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8 **Abstract**

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10 The size and location of the corpus luteum and the presence of coexistent follicles are crucial  
11 factors in synchronizing recipients and determining the suitability for embryo transfer. However,  
12 there has been a recent decline in conception rates after embryo transfer, which is attributed to  
13 environmental pollution, uterine inflammation, ovarian cysts, and other factors. Therefore, we  
14 conducted experiments to establish a novel criterion for successful embryo transfer assessment.  
15 To assess the suitability for embryo transfer one day before transfer, we conducted ultrasound  
16 examinations equipped with a vaginal probe to evaluate the corpus luteum and coexistent follicle.  
17 We found that instances with corpus luteum and coexistent follicles (diameter: >10 mm)  
18 constituted the majority (69.7%) of cases. When comparing the fertility rates of cases in which  
19 the corpus luteum and coexistent follicle (diameter: >10 mm) were located on the same ovary  
20 and cases in which they were not, higher fertility rates were observed when the corpus luteum  
21 and coexistent follicle (diameter: >10mm) were on different ovaries. Our study revealed a high  
22 incidence of corpus luteum and coexistent follicles with a diameter exceeding 10 mm. Therefore,  
23 our findings suggest that the co-occurrence of the corpus luteum and a large follicle can serve as  
24 a new standard for the evaluation of embryo transfer suitability.

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26 **Keywords:** corpus luteum, coexistent follicle, embryo transfer, conception rate, Hanwoo

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## Introduction

29 Embryo transfer has been a long-standing practice in both humans and livestock, including cows,  
30 pigs, and sheep. This technique is used in various fields, including biotechnology research,  
31 breeding improvement, preservation of genetic resources, and infertility resolution (1-3). In the  
32 case of cows, this technique is instrumental in producing offspring with exceptional genetic traits.  
33 Embryos are generated using superovulation methods, ovum pick-up (OPU), and ovaries  
34 obtained from slaughtered animals, which are then transplanted into recipient cows (3-5).  
35 Additionally, various synchronization methods, primarily centered around ovum synchronization,  
36 are employed to transfer embryos into multiple recipients simultaneously in cows (6-10).

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38 To ensure the success of embryo transfer, synchronizing the recipients and determining the  
39 presence or absence of the corpus luteum is crucial, which is typically accomplished through  
40 rectal palpation or ultrasound examination (6, 8, 11, 12). Furthermore, even in instances of  
41 synchronized estrus, various factors can impede recipient ovulation, including physiological  
42 irregularities, ovarian cysts, and endocrine inflammation (11, 13, 14). Additionally, research  
43 indicates that the highest conception rates are achieved when embryos are transferred into the  
44 uterine angle where the corpus luteum is present (15-17). Therefore, evaluating the presence,  
45 location, and size of the corpus luteum prior to embryo transfer is closely related to the  
46 pregnancy rate (11, 12, 18).

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48 Recent studies have demonstrated that the diameter of the corpus luteum and the coexistent  
49 follicle also play a significant role in affecting conception rates before embryo transfer (15, 18).  
50 Specifically, the size of the corpus luteum exhibits a positive correlation with fertility rates.  
51 Conversely, the size of coexistent follicles has a negative correlation with conception rates (12,  
52 15, 18, 19).

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54 Furthermore, research has indicated that conception rates are influenced by the content and ratio  
55 of the reproductive hormones progesterone and estrogen (12, 15, 18, 19). Progesterone is a  
56 hormone critical for maintaining pregnancy, whereas estrogen positively influences follicle  
57 development. Therefore, higher progesterone levels, lower estrogen levels, and a higher  
58 progesterone-to-estrogen ratio are associated with relatively higher fertility rates (12, 15, 18, 19).

59 Given that estrogen is derived from follicles, coexistent follicles must be absent or small in size  
60 to ensure successful embryo transfer (15, 18).

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62 Nevertheless, to the best of our knowledge, no previous studies have comprehensively compared  
63 and analyzed conception rates, progesterone, and estrogen concentrations when the corpus  
64 luteum and coexistent follicles are present on the same ovary compared to when they are located  
65 differently. Therefore, our study sought to compare the presence, size, and location of the corpus  
66 luteum and coexistent follicles in the context of the embryo transfer synchronization method and  
67 their impact on pregnancy rates. Additionally, we analyzed conception rates when the corpus  
68 luteum and coexistent follicle are on the same ovary versus when they are not, and compared the  
69 influence of progesterone and estrogen levels on fertility rates.

