12 mg; manganese, 120 mg; zinc 100 mg; iron, 80 mg; copper, 20 mg; iodine, 2 mg; selenium, 0.3 mg; cobalt, 0.5 mg

DDGS, Dried distiller's grains with soluble; MDCP, Mono-dicalcium phosphate; SID, standardized ileal digestibility; TSAA, total sulfur amino acid; Lys, lysine; Arg, Arginine; MET, methionine; P, phosphorus; ME, metabolizable energy

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit50, supplementation of Cit at 50% of Arg in Arg1.15

Ingredient	NC	PC	Arg1.15	Arg1.25	Cit33	Cit50
CORN	59.50	59.50	59.50	59.50	59.50	59.50
SBM (46%CP)	24.00	24.00	24.00	24.00	24.00	24.00
DDGS	5.00	5.00	5.00	5.00	5.00	5.00
Animal Fats	2.20	2.20	2.20	2.20	2.20	2.20
Animal Protein	6.25	6.25	6.25	6.25	6.25	6.25
Limestone	0.60	0.60	0.60	0.60	0.60	0.60
MDCP	0.40	0.40	0.40	0.40	0.40	0.40
SALT-Proc	0.20	0.20	0.20	0.20	0.20	0.20
Vit-PX ^a	0.12	0.12	0.12	0.12	0.12	0.12
Min-PX ^a	0.20	0.20	0.20	0.20	0.20	0.20
Liq-Choline	0.27	0.27	0.27	0.27	0.27	0.27
Sand	0.20	0.20	0.20	0.20	0.20	0.20
Glycine	0.40	0.27	0.14		0.31	0.27
DL-MET 99%	0.28	0.28	0.28	0.28	0.28	0.28
L-Lys-SO4(55%)	0.38	0.38	0.38	0.38	0.38	0.38
Arginine 98%		0.13	0.26	0.40		
Citrulline					0.09	0.13
Calculated nutrient value			X			
ME, Kcal/kg	3050	3050	3050	3050	3050	3050
C protein	21.73	21.84	21.95	22.07	21.82	21.82
Total Ca	0.92	0.92	0.92	0.92	0.92	0.92
Total P	0.69	0.69	0.69	0.68	0.69	0.69
Total Na	0.19	0.19	0.19	0.19	0.20	0.20
Total Cl	0.30	0.30	0.30	0.30	0.30	0.30
SID Lys	1.30	1.30	1.30	1.30	1.30	1.30
SID TSAA	0.82	0.82	0.82	0.82	0.82	0.82
SID Arg	1.24	1.36	1.49	1.63	1.24	1.24

^aSupplied per kilogram diet: vitamin A (retinyl acetate), 9,000 IU; vitamin D3, 3,000 IU; vitamin E (DL-αtocopheryl acetate), 48 mg; vitamin K, 3 mg; thiamin, 1.8 mg; riboflavin, 6 mg; pyridoxine, 3 mg; vitamin B12, 0.012 mg; niacin, 42 mg; folic acid, 1.2 mg; biotin, 0.24 mg; pantothenic acid, 12 mg; manganese, 120 mg; zinc 100 mg; iron, 80 mg; copper, 20 mg; iodine, 2 mg; selenium, 0.3 mg; cobalt, 0.5 mg

DDGS, Dried distiller's grains with soluble; MDCP, Mono-dicalcium phosphate; SID, standardized ileal digestibility; TSAA, total sulfur amino acid; Lys, lysine; Arg, Arginine; MET, methionine; P, phosphorus; ME, metabolizable energy

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit50, supplementation of Cit at 50% of Arg in Arg1.15

