# JAST (Journal of Animal Science and Technology) TITLE PAGE Upload this completed form to website with submission

ARTICLE INFORMATION	Fill in information in each box below
Article Type	Research article
Article Title (within 20 words without	Diversity of MHC-B SNP haplotypes in the Vietnamese
abbreviations) Running Title (within 10 words)	Ri chicken "MHC-B SNP haplotypes of the Ri chicken"
Running The (within 10 words)	мнс-в SNP haplotypes of the KI chicken
Author	Seonju Nam1, Prabuddha Manjula2, Jaewon Kim1, Minjun Kim1, Eunjin Cho3, Trisha Nicole Agulto1, Yeong Ho Hong4, Jun Heon Lee1, 3
Affiliation	<ol> <li>Division of Animal and Dairy Science, Chungnam National University, Daejeon 34134, Korea.</li> <li>Department of Animal Science, Uva Wellassa University, Badulla 90000, Sri Lanka.</li> <li>Department of Bio-AI Convergence, Chungnam National University, Daejeon 34134, Korea.</li> <li>Department of Animal Science and Technology, Chung-Ang University, Anseong 17546, Korea.</li> </ol>
ORCID (for more information, please visit https://orcid.org)	Seonju Nam (https://orcid.org/0009-0002-6731-121X) Prabuddha Manjula (https://orcid.org/0000-0001-8074- 8323)
	Jaewon Kim (https://orcid.org/0009-0006-0445-3025) Minjun Kim (https://orcid.org/0000-0002-8173-8431) Eunjin Cho (https://orcid.org/0000-0003-4800-1603) Trisha Nicole Agulto (https://orcid.org/0009-0006-3168- 8110) Yeong Ho Hong (https://orcid.org/0000-0002-4510- 7851) Jun Heon Lee (https://orcid.org/0000-0003-3996-9209)
Competing interests	No potential conflict of interest relevant to this article was reported.
<b>Funding sources</b> State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available.	This research was funded by a grant from the National Research Foundation, Republic of Korea (grant number 2022R1F1A1064025).
Acknowledgements	The authors thank the Department of Biochemistry and Immunology Laboratory of the NIVR-Vietnam National Veterinary Research for conducting the animal experiments described in this study.
Availability of data and material	Upon reasonable request, the datasets of this study can be available from the corresponding author.
Authors' contributions Please specify the authors' role using this form.	Conceptualization: Lee JH Data curation: Nam S, Kim J, Hong YH Formal analysis: Manjula P, Kim J, Kim M, Cho E Methodology: Nam S, Kim J Software: Kim J, Kim M Validation: Manjula P, Kim M, Cho E Investigation: Nam S, Manjula P, Agulto TN, Hong YH

	Writing - original draft: Nam S				
	Writing - review & editing: Nam S, Manjula P, Kim J				
	Kim M, Cho E, Agulto TN, Hong YH, Lee JH				
Ethics approval and consent to participate	The care and experimental use of the chickens were				
	approved by the Ministry of Agriculture and Rural				
	Development of Vietnam (TCVN 8402: 2010/TCVN				
	8400-26:2014).				

# CORRESPONDING AUTHOR CONTACT INFORMATION

For the corresponding author (responsible for correspondence, proofreading, and	Fill in information in each box below			
reprints)				
First name, middle initial, last name	Jun Heon Lee			
Email address – this is where your proofs will	junheon@cnu.ac.kr			
be sent				
Secondary Email address	seonju7740@yonsei.ac.kr			
Address	213, KTnG, Chungnam National University, 99, Daehak-			
	ro, Yuseong-gu, Daejeon, Republic of Korea			
Cell phone number	+82-10-5172-0816			
Office phone number	+82-42-821-7031			
Fax number	+82-42-825-9754			

