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Ethics approval and consent to	I he experimental protocol for this study was reviewed and approved by the
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#### 8 Abstract (up to 350 words)

9 In the swine industry, supplementation with high levels of zinc oxide and copper sulfate in the weaner diet could 10 be excreted through feces without being normally absorbed in the intestine, resulting in environmental pollution. 11 Therefore, the various methods have been proposed to address this issue. The objective of this study was to 12 investigate effects of a low dietary dose of coated copper sulfate ( $CuSO_4$ ) and zinc oxide (ZnO) on growth 13 performance, frequency of diarrhea, nutrient digestibility, and immune responses of weaned pigs. The four dietary 14 treatments were (1) a basal weaner diet based on corn and soybean meal (CON), (2) CON supplemented with 15 2,500 ppm standard ZnO (T1), (3) CON supplemented with 100 mg/kg dietary coated CuSO<sub>4</sub> and 100 mg/kg 16 dietary coated ZnO (T2), and (4) CON supplemented with 200 mg/kg dietary coated CuSO<sub>4</sub> and 200 mg/kg dietary 17 coated ZnO (T3). Dietary T2 and T3 increased (p < 0.05) the average daily gain for the first two weeks and the 18 overall experimental period compared to that with CON. In addition, the groups supplemented with Cu and Zn tended to have a decreased (p < 0.10) frequency of diarrhea. Pigs fed dietary T2 and/or T3 had lower (p < 0.10) 19 20 number of white blood cells on day 7 and hematocrit on day 14 compared to those fed CON. However, no 21 difference was observed in the number of red blood cells among the dietary treatments. Regarding immune responses, dietary T2 decreased (p < 0.10) serum tumor necrosis factor- $\alpha$  on day 7 and increased (p < 0.10) 22 23 immunoglobulin G on day 14 compared with CON. Moreover, pigs fed dietary T2 tended to have increased 24 *Limosilacatobacilus* (p < 0.10). Dietary T3 had higher (p < 0.05) relative abundance of the genus Agathobacter 25 compared to those fed CON and dietary T1 and decreased (p < 0.05) genus *Terrisporobacter* compared to those 26 fed dietary T1. These results suggested the supplementation of dietary coated ZnO and CuSO<sub>4</sub> enhanced growth 27 performance and modulated immune responses associated with changes in the fecal microbiota composition.

28 Keywords: Copper; Growth performance; Immune response; Microbiota; Weaned pig; Zinc

# **INTRODUCTION**

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32 Post-weaning diarrhea (PWD) is a potentially fatal disease in swine production worldwide, causing 33 dehydration, growth delay, and death in severe cases causes of negative impacts on the industry [1]. During the 34 post-weaning period, various stress factors, including dietary, environmental, social, and physiological stressors, 35 affect weaning pigs. These factors reduce feed intake and growth, thus resulting in intestinal dysfunction, and an 36 increased susceptibility to inflammation [2,3]. Furthermore, permeability increases as the intestinal barrier is 37 disrupted, leading to PWD due to infections by intestinal pathogens such as Escherichia coli (E. coli) [4]. To solve 38 these issues, in-feed antibiotics have been widely utilized in the livestock industry for growth promotion and 39 disease treatment including PWD. However, the incidence of antimicrobial resistance and residual issues remain 40 concerning for animal and public health due to the overuse of in-feed antibiotics[5]. Therefore, various nutritive 41 alternatives such as probiotics, enzymes, minerals, and others, have been utilized to enhance animal health and 42 performance [6].

43 It is known that copper (Cu) and zinc (Zn) are vital trace elements that act as components of metabolic 44 enzymes and perform biological functions [7]. Both support immunity, reproduction, and growth of animals [8,9]. 45 The most commercial forms in the swine industry are zinc oxide (ZnO) and copper sulfate (CuSO4) because of 46 their relatively low cost compared with other forms [10,11]. These act as alternatives to in-feed antibiotics for 47 promoting growth and have antimicrobial effects such as reducing diarrhea incidence in weaned pigs at 48 concentration levels exceeding normal nutritional requirements according to the National Research Council [11]. 49 Specifically, pharmacological levels (2,000 to 4,000 mg/kg) of ZnO have been commonly supplemented in 50 weanling diets to promote growth performance and reduce diarrhea frequency by enhancing morphological 51 structure and maintaining gut integrity [12,13]. In addition, previous studies reported that a dose of 150 to 250 52 mg/kg supplementation of CuSO<sub>4</sub> stimulates growth rate, and feed intake, and sustains fecal consistency via 53 modulating gut microbiota homeostasis in the intestine [14,15].

However, supplementation of high doses of CuSO<sub>4</sub> (200 to 250 mg/kg) and ZnO (2,000 to 4,000 mg/kg) in the nursery diet can be excreted through manure without being normally absorbed into the intestine, resulting in environmental pollution [16–18]. Based on these issues, the European Union has legislated new maximum levels to limit Zn and Cu supplementation to 150 mg/kg up to 4 weeks after weaning [19,20]. Thus, new forms of Cu and Zn at lower doses have been proposed to reduce excretions and improve growth performance during the 59 weaning phase. Lipid-coated and concentrated forms of ZnO and CuSO<sub>4</sub>, protect against the formation of insoluble 60 complexes with other minerals and dissociation in the stomach. They are dissociated by pancreatic lipase, and 61 efficiently absorbed in the small intestine of monogastric animals [21-23]. A previous in vitro study presented the 62 dissociation percentage of coated and uncoated ZnO in the stomach. The results showed that the percentages were 63 25.03% for coated and 85.26% in uncoated ZnO [24]. Consequently, this reduced the quantity released into the 64 soil through the manure. Therefore, the objective of this study was to evaluate the effects of low doses of lipid-65 coated CuSO<sub>4</sub> and ZnO on the growth performance, diarrhea, nutrient digestibility, immune responses, and fecal 66 microbiota of weaned pigs.

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# MATERIALS AND METHODS

All the experimental protocol for this study was reviewed and approved by the Institutional Animal Care
 and Use Committee of Chungnam National University, Daejeon, Republic of Korea. (approval# 202103A-CNU 080)

#### 72 Source of tested products

The lipid-coated ZnO and CuSO<sub>4</sub> practiced in this study were provided by a commercial company (ACC Inc., Seongnam, Republic of Korea). These products are concentrated forms of Zn and Cu from zinc oxide (ZnO) and copper sulfate pentahydrate (CuSO<sub>4</sub>·5H2O) which are microencapsulated in a lipid matrix by fatty acids and hydrogenated palm oil according to the manufacturer's information.