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## Materials and Methods

### Animals and Management

A total of 145 cows were employed in this experiment. The cows were reared at the Gyeongsangbuk-do Livestock Research Institute in accordance with the Hanwoo Korean Feeding Standard, and they were housed in a well-equipped space that provided ample room (300 m<sup>2</sup> for 15 cows) and stanchions. All experimental procedures involved in this study were approved by the Institutional Animal Care and Use Committee (IACUC) of the Gyeongsangbuk-do Livestock Research Institute.

### Experimental Design

Cows were synchronized using the E2/P4 (7), 2FTET (7), and J-synch (9) methods for embryo transfer. Detailed methods can be found in the related literature, as well as in Figure 1. The experiment included 40 cows subjected to the E2/P4 method, 73 cows in the 2FTET method, and 32 cows in the J-synch method (Fig. 1).

#### 1. E2/P4 method

For the E2/P4 synchronization method, a 2 mg intramuscular (i.m.) injection of estradiol benzoate (EB) (Samyang Anipharm Co., South Korea) was administered on day 0, along with the insertion of a 1.56 g progesterone-releasing device (Cue-Mate, Bioniche Animal Health, Australia) into the vagina at a random stage. On day 7, a 25 mg intramuscular injection of prostaglandin F2 $\alpha$  (PGF2 $\alpha$ ) (Lutalyse, Zoetis, USA) was administered, and the progesterone-releasing device was removed. On day 8, a 2 mg i.m. injection of EB was administered. Estrus was confirmed on day 9, and on day 15, the corpus luteum was assessed through rectal palpation via ultrasound examination. On day 16, one embryo was transferred (Fig. 1).

#### 2. 2FTET method

For the 2FTET synchronization method, a 2 mg i.m. injection of EB was administered on day 0, along with the insertion of a 1.56 g progesterone-releasing device into the vagina at a random stage. On day 6, a 25 mg intramuscular injection of prostaglandin F2 $\alpha$  (PGF2 $\alpha$ ) is given, and the progesterone-releasing device is removed. Estrus is confirmed on day 8. On day 9, 250  $\mu$ g of gonadotropin-releasing hormone (GnRH) (Gonadon, gonadorelin acetate,

103 Dong Bang Co., South Korea) was administered via i.m. injection. On day 15, the corpus  
104 luteum was assessed through rectal palpation via ultrasound examination. On day 16, one  
105 embryo was transferred (Fig. 1). Prior to conducting the pregnancy test, a 2 mg i.m. injection  
106 of EB was administered, and a 1.56 g progesterone-releasing device was inserted into the  
107 vagina on day 33. The progesterone-releasing device was then removed on day 39, and  
108 pregnancy was confirmed through rectal palpation via ultrasound examination. If the cow  
109 was found to be pregnant, the pregnancy was recorded without any further treatment. In the  
110 case of a non-pregnant cow, a 25 mg i.m. injection of PGF<sub>2</sub> $\alpha$  was administered on day 33,  
111 and estrus was confirmed on day 41. On day 42, 250  $\mu$ g of GnRH was i.m. injected. On day  
112 48, the corpus luteum was examined via rectal palpation using ultrasound examination, and  
113 on day 49, a second embryo was transferred (Fig. 1).

### 114 115 **3. J-synch method**

116 For the J-synch synchronization method, a 2 mg i.m. injection of EB was administered on  
117 day 0, along with the insertion of a 1.56 g progesterone-releasing device into the vagina at a  
118 random stage. On day 6, a 25 mg i.m. injection of prostaglandin F<sub>2</sub> $\alpha$  (PGF<sub>2</sub> $\alpha$ ) was  
119 administered, and the progesterone-releasing device was removed. Estrus was then  
120 confirmed, and 250  $\mu$ g of GnRH was administered via i.m. injection on day 9. On day 15,  
121 the corpus luteum was assessed through rectal palpation using ultrasound examination, after  
122 which one embryo was transferred on day 16 (Fig. 1).

### 123 124 **Distinguish experimental groups by measurement of corpus luteum and coexistent follicle**

125 The presence and diameter of the corpus luteum and coexistent follicle were measured using  
126 ultrasonic equipment equipped with vaginal probe ultrasonography (4Vet Slim, DRAMINKI,  
127 Poland). As illustrated in Supplementary Figure 1, the subjects were divided into four  
128 experimental groups. The “Only CL” group comprises cases where only the corpus luteum is  
129 present in the left or right ovary. The “CL+MF” group represents cases in which both the  
130 corpus luteum and a middle-sized (5–10 mm) coexistent follicle are observed. The “CL+LF”  
131 group consists of cases with the corpus luteum and a large-sized (>10 mm) coexistent follicle.  
132 The “LF” group includes cases where there is no corpus luteum, only a large-sized coexistent  
133 follicle. The “X” group pertains to cases in which neither a corpus luteum nor a follicle is

134 detected. For further analysis within the “CL+LF” groups, the combination of corpus luteum  
135 and the coexisting large follicle is designated as “Same side\_CL/LF” when they are within the  
136 same ovary and “Other side\_CL/LF” when they are found in different ovaries in  
137 Supplementary Figure 2.