Table 3. Ingredient compos	ition of expe	rimental diets	s (Phase 3/day	15–21)		
Ingredient	NC	PC	Arg1.15	Arg1.25	Cit33	Cit50
CORN	60.80	60.80	60.80	60.80	60.80	60.80
SBM (46%CP)	22.85	22.85	22.85	22.85	22.85	22.85
DDGS	5.00	5.00	5.00	5.00	5.00	5.00
Animal Fats	2.20	2.20	2.20	2.20	2.20	2.20
Animal Protein	6.25	6.25	6.25	6.25	6.25	6.25
Limestone	0.60	0.60	0.60	0.60	0.60	0.60
MDCP	0.40	0.40	0.40	0.40	0.40	0.40
SALT-Proc	0.20	0.20	0.20	0.20	0.20	0.20
Vit-PX ^a	0.12	0.12	0.12	0.12	0.12	0.12
Min-PX ^a	0.20	0.20	0.20	0.20	0.20	0.20
Liq-Choline	0.27	0.27	0.27	0.27	0.27	0.27
Sand	0.10	0.17	0.17	0.17	0.17	0.17
Glycine	0.45	0.26	0.19		0.32	0.28
DL-MET 99%	0.19	0.19	0.19	0.19	0.19	0.19
L-Lys-SO4(55%)	0.37	0.37	0.37	0.37	0.37	0.37
Arginine 98%		0.12	0.19	0.38		
Citrulline					0.06	0.10
Calculated nutrient value						
ME, Kcal/kg	3060	3060	3060	3060	3060	3060
C protein	21.30	21.32	21.43	21.54	21.31	21.30
Total Ca	0.91	0.91	0.91	0.91	0.91	0.91
Total P	0.68	0.68	0.68	0.68	0.68	0.68
Total Na	0.19	0.19	0.19	0.19	0.19	0.19
Total Cl	0.28	0.28	0.28	0.28	0.28	0.28
SID Lys	1.26	1.26	1.26	1.26	1.26	1.26
SID TSAA	0.72	0.72	0.72	0.72	0.72	0.72
SID Arg	1.21	1.32	1.45	1.58	1.21	1.21

^aSupplied per kilogram diet: vitamin A (retinyl acetate), 9,000 IU; vitamin D3, 3,000 IU; vitamin E (DL-αtocopheryl acetate), 48 mg; vitamin K, 3 mg; thiamin, 1.8 mg; riboflavin, 6 mg; pyridoxine, 3 mg; vitamin B12, 0.012 mg; niacin, 42 mg; folic acid, 1.2 mg; biotin, 0.24 mg; pantothenic acid, 12 mg; manganese, 120 mg; zinc 100 mg; iron, 80 mg; copper, 20 mg; iodine, 2 mg; selenium, 0.3 mg; cobalt, 0.5 mg

DDGS, Dried distiller's grains with soluble; MDCP, Mono-dicalcium phosphate; SID, standardized ileal digestibility; TSAA, total sulfur amino acid; Lys, lysine; Arg, Arginine; MET, methionine; P, phosphorus; ME, metabolizable energy

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit50, supplementation of Cit at 50% of Arg in Arg1.15

 Table 4. Ingredient composition of experimental diets (Phase 4/day 22–32)
 Ingredient NC PC Arg1.15 Arg1.25 Cit33 Cit50 TRT CORN 62.46 62.46 62.46 62.46 62.46 62.46 SBM (46%CP) 21.15 21.15 21.15 21.15 21.15 21.15 DDGS 5.00 5.00 5.00 5.00 5.00 5.00 2.20 2.20 2.20 Animal Fats 2.20 2.20 2.20 Animal Protein 6.25 6.25 6.25 6.25 6.25 6.25 Limestone 0.60 0.60 0.60 0.60 0.60 0.60 MDCP 0.40 0.40 0.40 0.40 0.40 0.40 0.20 SALT-Proc 0.20 0.20 0.20 0.20 0.20 Vit-PX^a 0.12 0.12 0.12 0.12 0.12 0.12 Min-PX^a 0.20 0.20 0.20 0.20 0.20 0.20 Liq-Choline 0.27 0.27 0.27 0.27 0.27 0.27 Sand 0.25 0.25 0.25 0.25 0.25 0.25 Glycine 0.13 0.36 0.25 0.29 0.24 **DL-MET 99%** 0.19 0.19 0.19 0.19 0.19 0.19 L-Lys-SO4(55%) 0.35 0.35 0.35 0.35 0.35 0.35 0.11 0.23 Arginine 98% 0.36 Citrulline 0.07 0.12 **Calculated nutrient value** 3070 ME, Kcal/kg 3070 3070 3070 3070 3070 C protein 20.52 20.62 20.72 20.83 20.61 20.60 0.91 Total Ca 0.91 0.91 0.91 0.91 0.91 0.69 0.69 Total P 0.69 0.69 0.69 0.69 0.19 0.19 Total Na 0.19 0.19 0.19 0.19 0.29 Total Cl 0.29 0.29 0.29 0.29 0.29 SID Lys 1.21 1.21 1.21 1.21 1.21 1.21 0.71 SID TSAA 0.71 0.71 0.71 0.71 0.71 SID Arg 1.16 1.27 1.38 1.51 1.16 1.16