#### 77 Experimental design, animals, and diets

78 A total of 96 weaned piglets [(Landrace  $\times$  Yorkshire)  $\times$  Duroc; 7.29  $\pm$  0.69 kg of average initial body 79 weight (BW)] were assigned to four dietary treatments (4 pigs/pen; 6 replicates/treatment) in a randomized 80 complete block design (block = initial BW). The dietary treatments were (1) basal weaner diet based on corn-81 soybean meal (CON), (2) CON supplemented with 2,500 ppm standard ZnO (T1), (3) CON supplemented with 82 100 mg/kg dietary coated CuSO<sub>4</sub> and 100 mg/kg dietary coated ZnO (T2), and (4) CON supplemented with 200 83 mg/kg dietary coated CuSO<sub>4</sub> and 200 mg/kg dietary coated ZnO (T3). The basal diet was mixed to meet or exceed 84 the nutritional requirements of the National Research Council (NRC, 2012) for weaned pigs. The study was 85 conducted for 6 weeks, and the pigs were allowed *ad libitum* access to feeders and water and were housed in the 86 same-sized pen  $(2 \text{ m} \times 2 \text{ m})$  throughout the experimental period.

#### 87 Data and sample collection

88 In each pen, pigs' BW and remaining feed were weighed on days 1, 14, and 42 to figure out the average 89 daily gain (ADG), average daily feed intake (ADFI), and gain to feed ratio (G:F). The fecal score of the pigs was 90 visually monitored in each pen by two independent observers during the first 2 weeks. The score ranged from 1 91 to 5 (1 = hard and dry feces, 2 = soft feces, 3 = moist feces, 4 = mild diarrhea, and 5 = mild and severe diarrhea) 92 and were calculated by counting the number of pen days with a diarrhea score of 4 or higher as a percentage. 93 Blood samples were collected from the jugular vein of one randomly selected pig per pen using 10 mL tubes with 94 or without ethylenediaminetetraacetic acid (EDTA) on days 1, 7, and 14. Blood samples from tubes without EDTA 95 were left to clot at room temperature for 2 hours and then centrifuged for 15 min at  $3,000 \times g$  at 4 °C to obtain 96 serum. These samples were stored at -80°C for immune response analysis. In the final week of the experiment, 97 0.2 % chromic oxide (Cr2O3) was fed to all pigs as an indigestible marker. Fecal samples were collected from a 98 randomly one selected pig per pen using rectal palpation for three days after the adaption period and stored at 99  $-20^{\circ}$ C to measure nutrient digestibility [25,26]. Fecal samples were obtained from three randomly chosen pigs in 100 each dietary treatment on the last day of the experiment and stored at -80°C until metagenomic and fecal microbial 101 analysis

#### 102 Nutrient digestibility analysis

Diets and fecal samples were dried using a forced-air drying oven at 65°C for 72 h and then ground 103 104 through a grinder (80350, Hamilton Beach Inc, Virginia, USA) for apparent total tract digestibility (ATTD) 105 analysis. All ground samples were examined for dry matter (DM), crude protein (CP) by Kjeldahl method, and 106 energy using a bomb calorimeter (Parr 1281 Bomb Calorimeter, Parr Instrument, Moline, IL, USA) following the 107 procedures of the Association of Official Analytical Chemists [27]. The concentration of chromium in the samples 108 was determined using an absorption spectrophotometer (Hitachi Z-5000 Absorption Spectrophotometer, Hitachi 109 High-Technologies Co., Tokyo, Japan). The ATTD of the DM, CP, and energy for each dietary treatment were 110 calculated according to the previous study [28].

#### 111 Blood samples analysis

112 Whole blood samples were collected in EDTA tubes using an automated hematology analyzer (scil Vet 113 abc hematology analyzer; scil animal care company, F-67120 Altorf, France) [29]. The measurements included 114 the numbers of white blood cell (WBC), red blood cell (RBC), and hematocrit (HCT). The serum samples were 115 applied to determine immune responses including tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and cortisol using porcine 116 enzyme-linked immunosorbent assay kits (ELISA) kits (R&D Systems Inc., Minneapolis, MN, USA). 117 Additionally, levels of serum immunoglobulin A (IgA), serum immunoglobulin G (IgG), and serum 118 immunoglobulin M (IgM) were determined using ELSA kits (Bethyl Laboratories, Inc., Waltham, MA, USA). All

assays were performed according to the manufacturer's instructions.

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121 Fecal microbiota analysis

122 DNA was extracted from fecal samples (200 mg of feces per sample) using the QIAamp Fast DNA Stool 123 Mini Kit (QIAGEN, Hilden, Germany) based on the manufacturer's protocol. The concentration of DNA was 124 measured using the Colibri Microvolume Spectrometer (Titertek Berthold, Pforzheim, Germany), and the samples 125 with OD260/280 ratios between 1.80 and 2.15 were used to additional analysis [30]. The V5 to V6 regions of the 126 16S rRNA genes were amplified using sets of polymerase chain reaction (PCR) primers consisted of 799F-mod6 and 127 114R [31]. After PCR amplification, the products were refined using a Wizard® SV Gel and PCR Clean Up System 128 purification kit (Promega, Madison, United States). The sequencing of purified 16S rRNA gene was performed using the Illumina MiSeq platform at BRD Inc (Dongtan, Republic of Korea) following the manufacturer's 129 130 protocols. Quality control of all raw sequence data was checked utilizing FastQC [32], and then the 16S rRNA 131 gene sequences were analyzed using the Deblur algorithm, which is executed in both QIIME2 software and the 132 Microbiome Helper pipeline [33]. After applying the algorithm, sequences were grouped into operational 133 taxonomic units (OTUs), determined at a similarity cutoff of 97% [34]. Alpha diversity indices such as the 134 observed OTUs, Shannon, Simpson, and Chaol were measured to compare the diversity of microbial communities 135 within each dietary treatment. In addition, principal coordinated analysis (PCoA) based on unweighted and 136 weighted UniFrac distances was used to visualize differences in microbial communities among the dietary 137 treatments. Taxonomic composition of the dietary treatments was expressed as a percentage at the phylum and 138 genus levels based on their relative abundance.