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### 139 **Embryo production, embryo transfer, and pregnancy test**

140 The embryos utilized in this experiment were previously described in detail in a paper  
141 published by our research team (5). Cumulus-oocyte complexes were collected and cultured  
142 through the OPU method, and fresh embryos were subsequently transferred to the recipient. To  
143 enhance the accuracy of the experiment and eliminate potential confounding factors that could  
144 impact conception rates, a single expert conducted both the measurement of the corpus luteum  
145 and coexistent follicle and embryo transfer. Pregnancy testing was carried out via rectal  
146 palpation and ultrasound equipment (HS-101V; Honda, Japan) at least 23 days after embryo  
147 transfer.

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### 149 **Plasma collection and concentration of progesterone and estrogen ELISA kit**

150 Blood was drawn from the cow’s jugular vein one day prior to the embryo transfer, followed  
151 by centrifugation to separate the plasma. Using the isolated plasma, the levels of progesterone  
152 and estrogen in the blood were analyzed. The Bovine Progesterone ELISA kit (CSB-E08172b,  
153 CUSABIO Co., USA) and the Bovine Estradiol ELISA kit (CSB-E08173b, CUSABIO Co.,  
154 USA) were employed for this analysis.

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### 156 **Statistical Analysis**

157 The chi-square test was used to analyze the conception rate according to the size and  
158 location of the corpus luteum and coexistent follicle. Additionally, the correlation between  
159 conception rates and the levels of progesterone and estrogen was statistically examined  
160 through a 2-way ANOVA, followed by Tukey’s multiple comparisons test for *post hoc*  
161 analysis (GraphPad Prism, version 8.0.1, GraphPad Software Inc., USA).

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## Results

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Table 1 summarizes the results related to the distribution of corpus luteum and coexistent follicle one day before embryo transfer, categorized by the synchronization method. The “Only CL” group accounted for 7.6%, the “CL+MF” group represented 8.3%, the “CL+LF” group comprised 69.7%, the “LF” group was at 11.7%, and the “X” group constituted 2.8% (Table 1). Notably, the “CL+LF” group exhibited a significantly higher percentage compared to the other groups, with a significant difference observed among the experimental groups ( $p < 0.001$ ). No significant differences were observed between the presence of corpus luteum and coexistent follicle based on the synchronization methods E2/P4, 2FTET, and J-synch. In instances where the “LF” group ( $n=17$ ) and the “X” group ( $n=4$ ) were urgently vaccinated against FMD to prevent disease transmission ( $n=28$ ), the vaccinated cows were subsequently excluded from the embryo transfer procedure (Table 2).

A total of 96 cows out of the 145 synchronized cows underwent fresh embryo transfer using the OPU method (Table 2). The conception rates for embryo transfer according to the synchronization methods were determined to be 57.1% for the E2/P4 method, 37.1% for the 2FTET method, and 48.1% for the J-synch method. Importantly, no significant differences in conception rates were observed among the synchronization methods examined herein (Table 2). Among the experimental groups categorized based on the presence of corpus luteum and coexistent follicle, the “Only CL” group had a conception rate of 28.6%, the “CL+MF” group achieved 33.3% conception rate, and the “CL+LF” group yielded 43.0% conception rate. Notably, there were no significant differences in conception rates between these experimental groups.

Our study confirmed that a larger corpus luteum size is associated with a higher conception rate. As illustrated in Fig. 2, there was a significant difference in corpus luteum size according to pregnancy status ( $p < 0.001$ ). However, no significant difference was observed when analyzing the relationship between the size of the coexistent follicle and pregnancy (Fig. 2).

Figure 3 illustrates the analysis of progesterone and estrogen levels in the blood for the experimental groups, categorized based on the presence of corpus luteum and coexistent follicle. In terms of progesterone content, the “Only CL” group exhibited the highest levels compared to



197 the other groups. Furthermore, there was a tendency for progesterone levels to decrease as the  
198 size of the follicle increased, with a significant difference observed between the groups (Fig. 3).  
199 Regarding estrogen content, the “Only CL” group had the lowest levels compared to the other  
200 groups. Notably, significant differences were detected only between the “Only CL” and  
201 “CL+MF” groups and between the “CL+MF” and “LF” groups ( $p < 0.05$ ).