#### 1 Abstract

2 Avian influenza (AI) is a serious global threat to poultry and public safety, although some 3 native chicken varieties show resilience, such as the Ri chicken in Vietnam. Major histocompatibility complex B (MHC-B), a critical component of the chicken immune system, 4 5 has been shown to influence disease resistance. This study examined the MHC-B haplotype diversity in a Ri chicken population that is sensitive to avian influenza. Ri chickens were 6 7 genotyped for MHC-B single nucleotide polymorphisms (SNPs) using the Kompetitive Allele 8 Specific Polymerase Chain Reaction (KASP). Statistical tests revealed no significant 9 differences in allele frequencies of the SNPs between resistant (R) and susceptible (S) groups. Haplotype analysis identified 32 unique haplotypes, with only one shared haplotype between 10 11 the R and S groups. However, a phylogenetic analysis did not find distinct clustering of MHC-12 B alleles of the Ri chicken groups. Further research with a larger sample size is recommended 13 to establish representative group-specific haplotypes and enhance our comprehension of the intricate genetic mechanisms underlying disease resistance in poultry. The implications of this 14 15 research extend to improving disease resistance strategies and guiding selective breeding 16 practices in the poultry industry.

17

Keywords: Avian influenza, Ri chicken, MHC-B variability, SNP genotyping 18

### Introduction

21 Avian flu, also known as avian influenza (AI), is a concern for both the global poultry industry and 22 public health. Identifying the responsible genes and related pathways is of paramount importance to 23 develop resistant chicken strains that can withstand the highly pathogenic avian influenza virus (HPAIV) 24 therefore minimizing the negative impact on poultry production. Certain gene families and alleles 25 associated with disease resilience in avian species are used as markers for selection and breeding [1]. 26 Alleles of the major histocompatibility complex (MHC) genes are often associated with disease 27 resistance and susceptibility in chickens and mammals. The MHC-B region is highly polymorphic and 28 has complex genetic loci that contain clusters of genes responsible for the immune response and immune 29 recognition in chickens [2]. It is responsible for the adaptive and innate immune responses in chickens 30 [3]. Variations in MHC-B affect specific disease resistance to several highly pathogenic viral and 31 bacterial diseases, as well as internal and external parasites in poultry [4, 5]. Previous studies have 32 reported that the MHC-B21 haplotype is associated with resistance to H5N1 virus infection, with a 100% 33 survival rate. By contrast, chickens with the MHC-B13 haplotype showed 100% mortality during 34 HPAIV outbreaks in Thailand [6]. The MHC-B21 haplotype is also associated with lower tumor-related 35 mortality due to Marek's herpes virus infection than other haplotypes [7]. Matsuu et al. [8] reported that 36 despite the presence of significant BF1 and BF2 allele variations in Thai native chickens, none of the 37 alleles, particularly BF1/BF2 alleles that are homologous to the MHC-B21 haplotype, were significantly associated with sensitivity to HPAIV infection. 38 39 A set of single nucleotide polymorphisms (SNPs) in the chicken MHC-B region, developed in the study 40 by Fulton et al. [3], has been used to identify haplotypes of the MHC-B region in various chicken breeds 41 around the world [3, 9, 10, 11, 12]. The panel consisted of 101 SNPs, but the set of 90 SNPs that cover 42 the region between MHCJ06 and MHC178 were utilized for defining haplotype [3]. SNP-based 43 techniques have favorable haplotype discriminating power, which is helpful before conducting a high-44 resolution haplotyping analysis. 45 In the present study, MHC-B SNP diversity was carried out with the genetically resistant and susceptible 46 lines of the Vietnamese Ri chicken group. A total of 20 Ri chickens, 10 each line as described in Lee et 47 al. [1], were used to examine the MHC-B haplotype diversity using an MHC-B SNP panel and their

48 distribution in relation to the avian influenza resilience of Ri chickens.

- 49
- 50

51

# **Materials and Methods**

52 Genomic DNA amplification from Ri chickens

53 The Vietnamese Ri chicken population used in the current study is also described in another source [1].

A total of 20 Ri chicken samples (i.e., HPAIV-resistant and susceptible lines) obtained from the Poultry
 Research Centre of the National Institute of Animal Science (Hanoi, Vietnam) were used. Specifically,

56 10 individuals were selected for each of the resistant and susceptible groups, designated as R and S

57 groups, respectively. These chickens were chosen based on the genotypes for the *BF2* and *MX1* alleles 58 to identify genetically resistant or susceptible individuals. Genomic DNA extracted from the said 59 chicken population was obtained from the previous research group [1] and diluted to a final DNA 60 concentration of 5 ng/µl before KASP genotyping.