#### 139 Statistical analyses

140 Data were subjected to the GLM procedure of SAS (SAS Inst., Cary, NC, USA) using a randomized 141 complete block design (block = initial BW). The experimental unit was the pen. The statistical models for growth 142 performance, nutrient digestibility, blood profiles, and immune responses of weaned pigs included the effects of 143 dietary treatment as the main effect and initial BW as a covariate. The frequency of diarrhea was analyzed using 144 the Chi-square test. The MicrobiomeAnalyst webtool (https://www.microbiomeanalyst.ca/) was used to analyze 145 alpha and beta diversity. STAMP software v. 2.1.3 [35] was used for taxonomic classification using a two-sided 146 Welch's t-test. Alpha diversity indices were measured using ANOVA and beta diversity based on unweighted and 147 weighted UniFrac distances was estimated using ANOSIM to determine the differences in microbial diversity 148 among the dietary treatments. Statistical difference and tendency for dietary treatment effects were set at p < 0.05

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### RESULTS

152 Growth performance, frequency of diarrhea, and nutrient digestibility

Pigs fed dietary T2 and T3 had higher (p < 0.05) ADG and G:F on day 1 to 14 than those fed CON (Table 2). Additionally, the groups supplemented with Cu and Zn tended to have a lower (p < 0.10) frequency of diarrhea than those fed CON. However, no differences were found in overall average ADFI for the first 14 days after weaning among the dietary treatments. In addition, dietary T2 and T3 increased (p < 0.05) ADG over the entire experimental period compared with the CON. As shown in Table 3, no differences were found ATTD of DM, CP and energy among dietary treatments.

#### 159 **Blood profiles and immune responses**

Pigs fed dietary T2 tended to have lower (p < 0.10) number of WBC on day 7 and HCT on day 14 than those fed CON (Table 4). However, there was no difference in RBC among dietary treatments. Regarding immune responses (Table 4), dietary T1 tended to have a lower (p < 0.10) concentration of serum cortisol on day 7 and a higher (p < 0.10) serum immunoglobulin M (IgM) than CON. In addition, dietary T2 tended to have decreased (p< 0.10) serum concentrations of TNF- $\alpha$  on day 7 and increased (p < 0.10) serum IgG on day 14 compared with CON. However, no differences were found in serum IgA levels among the dietary treatments.

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#### 167 Fecal microbiota

The microbial alpha diversity indices are shown in Fig. 1. Dietary T1 tended to have lower (p < 0.10) 168 169 Chao1, Simpson, and Shannon indices than CON and T3. The beta diversity of the fecal microbiota determined 170 using PCoA plots is presented in Fig. 2. PCoA plots based on the unweighted UniFrac distance, confirmed that 171 T1 had distinct clustering from other groups, but overlapped clustering with CON, T2, and T3 (R = 0.463,  $p < 10^{-1}$ 172 0.05). However, there was some distinct separation of fecal microbial communities based on the weighted UniFrac 173 distance among the dietary treatments (R = 0.201, p < 0.10). The relative abundances of the fecal microbes at the 174 phylum and genus level among the dietary treatments are shown in Figures 3 and 4, respectively. At the phylum 175 level (Fig. 3), Firmicutes were the most predominant bacteria in all dietary treatments (CON, 83.82%; T1, 82.98%; T2, 91.44%; T3, 80.19%), followed by Bacteroidetes in CON (5.83%) and T1 (14.67%), and Actinobacteria and 176 177 Proteobacteria in T2 (2.9%) and T3 (7.16%). At the genus level (Fig. 4), dietary T2 tended to increase (p < 0.10) 178 relative abundance of Limosilactobacilus (24.77%) compared to those fed dietary T1 (0.33%). In addition, pigs

179	fed dietary T3 had higher ( $p < 0.05$ ) relative abundance of <i>Agathobacter</i> (3.39%) than those fed CON (1.26%)
180	and dietary T1 (0.04%). Additionally, dietary T3 had a lower relative abundance of Terrisporobacter (1.74%)
181	compared to those fed dietary T1 (12.77%).

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### DISCUSSION

185 Minerals are inorganic elements that improve growth and reproduction in pigs [11]. In the livestock industry, micro minerals such as Cu and Zn, typically in the form of CuSO4 and ZnO are widely supplemented 186 187 into the diet in amounts that exceed the nutritional requirements of weanling pigs [24-26]. However, when these compounds reach the stomach of piglets, large amounts of ZnO and CuSO<sub>4</sub> are dissociated into Zn and 188 Cu ions, respectively, and great quantities are lost in the digestive tract. Because only a small amount of these 189 190 complexes can reach the intestinal tract, high doses are required. However, the inclusion level in feed must be 191 lowered owing to global regulations on environmental pollution through excretion and/or malabsorption by 192 overnutrition. Therefore, modified forms such as organic, nanoparticles and lipid-coated forms have been researched and utilized into swine feed [27-30]. Results from this study demonstrated that the dietary T2 and 193 T3 groups enhanced the ADG and G:F for the first two weeks and the overall period compared with that of the 194 195 CON group, which is consistent with previous studies where supplementation in coated or nano-sized forms 196 [31–33]. An improvement in growth performance was observed with the supplementation of a lower dosage of 197 dietary-coated CuSO<sub>4</sub> and ZnO in the present study due to the relatively high bioavailability and absorption of 198 Cu and Zn in the small intestine compared with standard forms of CuSO<sub>4</sub> and ZnO. In general, no additive 199 effects were observed when excess Zn was added to Cu [39]. Specifically, metallothionein in the intestinal 200 mucosa is induced by high concentrations of Zn, resulting in Cu binding, and causing Cu deficiency by 201 disturbing its absorption [40,41]. Therefore, the results suggested that balanced doses of coated CuSO<sub>4</sub> and 202 ZnO were better absorbed and enhanced growth rate compare with CON as same as supplementation of 203 pharmacological levels of ZnO.