^aSupplied per kilogram diet: vitamin A (retinyl acetate), 9,000 IU; vitamin D3, 3,000 IU; vitamin E (DL-αtocopheryl acetate), 48 mg; vitamin K, 3 mg; thiamin, 1.8 mg; riboflavin, 6 mg; pyridoxine, 3 mg; vitamin B12, 0.012 mg; niacin, 42 mg; folic acid, 1.2 mg; biotin, 0.24 mg; pantothenic acid, 12 mg; manganese, 120 mg; zinc 100 mg; iron, 80 mg; copper, 20 mg; iodine, 2 mg; selenium, 0.3 mg; cobalt, 0.5 mg

DDGS, Dried distiller's grains with soluble; MDCP, Mono-dicalcium phosphate; SID, standardized ileal digestibility; TSAA, total sulfur amino acid; Lys, lysine; Arg, Arginine; MET, methionine; P, phosphorus; ME, metabolizable energy

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit50, supplementation of Cit at 50% of Arg in Arg1.15

Table 5. Effects of dietary arginine or citrulline on growth performance in broiler

Items	NC	PC	Arg1.15	Arg1.25	Cit33	Cit50	SEM	<i>p</i> -value
BW, g								
D0	34.50	34.44	34.56	34.50	34.28	34.33	0.439	0.997
D7	127.39	136.06	138.28	133.28	126.61	124.00	4.388	0.162
D21	913.56	902.17	930.56	972.17	929.78	903.89	18.872	0.125
D32	1658.50 ^b	1673.72 ^{ab}	1729.67 ^{ab}	1788.72ª	1726.17 ^{ab}	1671.94 ^{ab}	29.621	0.035
BWG, g								
D 0 to 7	92.89	101.61	103.72	98.78	92.33	89.67	4.490	0.192
D 7 to 21	786.17	766.11	792.28	838.89	803.17	779.89	16.425	0.067
D 21 to 32	744.94	771.56	799.11	816.56	796.39	768.06	32.462	0.672
D 0 to 32	1624.00 ^b	1639.28 ^{ab}	1695.11 ^{ab}	1754.22ª	1691.89 ^{ab}	1637.61 ^{ab}	28.430	0.018
FI, g								
D 0 to 7	110.77	114.48	122.23	114.61	112.30	111.62	6.371	0.826
D 7 to 21	1056.80	1059.65	1056.94	1085.28	1034.26	1027.69	26.44	0.698
D 21 to 32	1413.32	1389.52	1318.09	1297.02	1371.02	1354.94	30.33	0.097
D 0 to 32	2580.88	2563.66	2497.26	2496.91	2517.57	2494.25	39.77	0.489
FCR		C						
D 0 to 7	1.22	1.16	1.21	1.19	1.26	1.27	0.032	0.196
D 7 to 21	1.36	1.39	1.35	1.31	1.30	1.33	0.026	0.169
D 21 to 32	1.93	1.85	1.69	1.61	1.76	1.80	0.073	0.063
D 0 to 32	1.60 ^a	1.58ª	1.49 ^{ab}	1.44 ^b	1.51 ^{ab}	1.54 ^{ab}	0.028	0.002
FCR at 1.5kg	1.56 ^a	1.53 ^a	1.42 ^{ab}	1.35 ^b	1.44 ^{ab}	1.48 ^{ab}	0.033	0.002
PI	329.43 ^b	337.52 ^b	371.23 ^{ab}	395.96 ^a	366.04 ^{ab}	346.01 ^b	10.803	0.001

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit50, supplementation of Cit at 50% of Arg in Arg1.15

BW, body weight; BWG, body weight gain; FI, feed intake; FCR, feed conversion ratio; PI, production index SEM, stand error of means

^{a, b} Means within a column with different superscripts differ significantly (p < 0.05).

Table 6. Effects of dietary arginine or citrulline on nutrient apparent ileal digestibility

Items, %	NC	PC	Arg1.15	Arg1.25	Cit33	Cit50	SEM	<i>p</i> -value
DM	67.64	67.91	68.09	68.15	67.90	67.78	0.158	0.241
СР	77.62 ^c	78.50 ^{bc}	79.35 ^{ab}	80.61ª	79.62 ^{ab}	79.36 ^{ab}	0.351	< 0.001
GE	72.71	72.66	72.83	72.98	72.97	72.63	0.256	0.872

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit50, supplementation of Cit at 50% of Arg in Arg1.15

DM, dry matter; CP, crude protein; GE, gross energy; SEM, stand error of means

^{a-c} Means within a column with different superscripts differ significantly (p < 0.05).