61

### 62 MHC-B SNP genotyping

To further understand the genetic diversity within MHC-B, we genotyped 20 individuals using an MHC-B SNP panel described in the study by Fulton et al. [3]. The panel consists of 90 previously identified SNPs and is subjected to a fluorescence-based genotyping method called Kompetitive Allele Specific Polymerase Chain Reaction (KASP) [3].

67

### 68 Statistical analysis

To test statistically significant differences in allele frequencies for the 89 SNP markers between the R and S groups, we used the chi-square test [13]. Statistical tests were conducted individually for each SNP marker. The applied statistical formula is as follows.

72 
$$x_c^2 = \sum \frac{(O_i - E_i)^2}{E_i}$$

73 In this context,  $x^2$  stands for the chi-square statistic and c denotes the degrees of freedom, which was 74 consistently set to 1 for all markers. *O* represents the observed allele frequency values while *E* stands 75 for the expected values and the variable *i* refers to each specific SNP marker.

- 76 The statistical analysis was carried out using R software version 4.3 and the significance of the results
- was determined based on a *p*-value threshold of 0.05 for each SNP marker's allele frequencycalculations and testing.

79

#### 80 MHC-B SNP haplotype identification and phylogenetic analysis

81 In contrast to previous studies of BSNP haplotypes in native chicken populations [9, 11], we excluded 82 the "MHC065" SNP from BSNP haplotypes therefore utilizing only 89 SNPs. Because the MHC065 83 was not genotyped in the current population. MHC-B haplotypes were then inferred from the genotypes 84 obtained from the 89 SNPs in all 20 Ri chickens along with homozygous samples with known MHC-B 85 SNP genotypes from the previous study of Manjula et al. [11]. Haplotype analysis was implemented 86 using PHASE 2.1 software using the -MS model with no recombination to iterate all possible haplotypes. 87 The haplotypes were named after their origin within the Ri chicken population followed by haplotype 88 number (e.g., "Ri\_S/R\_Hap01"). 89 To distinguish the haplotype diversity among Ri chickens and other native chicken breeds, a

90 phylogenetic tree was constructed using all defined haplotypes based on the 89 SNPs and compared to

91 global chicken populations including the Korean native chicken, Sri Lankan chicken, Bangladesh

92 chicken, and MHC-B standard haplotypes defined using the same set of SNPs in previous studies [3,

93	11, 12, 14]. The analysis was conducted using the Bayes r option, utilizing sequence differences
94	between haplotypes, in Geneious Prime software v2023.1.2.
95	
96	
97	<b>Results and Discussion</b>
98	Differences of SNP allele frequencies in Ri chicken
99	Table 1 summarizes the differences in allele frequency in the Ri chicken population, confirmed by
100	KASP genotyping, presenting the top 10 SNPs with chi-square test statistics. No significant differences
101	in allele frequencies were observed between the R and S groups. The SNP that had the most notable
102	frequency difference between the groups was MHC127, with a fixed frequency of allele 1 at 1.00 in the
103	R group and an observed frequency of 0.50 in the S group. The chi-square statistic for this SNP was
104	0.667 and the calculated <i>p</i> -value was 0.414.
105	
106	BSNP haplotypes in the Ri chicken population
107	A total of 32 BSNP haplotypes (shown in Figure 1) were identified from the 20 Ri chickens tested. The
108	number of haplotypes present in each group is summarized in Table 2. Haplotypes prefixed with "Ri_R
109	or Ri_S" were exclusive to the R and S groups, respectively. These haplotypes had varying frequencies,
110	with some occurring only once (e.g., Ri_R_Hap01) and others being more prevalent.
111	Of the 32 haplotypes, Ri_RS_Hap32 was present in both the R and S groups with a frequency of 1 in
112	both groups indicating a shared genetic characteristic. Within the R group, the most prevalent haplotype
113	was Ri_R_Hap09, observed in three individuals, followed by Ri_R_Hap13 and Ri_R_Hap14, each
114	present in two individuals. In the S group, haplotype Ri_S_Hap21 was the most frequent, followed by
115	haplotype <i>Ri_S_Hap17</i> .
116	However, the relatively low frequency of group-specific haplotypes makes it challenging to determine
117	the presence or absence of haplotype sharing at the group level as each group-specific haplotype was
118	found only in a few individuals. Similar results were also observed in the native Bangladesh chicken
119	and red jungle fowl with many alleles appearing only once or twice in the population [14]. Some of the
120	new haplotypes differed from each other at only one or two loci. The identification of recombinant
121	haplotypes is unlikely because of the limited population size.
122	In comparison to other similar studies [11, 12], the limited sample size in the current study was not
123	necessarily due to a shortage of individuals but rather a reflection of the substantial diversity in the
124	MHC-B region within the Ri chicken population. Given that all the animals are heterozygous, it is
125	evident that the Ri chicken population is very diverse in the MHC-B region. Additional research on a
126	larger number of Ri chickens is necessary to establish representative group-specific haplotypes.
127	
128	Phylogenetic tree based on BSNP haplotypes in the Ri chicken population
129	Figure 2 shows the constructed phylogenetic tree of the 32 haplotypes in the Ri chicken population. The