In this study, supplementation with dietary coated  $CuSO_4$  and ZnO tended to have decreased the frequency of diarrhea. In addition, HCT in the blood profiles, which increases with dehydration and is used as an indicator of increases in diarrhea [42], did not differ among the dietary treatments in the first week after 207 weaning. However, supplementation with low dose of dietary coated microminerals tended to decrease HCT 208 on day 14, and we demonstrated that this supplementation positively affected the fecal score. Furthermore, no 209 differences in the ATTD of DM, CP, and energy among the dietary treatments. However, several previous 210 studies have reported that ZnO and CuSO<sub>4</sub> supplementation in the form of lipid-coated or nano-type positively 211 affected nutrient and energy digestibility, which were attributed to digestive enzymes and morphological 212 changes in the small intestine [43-45]. Therefore, additional research on digestive enzyme activity and nutrient 213 digestibility should be conducted because they may differ depending on the processing type, method, or 214 concentrations of the microminerals.

215 As intestinal permeability increases due to weaning stress, potentially pathogenic bacteria can penetrate and cause not only intestinal inflammation but also systemic inflammatory responses [42]. Changes in the WBC 216 217 count, indicative of systemic inflammation, are associated with alterations in the levels of cytokines involved in maintaining immunity and homeostasis [46,47]. Serum TNF-a is one of the pro-inflammatory cytokines, 218 used as a potential indicator of inflammatory reactions and damaging the mucosal barrier system [48]. Changes 219 220 in these parameters regulate the systemic immune responses against infections or diseases caused by weaning 221 stress. Previous studies reported that supplementation of CuSO<sub>4</sub> and ZnO reduced and downregulated the 222 concentrations and mRNA levels of inflammatory cytokines including TNF- $\alpha$  in the intestinal mucosa of weaned pigs [24,37]. The current study further demonstrated that dietary coated CuSO<sub>4</sub> and ZnO alleviated 223 224 systemic immune responses caused by weaning stress through the reduction of WBC and serum TNF- $\alpha$  levels. 225 Serum IgA, IgM, and IgG levels, which are the major components of humoral immunity, are reduced by 226 weaning stress and immature immunity of piglets [49]. In this study, dietary coated CuSO<sub>4</sub> and ZnO improved 227 serum IgG levels of weaned pigs, consistent with the results of previous studies [17,50]. IgG is a type of 228 antibody that leads to control the infection via binding many pathogens such as bacteria and viruses by immune 229 cells such as macrophages. Furthermore, it has been reported that free Cu and Zn ions possess antimicrobial 230 properties against *E.coli* in the small intestine [51]. Although the specific mechanisms of microminerals 231 activities are not entirely elucidated yet, it is widely accepted that these metallic ions degrade bacterial cell 232 membranes by disrupting the integrity of bacterial cell membranes, inducing cell death. In addition, ions 233 increase the release of reactive oxygen species within microorganisms, leading to pathogen destruction [52]. 234 Collectively, dietary supplementation with coated Cu and Zn modulates immune responses by preventing the 235 breakdown of the intestinal barrier and overproduction of pro-inflammatory cytokines.

236 Many studies have reported that dietary supplementation of pharmacological concentration of Cu and Zn 237 improves intestinal microorganisms by increasing the number of beneficial bacteria and reducing pathogenic 238 microbial composition [53,54]. The diversity and composition of the intestinal microbiota in pigs are 239 considerably affected by health condition and the digestion of nutrients compositions through physiological 240 functions [55]. In our study, the Shannon index tended to increase with coated CuSO<sub>4</sub> and ZnO compared to 241 that with the standard dosage of ZnO, indicating greater diversity in the fecal microbiota of piglets. Shen et al. 242 [24] showed that a pharmacological dosage of ZnO reduced the richness of microbial populations in the 243 jejunum and feces of weaning pigs, which was consistent with the results of our study. In general, diversity is 244 often associated with the presence of beneficial bacteria that can counteract pathogens [56]. However, the relative abundance of bacteria among dietary treatments was compared through taxonomic classification for a 245 246 more precise interpretation. At the phylum level, the Firmicutes and Bacteroidetes accounted for approximately 247 90% of all treatments in the fecal microbiomes of weaned pigs. Among the dietary treatments, dietary T2 had the highest proportion of Firmicutes. Since Lactobacillus and Clostridium were dominant genera within 248 249 Firmicutes, we determined that the overall portion of Lactobacillus (22.4%), Limosilactobacilus (24.78%), and 250 Clostridium sensu stricto (10.23%) in the T2 was higher than in the other treatments. At the genus level, the 251 present study showed that pigs fed dietary T2 had a higher relative abundance of genus Limosilactobacilus 252 compared with T1. Limosilactobacilus is a genus of lactic acid bacteria that recently split from Lactobacillus 253 and includes the species Limosilactobacilus reuteri, which is a microorganism with properties that promotes intestinal health and is widely used as a probiotic strain [54,57]. Furthermore, the relative abundance of fecal 254 255 microbiota increased Agathobacter and decreased Terrisporobacter in dietary T3 compared with that in CON 256 and dietary T1. Agathobacter is a beneficial bacterium that contributes to short-chain fatty acid production, 257 particularly butyrate, and is positively correlated with overall gut health in humans through metabolic 258 interactions of the gut microbiota [58]. Furthermore, Terrisporobacter shows a positive correlation with 259 increased serum markers such as endotoxin and  $TNF-\alpha$ , which promote oxidative stress, inflammation, and 260 malnutrition of gut microbiota in weaned pigs [59,60]. Therefore, the higher relative abundance of 261 Limosilactobacilus and Agathobacter may have contributed to the suppression of Terrisporobacter and 262 stabilization of the intestinal environment, thereby enhancing the growth performance of pigs fed dietary coated 263 CuSO<sub>4</sub> and ZnO than those fed CON and standard ZnO diets.