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203 Furthermore, as indicated in Table 3, the conception rates were compared by distinguishing  
204 between cases in which the corpus luteum and coexistent follicle ( $>10$  mm) were present in the  
205 same ovary (“Same side CL/LF” group) and cases where they were located in different ovaries  
206 (“Other side CL/LF” group). Upon comparing the fertility rates, we found that the “Other side  
207 CL/LF” group tended to have a higher fertility rate than the “Same side CL/LF” group, although  
208 this difference did not reach statistical significance (Table 3).

209

210 Although the results were not statistically significant, a comparison of the progesterone levels in  
211 the blood showed that the “Other side CL/LF” group had higher progesterone content than the  
212 “Same side CL/LF” group. Conversely, the blood estrogen levels were higher in the “Same side  
213 CL/LF” group compared to the “Other side CL/LF” group (Fig. 4).

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## Discussion

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Studies are actively underway to investigate synchronization methods aimed at enhancing the fertility rate of embryo transfer in cows (6, 7, 10, 11, 17, 20). Given that cows are domestic animals, the primary objective of embryo transfer tends to be profit-driven rather than genetic resource preservation. Furthermore, embryo transfer can result in the production of offspring with outstanding genetic traits, making it a potentially more lucrative option compared to artificial insemination (2, 3, 17). However, it is important to note that embryo transfer complex preparation procedures, advanced technology, and additional expenses related to purchasing embryos. Therefore, its utilization rate is lower when compared to artificial insemination (9, 15, 21).

Additional research efforts are thus needed to address issues such as reducing the acquisition costs and improving the low conception rates associated with embryo transfer. The outcomes of these efforts could be highly promising, as they would establish a basis for the generation of substantial profits through transplantation, in addition to significantly expediting the genetic improvement process. Traditionally, rather than conducting a detailed confirmation of the size of the corpus luteum and coexistent follicle prior to embryo transfer through ovarian ultrasound, a rectal test relying on palpation is commonly performed (11, 19). Technology based on ultrasonic equipment with rectal or vaginal probes has recently gained widespread popularity, albeit with the drawback of requiring specialized expertise. Our research team focuses on oocyte collection using OPU methods, embryo production, and embryo transfer, and therefore our team members are highly skilled in handling ultrasound equipment equipped with a vaginal probe (5). By leveraging this expertise, our study confirmed that the ratio of both corpus luteum and coexistent follicle (>10 mm) was notably high, reaching approximately 69.7% (101 out of 145 cows), thus exceeding previous findings. For instance, Msahiko et al. (15) reported that the ratio of both the corpus luteum and coexistent follicle (>10mm) was 32.8% (24/73 cows).

244 Numerous studies have demonstrated that the corpus luteum secretes progesterone, a hormone  
245 crucial for maintaining pregnancy, whereas the follicle secretes estrogen, a hormone necessary  
246 for follicle development (1, 3, 16, 19, 22). Therefore, we inferred that the presence of only the  
247 corpus luteum during embryo transfer positively impacts conception rates. However, the  
248 presence of both the corpus luteum and coexistent follicle has an adverse effect on pregnancy  
249 maintenance, thereby negatively affecting conception rates. Although it is physiologically ideal  
250 for only the corpus luteum to be present during embryo transfer, the exact mechanism underlying  
251 the simultaneous presence of the coexistent follicle remains to be fully understood. Hypotheses  
252 have been proposed, suggesting that cow-related diseases and environmental factors, such as  
253 environmental pollution, uterine inflammation, and ovarian cysts, may be primary contributing  
254 factors (1, 3, 13, 22).

255  
256 Previous literature has discouraged embryo transfer when both the corpus luteum and coexistent  
257 follicle are simultaneously present (12, 15, 23). Excluding the “CL+LF” (69.7%), “LF” (11.7%),  
258 and “X” (2.8%) groups, our findings confirmed that the aforementioned strategy is rather  
259 inefficient, as only 15.9% of cases allowed for embryo transfer (7.6% for “Only CL” and 8.3%  
260 for “CL+MF”). To address these limitations, we divided the CL+LF group into the “Same side  
261 CL/LF” and “Other side CL/LF” groups and compared the levels of progesterone and estrogen in  
262 the blood. Existing literature has already reported that successful embryo transfer is associated  
263 with high progesterone concentration and low estrogen concentration (12, 15, 19). In this study,  
264 we confirmed that the “Other side CL/LF” group exhibited higher progesterone levels and lower  
265 estrogen levels compared to the “Same side CL/LF” group, although this difference was not  
266 statistically significant. Therefore, our findings suggest that embryo transfer can be considered  
267 when the corpus luteum and coexistent follicle are present in different ovaries.