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Items, %	NC	РС	Arg1.15	Arg1.25	Cit33	Cit50	SEM	<i>p</i> -value
EAA								
Arginine	86.33°	87.10 ^{bc}	88.09 ^{abc}	89.45ª	88.46 ^{ab}	88.92 ^{ab}	0.457	< 0.001
Histidine	85.54	86.38	87.63	87.29	88.41	88.01	1.184	0.548
Isoleucine	86.78	86.73	87.10	87.45	86.94	87.09	0.637	0.973
Leucine	86.94	86.98	87.14	87.34	87.39	87.62	0.212	0.208
Lysine	86.49°	86.91 ^{bc}	87.88 ^{ab}	88.90 ^a	88.40 ^a	88.19 ^{ab}	0.316	< 0.001
Methionine	84.77 ^b	85.59 ^{ab}	86.60 ^{ab}	88.90ª	87.84 ^{ab}	88.64ª	0.789	0.003
Phenylalanine	88.16	88.40	87.94	88.46	88.09	88.03	0.155	0.142
Threonine	87.73	87.70	87.81	87.78	87.72	87.92	0.109	0.752
Valine	88.88	89.27	88.86	88.73	88.83	88.80	0.283	0.801
Tryptophan	86.34	87.15	87.45	87.49	88.25	87.10	0.777	0.671
NEAA				X				
Alanine	87.98	86.99	87.42	88.41	88.35	87.93	0.438	0.200
Aspartic	88.81	89.12	89.28	89.58	88.87	89.19	0.304	0.513
Cystine	87.18 ^b	88.41 ^{ab}	88.59 ^{ab}	89.30ª	88.90 ^{ab}	88.34 ^{ab}	0.404	0.021
Glycine	88.38	88.29	88.85	89.23	88.48	88.95	0.404	0.530
Glutamic acid	88.19	88.65	88.07	88.59	88.19	87.98	0.194	0.102
Proline	87.24	87.36	87.38	87.15	88.04	87.34	0.305	0.385
Serine	86.38	86.51	86.71	87.15	86.23	86.56	0.379	0.624
Tyrosine	87.24	87.10	86.67	87.34	86.19	87.32	0.940	0.944

Table 7. Effects of dietary arginine or citrulline on amino acid apparent ileal digestibility

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit2, supplementation of Cit at 50% of Arg in Arg1.15

EAA, essential amino acid; NEAA, non-essential amino acid; SEM, stand error of means

^{a-c} Means within a column with different superscripts differ significantly (p < 0.05).

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Items	NC	РС	Arg1.15	Arg1.25	Cit33	Cit50	SEM	<i>p</i> -value
TP, (g/dL)	3.10	3.30	3.25	2.97	3.22	3.13	0.081	0.081
BUN, (mg/dL)	2.17	1.83	1.83	2.17	2.33	2.17	0.175	0.266
Cortisol, (µg/dL)	0.04	0.04	0.04	0.04	0.04	0.03	0.006	0.831
Arginine, (µmol/L)	27.98	29.98	30.67	30.59	30.10	27.68	1.505	0.576
NO, (µmol/L)	2.12 ^b	3.09 ^{ab}	3.49 ^a	3.65 ^a	2.91 ^{ab}	2.36 ^b	0.248	0.001
Citrulline, (nmol/mL)	31.94 ^b	33.18 ^{ab}	33.87 ^{ab}	38.70ª	35.16 ^{ab}	32.32 ^{ab}	1.512	0.039

Table 8. Effects of dietary arginine or citrulline on blood profile in broiler

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit50, supplementation of Cit at 50% of Arg in Arg1.15

TP, total protein; BUN, blood urea nitrogen; NO, nitric oxide; SEM, stand error of means

^{a, b} Means within a column with different superscripts differ significantly (p < 0.05).