265	CONCLUSION
266	Our study demonstrated that dietary coated CuSO4 and ZnO supplementation in a nursery diet had
267	beneficial effects on growth performance and modulation of immune response and gut microbiota in weaned pigs.
268	These results indicate that improvement in growth performance and immune response may be associated with
269	changes in the fecal microbiota composition compared with CON group. In conclusion, dietary coated CuSO4 and
270	ZnO have positive effects in weaned pigs and represent potential alternatives to high levels of ZnO diets.
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# 277 **References**

- Rhouma M, Fairbrother JM, Beaudry F, Letellier A. Post weaning diarrhea in pigs: Risk factors and noncolistin-based control strategies. Acta Vet Scand. BioMed Central Ltd.; 2017
- Lallès JP, Boudry G, Favier C, Le Floc'h N, Luron I, Montagne L, et al. Gut function and dysfunction in young pigs: Physiology. Animal Res. 2004;53:301-16
- Campbell JM, Crenshaw JD, Polo J. The biological stress of early weaned piglets. J Anim Sci Biotechnol.
   2013;4:19
- Moeser AJ, Klok C Vander, Ryan KA, Wooten JG, Little D, Cook VL, et al. Stress signaling pathways activated by weaning mediate intestinal dysfunction in the pig. Am J Physiol Gastrointest liver Physiol. 2007;292(1):G173-181
- 5. Girard M, Bee G. Invited review: Tannins as a potential alternative to antibiotics to prevent coliform diarrhea
   in weaned pigs. Anim. Cambridge University Press; 2020;14;95–107
- Stein HH, Kil DY. Reduced use of antibiotic growth promoters in diets fed to weanling pigs: Dietary tools, part 2. Anim Biotechnol. 2006;17:217–31
- Yazdankhah S, Rudi K, Bernhoft A. Zinc and copper in animal feed development of resistance and coresistance to antimicrobial agents in bacteria of animal origin. Microb Ecol Health Dis. 2014. doi: 10.3402/mehd.v25.25862
- Hedemann MS, Jensen BB, Poulsen HD. Influence of dietary zinc and copper on digestive enzyme activity and intestinal morphology in weaned pigs. J Anim Sci. 2006;84:3310–20
- Bonetti A, Tugnoli B, Piva A, Grilli E. Towards zero zinc oxide: Feeding strategies to manage post-weaning diarrhea in piglets. Animals. MDPI AG; 2021;11:1-24
- Wen Y, Li R, Piao X, Lin G, He P. Different copper sources and levels affect growth performance, copper content, carcass characteristics, intestinal microorganism and metabolism of finishing pigs. Anim Nutr. 2022;8:321–30
- Liu Y, Espinosa CD, Abelilla JJ, Casas GA, Lagos LV, Lee SA, et al. Non-antibiotic feed additives in diets
   for pigs: A review. Anim Nutr. 2018;4(2):113-25
- Hu C, Song J, Li Y, Luan Z, Zhu K. Diosmectite-zinc oxide composite improves intestinal barrier function, modulates expression of pro-inflammatory cytokines and tight junction protein in early weaned pigs. British Journal of Nutrition. 2013;110:681–8
- Zhu C, Lv H, Chen Z, Wang L, Wu X, Chen Z, et al. Dietary Zinc Oxide Modulates Antioxidant Capacity,
   Small Intestine Development, and Jejunal Gene Expression in Weaned Piglets. Biol Trace Elem Res.

- 308 2017;175:331-8
- Bikker P, Jongbloed AW, Van Baal J. Dose-dependent effects of copper supplementation of nursery diets on growth performance and fecal consistency in weaned pigs. J Anim Sci. 2016;94:181–6.
- Stahly TS, Monegue HJ. Effects of source and level of copper on performance and liver copper stores in weanling pigs. J Anim Sci. 1989;67:2996-3002
- Zhao J, Allee G, Gerlemann G, Ma L, Gracia MI, Parker D, et al. Effects of a chelated copper as growth
   promoter on performance and carcass traits in pigs. Asian-Australas J Anim Sci. 2014;27:965–73
- Sun YB, Xia T, Wu H, Zhang WJ, Zhu YH, Xue JX, et al. Effects of nano zinc oxide as an alternative to pharmacological dose of zinc oxide on growth performance, diarrhea, immune responses, and intestinal microflora profile in weaned piglets. Anim Feed Sci Technol. 2019;258:114312
- Lei XJ, Liu ZZ, Park JH, Kim IH. Novel zinc sources as antimicrobial growth promoters for monogastric animals: a review. J Anim Sci Technol. 2022;64:187
- Blavi L, Villagómez-Estrada S, Solà-Oriol D, Pérez JF. Exploring zinc deficiency using serum Zn levels:
   consequences and potential solutions in suckling pigs. J Anim Sci. 2024;102
- 322 20. Revision of the currently authorised maximum copper content in complete feed. EFSA Journal. 2016;14
- Jang I, Kwon C, Ha D, Jung D, Kang S, Park M, et al. Effects of a lipid-encapsulated zinc oxide supplement
   on growth performance and intestinal morphology and digestive enzyme activities in weanling pigs. J Anim
   Sci Technol. 2014;56:29
- Wilk M, Pecka-Kiełb E, Pastuszak J, Asghar MU, Mól L. Effects of Copper Sulfate and Encapsulated Copper
   Addition on In Vitro Rumen Fermentation and Methane Production. Agriculture (Switzerland). 2022;12:1943
- Liu Y, Ma YL, Zhao JM, Vazquez-Añón M, Stein HH. Digestibility and retention of zinc, copper, manganese,
   iron, calcium, and phosphorus in pigs fed diets containing inorganic or organic minerals 1. J Anim Sci.
   2014;92:3407–15
- Shen J, Chen Y, Wang Z, Zhou A, He M, Mao L, et al. Coated zinc oxide improves intestinal immunity
   function and regulates microbiota composition in weaned piglets. British Journal of Nutrition.
   2014;111:2123–34
- Song M, Kim B, Cho JH, Kyoung H, Park S, Cho JY, et al. Effects of dietary protease supplementation on
   growth rate, nutrient digestibility, and intestinal morphology of weaned pigs. J Anim Sci Technol.
   2022;64:462–70
- Park S, Choe J, Cho JH, Jang KB, Kyoung H, Park KI, et al. Determination of optimal energy system and
   level for growing pigs. J Anim Sci Technol. 2024;66:514–22