130 tree is broadly divided into two major clades, with intermixed group specificities within these clades.

132	other global chicken haplotypes. Although no study has been conducted on the genetic distance between
133	Ri chicken and Bangladesh chicken breeds, it can be inferred from these results that the characteristics
134	of the MHC-B region are somehow similar between two populations. Consequently, the phylogenetic
135	tree does not reveal distinct similarities in BSNP haplotypes specific to the R and S groups. This concurs
136	with the finding that no SNPs had significant differences in allele frequencies between the groups.
137	This study analyzed chicken populations using SNP haplotypes that excluded the MHC065 marker.
138	One of the reasons why SNP genotyping failed may be due to the highly polymorphic nature of the
139	MHC region. In the future, when it is possible to confirm the genotype, updated research in
140	the region could be reported.
141	
142	Conclusions
143	The present study showed that none of the MHC-B SNP alleles were significantly associated with avian
144	influenza resistance in the Ri chicken population. Unique MHC-B haplotypes were also discovered
145	wherein phylogenetic analysis of these haplotypes showed a closer relationship between the Ri and
146	Bangladesh chickens than other chicken populations. However, further investigations are still needed
147	to evaluate the relationships of the Ri chicken with other chicken populations.
148	
149	Funding source
150	This research was funded by a grant from the National Research Foundation, Republic of Korea (grant
151	number 2022R1F1A1064025).
152	
153	Acknowledgements
154	The authors thank the Department of Biochemistry and Immunology Laboratory of the NIVR-Vietnam
154	National Veterinary Research for conducting the animal experiments described in this study.
155	Ivational veterinary Research for conducting the annual experiments described in this study.
157	
1.57	

Ri chicken clustered with the Bangladesh chicken regardless of the sub-group but separated from the