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Items, µm	NC	PC	Arg1.15	Arg1.25	Cit33	Cit50	SEM	<i>p</i> -value
Duodenum								
VH	148.58 ^b	158.74 ^{ab}	170.76ª	173.44 ^a	167.05ª	160.74 ^{ab}	6.998	0.003
CD	10.88	10.67	11.56	9.64	11.09	11.85	0.593	0.162
VCR	13.96 ^b	15.17 ^{ab}	14.89 ^{ab}	18.03 ^a	15.09 ^{ab}	14.10 ^b	0.898	0.039
Jejunum								
VH	116.74 ^b	139.95ª	142.38ª	155.46 ^a	149.89ª	134.87 ^{ab}	5.250	< 0.001
CD	18.82ª	15.52 ^{ab}	15.15 ^{ab}	13.15 ^b	13.77 ^b	15.34 ^{ab}	0.959	0.005
VCR	6.28 ^b	9.65 ^{ab}	9.70 ^{ab}	11.98 ^a	11.07ª	8.84 ^{ab}	0.839	0.001
Ileum					\mathcal{A}			
VH	75.29 ^b	82.51 ^{ab}	84.12 ^{ab}	101.17 ^a	96.16ª	84.98 ^{ab}	4.378	0.003
CD	18.89ª	15.82 ^{ab}	14.79 ^{abc}	10.61°	11.61 ^{bc}	14.48 ^{abc}	1.074	< 0.001
VCR	4.00 ^c	5.33°	5.80 ^{bc}	9.82ª	8.66 ^{ab}	6.21 ^{bc}	0.669	< 0.001

 Table 9. Effects of dietary arginine or citrulline on intestinal morphology of small intestine in broiler

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit50, supplementation of Cit at 50% of Arg in Arg1.15

VH, villus height; CD, crypt depth; VCR, villus to crypt ratio; SEM, stand error of means

^{a-c} Means within a column with different superscripts differ significantly (p < 0.05).

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Items, (g/kg of live body weight)	NC	РС	Arg1.15	Arg1.25	Cit33	Cit50	SEM	<i>p</i> -value
Gizzard	21.52	22.99	18.56	21.79	24.40	21.92	1.893	0.410
Liver	22.17 ^{ab}	20.43 ^b	20.09 ^b	20.42 ^b	20.10 ^b	25.05ª	0.903	0.003
Spleen	0.76 ^{abc}	0.97ª	0.66 ^{bc}	0.79 ^{ab}	0.63 ^{bc}	0.50°	0.065	0.001
Bursa of Fabricius	1.55	1.95	1.55	1.63	1.92	1.32	0.244	0.445
Small intestine	40.70	33.76	38.31	40.38	38.24	39.01	1.581	0.053

Table 10. Effects of dietary arginine or citrulline on internal organ weight in broiler

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit50, supplementation of Cit at 50% of Arg in Arg1.15

SEM, stand error of means

^{a-c} Means within a column with different superscripts differ significantly (p < 0.05).

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Table 11. Effects of dietary arginine or citrulline on carcass weight in broiler

Items, (g/kg of live body weight)	NC	PC	Arg1.15	Arg1.25	Cit33	Cit50	SEM	<i>p</i> -value
Abdominal fat	10.40	9.63	11.75	10.00	9.79	10.49	0.653	0.258
Thigh	83.69	86.91	85.71	82.07	81.56	86.74	3.172	0.740
Drumstick	92.46	93.79	91.11	94.27	87.99	90.44	3.202	0.751
Breast	218.41	211.51	213.62	215.19	208.41	213.59	6.084	0.905

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit50, supplementation of Cit at 50% of Arg in Arg1.15

SEM; stand error of means

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Table 12. Effects of dietary arginine or citrulline on enzyme activity

Items, (ng/mg protein)	NC	РС	Arg1.15	Arg1.25	Cit33	Cit50	SEM	<i>p</i> -value
Liver								
Total NO synthase	17.71	18.69	22.67	22.81	21.27	19.95	1.659	0.191
Arginase	2.86b	3.39b	3.99ab	5.06a	3.88ab	3.97ab	0.276	< 0.001
Kidney								
Total NO synthase	15.61b	16.10b	17.69b	24.53a	16.55b	15.29b	1.150	< 0.001
Arginase	2.52c	3.69b	3.68b	4.59a	3.69b	3.79b	0.138	< 0.001

NC, Arg: Lys=0.95; PC, Arg: Lys=1.05; Arg1.15, Arg: Lys=1.15; Arg1.25, Arg: Lys=1.25. Arg, arginine; Lys, lysine; Cit33, supplementation of Cit at 33% of Arg in Arg1.15; Cit50, supplementation of Cit at 50% of Arg in Arg1.15

NO, nitric oxide; SEM, stand error of means

a, b Means within a column with different superscripts differ significantly (p < 0.05).

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