- 339 27. AOAC.2010. Official Methods of Analysis of AOAC International 962.09 2-4
- Stein HH, Sève B, Fuller MF, Moughan PJ, De Lange CFM. Invited review: Amino acid bioavailability and digestibility in pig feed ingredients: Terminology and application. J Anim Sci. 2007;85(1):172-80
- Nam J, Kyoung H, Kim JN, Cho JH, Kim Y, Ahn J, et al. Effects of dietary aluminosilicate on growth
   performance, frequency of diarrhea, and blood profiles of weaned pigs. J Anim Sci Technol.
   2024;jast.2024.e21
- 345 30. Keum GB, Kim ES, Cho J, Song M, Oh KK, Cho JH, et al. Analysis of antibiotic resistance genes in pig
   346 feces during the weaning transition using whole metagenome shotgun sequencing. J Anim Sci Technol.
   347 2023;65:175–82
- 348 31. Lee JH, Kim S, Kim ES, Keum GB, Doo H, Kwak J, et al. Comparative analysis of the pig gut microbiome
   349 associated with the pig growth performance. J Anim Sci Technol. 2023;65:856–64
- 350 32. Kim H, Cho JH, Cho JH, Song M, Shin H, Kim S, et al. Complete genome sequence of Escherichia coli
   351 K\_EC180, a bacterium producing Shiga-like toxin isolated from swine feces. J Anim Sci Technol.
   352 2021;63:461-4
- 353 33. Kim S, Kwak J, Song M, Cho J, Kim ES, Keum GB, et al. Effects of Lacticaseibacillus casei (Lactobacillus casei) and Saccharomyces cerevisiae mixture on growth performance, hematological parameters, immunological responses, and intestinal microbiome in weaned pigs. Front Vet Sci. 2023;10:1140718
- 34. Li W, Fu L, Niu B, Wu S, Wooley J. Ultrafast clustering algorithms for metagenomic sequence analysis. Brief
   Bioinform. 2012;13:656–68
- 358 35. Parks DH, Tyson GW, Hugenholtz P, Beiko RG. STAMP: Statistical analysis of taxonomic and functional
   profiles. Bioinformatics. 2014;30:3123–4
- 36. Schell TC, Kornegay ET. Zinc Concentration in Tissues and Performance of Weanling Pigs Fed
   Pharmacological Levels of Zinc from ZnO, Zn-Methionine, Zn-Lysine, or ZnSO4. J Anim Sci.
   1996;74:1584-93
- 363 37. Shelton NW, Tokach MD, Nelssen JL, Goodband RD, Dritz SS, Derouchey JM, et al. Effects of copper
  364 sulfate, tri-basic copper chloride, and zinc oxide on weanling pig performance. J Anim Sci. 2011;89:2440–
  365 51
- 366 38. Oh HJ, Kim MH, Lee JH, Kim YJ, An JW, Chang SY, et al. Effects of different inorganic: organic zinc ratios
   367 or combination of low crude protein diet and mixed feed additive in weaned piglet diets. J Anim Sci Technol.
   368 2022;64:23–37
- 369 39. Esparza Gonzalez BP, Fong RN, Gibson CJ, Fuentealba IC, George Cherian AM. Zinc Supplementation
   370 Decreases Hepatic Copper Accumulation in LEC Rat A Model of Wilson's Disease. Biol Trace Elem Res.
   371 2005;105:117-134

- 40. Cousins RJ. Absorption, Transport, and Hepatic Metabolism of Copper and Zinc: Special Reference to
   Metallothionein and Ceruloplasmin. Physiol Rev. 1985;65:238-309
- 41. Jeng SS, Chen YH. Association of Zinc with Anemia. Nutrients. 2022;14:4918
- 42. Moeser AJ, Pohl CS, Rajput M. Weaning stress and gastrointestinal barrier development: Implications for
   lifelong gut health in pigs. Anim Nutr. 2017;3(4):313-21
- 43. Oh HJ, Park YJ, Cho JH, Song MH, Gu BH, Yun W, et al. Changes in diarrhea score, nutrient digestibility,
   zinc utilization, intestinal immune profiles, and fecal microbiome in weaned piglets by different forms of
   zinc. Animals. 2021;11:1356
- 44. Lei XJ, Kim IH. Low dose of coated zinc oxide is as effective as pharmacological zinc oxide in promoting
   growth performance, reducing fecal scores, and improving nutrient digestibility and intestinal morphology
   in weaned pigs. Anim Feed Sci Technol. 2018;245:117–25
- 45. Kim M, Hosseindoust A, Choi Y, Lee J, Kim K, Kim T, et al. Effects of Hot-Melt Extruded Nano-Copper as
   an Alternative for the Pharmacological Dose of Copper Sulfate in Weanling Pigs. 2021;199:2925-35
- 46. Han Y, Zhan T, Tang C, Zhao Q, Dansou DM, Yu Y, et al. Effect of replacing in-feed antibiotic growth promoters with a combination of egg immunoglobulins and phytomolecules on the performance, serum immunity, and intestinal health of weaned pigs challenged with escherichia coli k88. Animals. 2021;11:1292
- Liu Y, Song M, Che TM, Almeida JAS, Lee JJ, Bravo D, et al. Dietary plant extracts alleviate diarrhea and alter immune responses of weaned pigs experimentally infected with a pathogenic. J Anim Sci. 2013;91:5294-306
- 48. Zhang L, Xu YQ, Liu HY, Lai T, Ma JL, Wang JF, et al. Evaluation of Lactobacillus rhamnosus GG using an
   Escherichia coli K88 model of piglet diarrhoea: Effects on diarrhoea incidence, faecal microflora and
   immune responses. Vet Microbiol. 2010;141:142–8
- Klobasa F, Butler JE, Werhahn E, Habe F. Maternal-neonatal immunoregulation in swine. II. Influence of
   multiparity on de novo immunoglobulin synthesis by piglets. Vet Immunol Immunopathol. 1986;11:149-59
- 396 50. Gonzales-Eguia A, Fu CM, Lu FY, Lien TF. Effects of nanocopper on copper availability and nutrients
   397 digestibility, growth performance and serum traits of piglets. Livest Sci. 2009;126:122–9
- Raja FNS, Worthington T, Martin RA. The antimicrobial efficacy of copper, cobalt, zinc and silver nanoparticles: alone and in combination. Biomedical Materials (Bristol). 2023;18:0405003
- 52. Duarte ME, Garavito-Duarte Y, Kim SW. Impacts of F18+ Escherichia coli on Intestinal Health of Nursery
   Pigs and Dietary Interventions. Animals. 2023;13:2791
- 402 53. Zheng J, Wittouck S, Salvetti E, Franz CMAP, Harris HMB, Mattarelli P, et al. A taxonomic note on the 403 genus Lactobacillus: Description of 23 novel genera, emended description of the genus Lactobacillus