- 158 **References**159

  Lee J, Hong Y, Vu TH, Lee S, Heo J, Truong AD, et al. Influenza a Pathway Analysis of Highly Pathogenic Avian Influenza Virus (H5N1) Infection in Genetically Disparate Ri Chicken Lines. Vet. Immunol. Immunopathol. 2022;246:110404. https://doi.org/10.1016/j.vetimm.2022.110404.
- 162 2. Delany ME, Robinson CM, Goto RM, Miller MM. Architecture and organization of chicken
  163 microchromosome 16: order of the NOR, MHC-Y, and MHC B subregions. J Hered.
  164 2009;100:507-14. https://doi.org/10.1093/jhered/esp044.
- Fulton JE, McCarron AM, Lund AR, Pinegar KN, Wolc A, Chazara O, et al. A high-density SNP
  panel reveals extensive diversity, frequent recombination and multiple recombination hotspots
  within the chicken major histocompatibility complex B region between BG2 and CD1A1. Genet
  Sel Evol. 2016;48:1-15. https://doi.org/10.1186/s12711-015-0181-x.
- Quach CC, Fulton JE, Benson JD, Walker P, Auckland C, Lessard Carl. Major Histocompatibility
   Complex-B Haplotype and Ovarian Graft Response. Poult. Sci. 2023;102:102850.
   https://doi.org/10.1016/j.psj.2023.102850.
- Liu W, Miller MM, Lamont SJ. Association of MHC class I and class II gene polymorphisms with
  vaccine or challenge response to *Salmonella enteritidis* in young chicks.
  Immunogenetics. 2002;54:582-590. https://doi.org/10.1007/s00251-002-0495-z.
- Boonyanuway K, Thummabutra S, Sookmanee N, Vatchavalkhu V, Siripholvat V, Mitsuhashi T.
  Influence of MHC class II haplotypes on avian influenza traits in Thai indigenous chicken. J Poult
  Sci. 2006;43:120–125. https://doi.org/10.2141/jpsa.43.120.
- 178 7. Briles WE, Stone HA, Cole RK. Marek's disease: effects of B histocompatibility alloalleles in
  179 resistant and susceptible chicken lines. *Science*. 1977;195:193–
  180 195. https://doi.org/10.1126/science.831269.
- Matsuu A, Kobayashi T, Patchimasiri T, Shiina T, Suzuki S, Chaichoune K, et al. Pathogenicity of Genetically Similar, H5N1 Highly Pathogenic Avian Influenza Virus Strains in Chicken and the Differences in Sensitivity among Different Chicken Breeds. PLoS ONE. 2016;11:e0153649.
   https://doi.org/10.1371/journal.pone.0153649.
- 185 9. Fulton JE, Berres ME, Kantanen J, Honkatukia M. MHC-B variability within the Finnish
  186 Landrace chicken conservation program, Poult. Sci. 2017;96:3026-3030.
  187 https://doi.org/10.3382/ps/pex102.
- 188 10. Iglesias GM, Canet Z, Cantaro H, Miquel MC, Melo JE, Miller MM, et al. Mhc-B haplotypes in
  "Campero-Inta" chicken synthetic line, Poult. Sci. 2019;98:5281-5286.
  190 https://doi.org/10.3382/ps/pez431.
- Manjula P, Fulton JE, Seo D, Lee JH. Major Histocompatibility Complex B Variability in Korean Native Chicken Breeds. Poult. Sci. 2020;99:4704–4713. https://doi.org/10.1016/j.psj.2020.05.049.

- Manjula P, Fulton JE, Seo D, Lee JH. Comparison of major histocompatibility complex-B
   variability in Sri Lankan indigenous chickens with five global chicken populations using MHC-B
   SNP panel. Anim. Genet. 2021;52:824-833. https://doi.org/10.1111/age.13137.
- Pirhaji L, Kargar M, Sheari A, Poormohammadi H, Sadeghi M, Pezeshk H, et al. The Performances of the Chi-square Test and Complexity Measures for Signal Recognition in Biological Sequences.
   J. Theor. Biol. 2008;251:380–387. https://doi.org/10.1016/j.jtbi.2007.11.021.
- 14. Ediriweera TK, Manjula P, Kim J, Kim JH, Nam S, Kim M, et al., Identification of new major
   histocompatibility complex-B haplotypes in Bangladesh native chickens. Anim. Biosci. (Accepted
   and in process). https://doi.org/10.5713/ab.23.0295.

# **Tables and Figures**

CND	A 11 a 1 a	R group			S group		
SNP	Allele	Freq <sub>A1</sub>	Freq <sub>A2</sub>	Freq <sub>A1</sub>	Freq <sub>A2</sub>	$-X^2$	<i>p</i> -value
MHC127	G > A	1.00	0.00	0.50	0.50	0.667	0.414
MHC025	T > C	0.55	0.45	0.05	0.95	0.595	0.440
MHC114	A > G	0.60	0.40	0.15	0.85	0.432	0.511
MHC079	A > G	0.65	0.35	0.20	0.80	0.414	0.520
MHC008	A > G	0.60	0.40	0.95	0.05	0.351	0.554
MHC011	T > C	0.60	0.40	0.20	0.80	0.333	0.564
MHC056	A > G	0.55	0.45	0.90	0.10	0.307	0.580
MHC015	C > G	0.75	0.25	1.00	0.00	0.286	0.593
MHC060	G > T	0.55	0.45	0.20	0.80	0.261	0.609
MHC081	C > T	0.80	0.20	0.45	0.55	0.261	0.609

204 Table 1. Differences in allele frequency in single nucleotide polymorphisms between the R and S groups showing the top 10 SNPs with chi-square statistics. 205

SNP, single nucleotide polymorphism; Allele, allele 1 and allele 2; Freq<sub>A1</sub>, frequency of allele 1 in a 206

group;  $Freq_{A2}$ , frequency of allele 2 in a group; X<sup>2</sup>, calculated chi-square statistic; R group, resistant group to avian influenza; S group, susceptible group to avian influenza 207