- beijerinck 1901, and union of Lactobacillaceae and Leuconostocaceae. Int J Syst Evol Microbiol.
   2020;70:2782-858
- 406 54. Abuqwider J, Altamimi M, Mauriello G. Limosilactobacillus reuteri in Health and Disease. Microorganisms.
   407 2022;10:522
- 408 55. Rist VTS, Weiss E, Eklund M, Mosenthin R. Impact of dietary protein on microbiota composition and activity
   409 in the gastrointestinal tract of piglets in relation to gut health: A review. Animal. 2013;7:1067–78
- 56. Schokker D, Kar SK, Willems E, Bossers A, Dekker RA, Jansman AJM. Dietary supplementation of zinc oxide modulates intestinal functionality during the post-weaning period in clinically healthy piglets. J Anim Sci Biotechnol. 2023;14
- 57. Duar RM, Frese SA, Lin XB, Fernando SC, Burkey TE, Tasseva G, et al. Experimental evaluation of host adaptation of Lactobacillus reuteri to different vertebrate species. Appl Environ Microbiol. 2017;83
- 415 58. Granado-Serrano AB, Martín-Garí M, Sánchez V, Riart Solans M, Berdún R, Ludwig IA, et al. Faecal bacterial and short-chain fatty acids signature in hypercholesterolemia. Sci Rep. 2019;9:1772
- 417 59. Cai C, Zhang Z, Morales M, Wang Y, Khafipour E, Friel J. Feeding practice influences gut microbiome composition in very low birth weight preterm infants and the association with oxidative stress: A prospective cohort study. Free Radic Biol Med. 2019;142:146–54
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425 **Table 1.** Composition of basal diet for weaned pigs (as-fed basis)

Item	Basal diet
Ingredient, %	
Corn	49.86
Soybean meal, 44%	25.00
Whey powder	12.50
Soy protein concentrate	6.25
Soybean oil	3.00
Limestone	1.14
Monocalcium phosphate	1.05
Vitamin premix <sup>1)</sup>	0.20
Mineral premix <sup>2)</sup>	0.20
L-Lysine·HCl	0.45
DL-Methionine	0.16
L-Threonine	0.13
L-Valine	0.06
Total	100.00
Calculated energy and nutrient	
Metabolizable energy, kcal/kg	3,465
Crude protein, %	21.26
Calcium, %	0.81
Phosphorous, %	0.65
Lysine, %	1.53
Methionine, %	0.47
Threonine, %	0.95
Tryptophan, %	0.25

<sup>426 &</sup>lt;sup>1</sup>Vitamin premix provided the following quantities of vitamin per kilogram of complete diet: vitamin A, 12,000

427 IU; vitamin D<sub>3</sub>, 2,500 IU; vitamin E, 30 IU; vitamin K<sub>3</sub>, 3 mg; D-pantothenic acid, 15 mg; nicotinic acid, 40 mg;
428 choline, 400 mg; and vitamin B<sub>12</sub>, 12 μg.

429 <sup>2)</sup>Mineral premix provided the following quantities of mineral per kilogram of complete diet: Fe, 90 mg from

430 iron sulfate; Cu, 8.8 mg from copper sulfate; Zn, 100 mg from zinc oxide; Mn, 54 mg from manganese oxide; I,

431 0.35 mg from potassium iodide; Se, 0.30 mg from sodium selenite.

433 **Table 2.** Effects of dietary coated CuSO<sub>4</sub> and ZnO on growth performance of weaned pigs<sup>1)</sup>

Item <sup>2)</sup>	CON	T1	T2	Т3	SEM	<i>p</i> -value
Day 1 to 14						
Initial BW, kg	7.30	7.31	7.29	7.27	0.31	1.000
Final BW, kg	10.73	11.27	11.75	11.87	0.44	0.503
ADG, g/d	245 <sup>a</sup>	283 <sup>ab</sup>	319 <sup>b</sup>	328 <sup>b</sup>	20.72	0.040
ADFI, g/d	437	429	415	429	26.52	0.994
G:F, g/g	0.561ª	0.659 <sup>ab</sup>	$0.768^{b}$	0.766 <sup>b</sup>	0.042	0.021
Day 15 to 42						
Initial BW, kg	10.73	11.27	11.75	11.87	0.44	0.503
Final BW, kg	21.49 <sup>a</sup>	22.23 <sup>ab</sup>	22.85 <sup>b</sup>	22.98 <sup>b</sup>	0.31	0.030
ADG, g/d	384	391	396	397	13.02	0.925
ADFI, g/d	956	951	963	965	29.55	0.999
G:F, g/g	0.402	0.412	0.412	0.411	0.020	0.993
Day 1 to 42						
Initial BW, kg	7.30	7.31	7.29	7.27	0.31	1.000
Final BW, kg	21.49 <sup>a</sup>	22.23 <sup>ab</sup>	22.85 <sup>b</sup>	22.98 <sup>b</sup>	0.31	0.030
ADG, g/d	338 <sup>a</sup>	355 <sup>ab</sup>	371 <sup>b</sup>	374 <sup>b</sup>	7.28	0.020
ADFI, g/d	783	777	780	786	24.78	0.999
G:F, g/g	0.431	0.457	0.475	0.476	0.016	0.419
Frequency of diarrhea <sup>3</sup> , %	13.87	9.28	10.14	9.92		0.051

434 <sup>1)</sup>Each value is the mean of 6 replicates (4 pigs/pen).

 $^{2)}$  CON = basal weaner diet based on corn and soybean meal, T1 = CON + 2,500 ppm standard ZnO, T2 = CON

436 + 100 mg/kg dietary coated CuSO<sub>4</sub> and 100 mg/kg dietary coated ZnO, T3 = CON + 200 mg/kg dietary coated

437 CuSO<sub>4</sub> and 200 mg/kg dietary coated ZnO, BW = body weight, ADG = average daily gain, ADFI = average daily

438 feed intake, G:F = gain to feed ratio.

 $^{3)}$  Frequency of diarrhea for the first 2 weeks after weaning (%) = (number of diarrhea score of 4 or higher / number

440 of pen days)  $\times$  100. Data was analyzed using the Chi-square test.