208

209

Haplotype	Number of haplotypes		Specificity	Honlotyno -	Number of haplotypes		Specificity
	R group	S group	specificity	Haplotype –	R group	S group	specificity
Ri_R_Hap01	1	0	R group	Ri_S_Hap17	0	2	S group
Ri_R_Hap02	1	0	R group	Ri_S_Hap18	0	1	S group
Ri_R_Hap03	1	0	R group	Ri_S_Hap19	0	1	S group
Ri_R_Hap04	1	0	R group	Ri_S_Hap20	0	1	S group
Ri_R_Hap05	1	0	R group	Ri_S_Hap21	0	3	S group
Ri_R_Hap06	1	0	R group	Ri_S_Hap22	0	1	S group
Ri_R_Hap07	1	0	R group	Ri_S_Hap23	0	1	S group
Ri_R_Hap08	1	0	R group	Ri_S_Hap24	0	1	S group
Ri_R_Hap09	3	0	R group	Ri_S_Hap25	0	1	S group
Ri_R_Hap10	1	0	R group	Ri_S_Hap26	0	1	S group
Ri_R_Hap11	1	0	R group	Ri_S_Hap27	0	1	S group
Ri_R_Hap12	1	0	R group	Ri_S_Hap28	0	1	S group
Ri_R_Hap13	2	0	R group	Ri_S_Hap29	0	1	S group
Ri_R_Hap14	2	0	R group	Ri_S_Hap30	0	1	S group
Ri_R_Hap15	1	0	R group	Ri_S_Hap31	0	1	S group
Ri_R_Hap16	1	0	R group	Ri_RS_Hap_32	1	1	Common

**Table 2.** Major histocompatibility complex B haplotypes in Ri chicken population.

211 R group, resistant group to avian influenza; S group, susceptible group to avian influenza

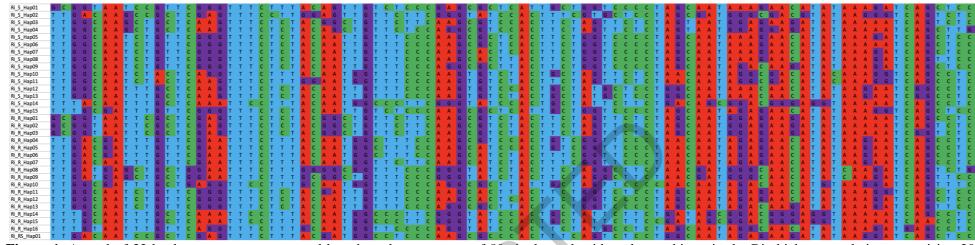
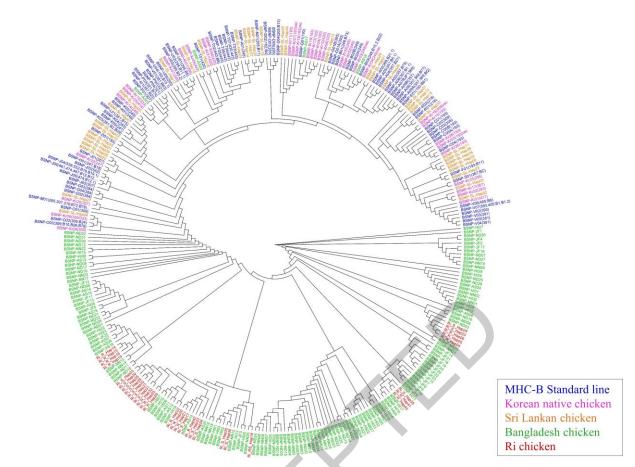


Figure 1. A total of 32 haplotypes were constructed based on the genotypes of 89 single nucleotide polymorphisms in the Ri chicken population comprising 20

 $\left( \right)$ 

С,

- 214 individuals.
- 214 215



- 217 218 219 Figure 2. Phylogenetic tree based on haplotypes in the major histocompatibility complex B standard lines, Korean native chicken, Sri Lankan chicken, Bangladesh chicken, and Ri chicken. Ri chicken
- 220 haplotypes grouped with Bangladesh chicken haplotypes while separated from other global chicken
- 221 haplotype groups.