441 <sup>a,b</sup>Means in the same row with different superscripts are different (p < 0.05)

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**Table 3.** Effects of dietary coated CuSO<sub>4</sub> and ZnO on apparent total tract digestibility of weaned pigs<sup>1)</sup>

Item <sup>2)</sup>	CON	T1	T2	T3	SEM	<i>p</i> -value
DM, %	70.68	75.27	80.17	81.15	5.67	0.324
Energy, %	75.34	77.83	79.74	84.33	5.21	0.414
CP, %	69.84	73.33	76.40	82.59	6.48	0.542

<sup>1)</sup>Each value is the mean of 6 replicates (1 pig/pen).
<sup>2)</sup>CON = basal weaner diet based on corn and soybean meal, T1 = CON + 2,500 ppm standard ZnO, T2 = CON + 100 mg/kg dietary coated CuSO<sub>4</sub> and 100 mg/kg dietary coated ZnO, T3 = CON + 200 mg/kg dietary coated 447 CuSO<sub>4</sub> and 200 mg/kg dietary coated ZnO, DM= dry matter, CP= crude protein.



Item <sup>2)</sup>	CON	T1	T2	Т3	SEM	<i>p</i> -valu
WBC, ×10 <sup>3</sup> /µL						
Day 1	13.88	13.56	12.18	14.88	2.48	0.474
Day 7	23.08	19.76	18.70	20.42	1.31	0.074
Day 14	24.70	23.32	19.20	19.14	2.53	0.459
RBC, ×10 <sup>6</sup> /µL						
Day 1	4.70	4.94	4.54	4.65	0.18	0.355
Day 7	6.00	6.33	6.19	6.10	0.41	0.984
Day 14	6.45	6.50	6.70	6.74	0.32	0.325
НСТ, %						
Day 1	25.08	26.02	25.02	24.92	0.87	0.885
Day 7	30.66	29.74	29.66	29.54	2.05	0.985
Day 14	35.56	30.30	29.26	29.56	1.85	0.084
TNF-α, pg/mL						
Day 1	141.21	131.53	125.35	121.35	30.25	0.506
Day 7	108.81	78.57	77.79	78.97	10.08	0.095
Day 14	104.99	84.44	77.27	81.43	38.91	0.550
Cortisol, ng/mL						
Day 1	115.05	118.74	107.32	105.83	18.64	0.691
Day 7	122.98	81.18	83.77	90.17	13.84	0.063
Day 14	105.50	98.64	92.11	94.88	17.39	0.330
IgG, mg/mL						
Day 1	5.37	4.86	4.91	5.24	0.71	0.721
Day 7	3.72	3.84	3.77	3.94	1.01	0.650
Day 14	3.86	4.55	4.79	4.75	0.31	0.076
IgM, mg/mL	Y					
Day 1	1.36	1.57	1.15	1.43	0.27	0.413
Day 7	1.35	1.37	1.44	1.56	0.17	0.361
Day 14	1.23	1.62	1.60	1.51	0.13	0.067
IgA, mg/mL						
Day 1	0.23	0.36	0.25	0.26	0.08	0.385
Day 7	0.32	0.26	0.28	0.30	0.11	0.857
Euj						

450 **Table 4.** Effects of dietary coated CuSO<sub>4</sub> and ZnO on blood profiles and immune responses of weaned pigs<sup>1)</sup>

451 <sup>1)</sup>Each value is the mean of 6 replicates (1 pig/pen).

<sup>2)</sup> CON = basal weaner diet based on corn and soybean meal, T1 = CON + 2,500 ppm standard ZnO, T2 = CON + 100 mg/kg dietary coated CuSO<sub>4</sub> and 100 mg/kg dietary coated ZnO, T3 = CON + 200 mg/kg dietary coated CuSO<sub>4</sub> and 200 mg/kg dietary coated ZnO, WBC = white blood cell, RBC = red blood cell, HCT = hematocrit, TNF- $\alpha$  = tumor necrosis factor-alpha, IgG = immunoglobulin G, IgM = immunoglobulin M, IgA = immunoglobulin A.



Fig. 1. Effects of dietary coated CuSO<sub>4</sub> and ZnO on alpha diversity of fecal microbiota of
weaned pigs (n = 3). Alpha diversity indices were (a) observed OTUs, (b) Chao1, (c) Shannon,
and (d) Simpson. Statistical difference was performed using the analysis of ANOVA. CON =
basal weaner diet based on corn and soybean meal, T1 = CON + 2,500 ppm standard ZnO, T2
= CON + 100 mg/kg dietary coated CuSO<sub>4</sub> and 100 mg/kg dietary coated ZnO, T3 = CON +
200 mg/kg dietary coated CuSO<sub>4</sub> and 200 mg/kg dietary coated ZnO.



Fig. 2. Effects of dietary coated CuSO<sub>4</sub> and ZnO on beta diversity of fecal microbiota of weaned pigs (n = 3). Principal coordinated analysis based on (a) unweighted and (b) weighted Unifrac distances. The ANOSIM test was used for statistically significant distances. T1 = CON + 2,500 ppm standard ZnO, T2 = CON + 100 mg/kg dietary coated CuSO<sub>4</sub> and 100 mg/kg dietary coated ZnO, T3 = CON + 200 mg/kg dietary coated CuSO<sub>4</sub> and 200 mg/kg dietary coated ZnO.



Fig. 3. Effects of dietary coated CuSO<sub>4</sub> and ZnO on relative taxonomic abundance at the phylum level of fecal microbiota of weaned pigs (n = 3). CON = basal weaner diet based on corn and soybean meal, T1 = CON + 2,500 ppm standard ZnO, T2 = CON + 100 mg/kg dietary coated CuSO<sub>4</sub> and 100 mg/kg dietary coated ZnO, T3 = CON + 200 mg/kg dietary coated CuSO<sub>4</sub> and 200 mg/kg dietary coated ZnO.

**(a)** 







**Fig. 4.** Effects of dietary coated CuSO<sub>4</sub> and ZnO on relative taxonomic abundance at the genus level (a) and relative abundances of *Agathobacter, Terrisporobacter,* and *Limosilactobacilus* (b) of fecal microbiota of weaned pigs (n = 3). CON = basal weaner diet based on corn and soybean meal, T1 = CON + 2,500 ppm standard ZnO, T2 = CON + 100 mg/kg dietary coated CuSO<sub>4</sub> and 100 mg/kg dietary coated ZnO, T3 = CON + 200 mg/kg dietary coated CuSO<sub>4</sub> and 200 mg/kg of dietary coated ZnO.

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