268  
269 Our findings highlighted the importance of meticulously assessing the presence and size of both  
270 the corpus luteum and coexistent follicle through ultrasound equipment to ensure the successful  
271 embryo transfer. Moreover, our findings provide foundational insights to study the mechanisms  
272 underlying the simultaneous presence of the corpus luteum and coexistent follicle. Therefore, the  
273 results of this study offer a valuable theoretical basis to guide the decision-making process  
274 regarding embryo transfer in cows, thus contributing to the improvement of farmers’ income, as  
275 well as conception rates.

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283 “Development of techniques to improve the reproductive performance in Korean native cows for  
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**References (Vancouver or NLM style)**

- 288 1. Ferre LB, Kjelland ME, Strobecch LB, Hyttel P, Mermillod P, Ross PJ. Review: Recent  
289 advances in bovine in vitro embryo production: reproductive biotechnology history and  
290 methods. *Animal*. 2020;14(5):991-1004. Epub 2019/11/26. doi:  
291 10.1017/S1751731119002775. PubMed PMID: 31760966.
- 292 2. Smith MF, Geisert RD, Parrish JJ. Reproduction in domestic ruminants during the past 50  
293 yr: discovery to application. *J Anim Sci*. 2018;96(7):2952-70. Epub 2018/04/24. doi:  
294 10.1093/jas/sky139. PubMed PMID: 29684167; PubMed Central PMCID:  
295 PMCPMC6095338.
- 296 3. Lovarelli D, Bacenetti J, Guarino M. A review on dairy cattle farming: Is precision livestock  
297 farming the compromise for an environmental, economic and social sustainable production?  
298 *Journal of Cleaner Production*. 2020;262. doi: 10.1016/j.jclepro.2020.121409.
- 299 4. Pontes JH, Melo Sterza FA, Basso AC, Ferreira CR, Sanches BV, Rubin KC, et al. Ovum  
300 pick up, in vitro embryo production, and pregnancy rates from a large-scale commercial  
301 program using Nelore cattle (*Bos indicus*) donors. *Theriogenology*. 2011;75(9):1640-6.  
302 Epub 2011/02/22. doi: 10.1016/j.theriogenology.2010.12.026. PubMed PMID: 21334055.
- 303 5. Kim D, Yi J. Improving Cryopreservation Efficiency and Pregnancy Rate through  
304 Superovulation with Follicle-Stimulating Hormone in Korean Hanwoo Cows via Ovum Pick  
305 Up. *Vet Sci*. 2023;10(9). Epub 2023/09/27. doi: 10.3390/vetsci10090578. PubMed PMID:  
306 37756101; PubMed Central PMCID: PMCPMC10534669.
- 307 6. Baruselli PS, Ferreira RM, Sales JN, Gimenes LU, Sa Filho MF, Martins CM, et al. Timed  
308 embryo transfer programs for management of donor and recipient cattle. *Theriogenology*.  
309 2011;76(9):1583-93. Epub 2011/07/30. doi: 10.1016/j.theriogenology.2011.06.006. PubMed  
310 PMID: 21798580.
- 311 7. Bo GA, Cedeno A, Mapletoft RJ. Strategies to increment in vivo and in vitro embryo  
312 production and transfer in cattle. *Anim Reprod*. 2019;16(3):411-22. Epub 2020/05/22. doi:  
313 10.21451/1984-3143-AR2019-0042. PubMed PMID: 32435285; PubMed Central PMCID:  
314 PMCPMC7234104.
- 315 8. Bo GA, Peres LC, Cutaia LE, Pincinato D, Baruselli PS, Mapletoft RJ. Treatments for the  
316 synchronisation of bovine recipients for fixed-time embryo transfer and improvement of  
317 pregnancy rates. *Reprod Fertil Dev*. 2011;24(1):272-7. Epub 2012/03/08. doi:  
318 10.1071/RD11918. PubMed PMID: 22394969.
- 319 9. Perez-Mora A, Segura-Correa JC, Peralta-Torres JA. Factors associated with pregnancy rate

- 320 in fixed-time embryo transfer in cattle under humid-tropical conditions of Mexico. *Anim*  
321 *Reprod.* 2020;17(2):e20200007. Epub 2020/07/28. doi: 10.1590/1984-3143-AR2020-0007.  
322 PubMed PMID: 32714459; PubMed Central PMCID: PMC7375863.
- 323 10. Bo GA, de la Mata JJ, Baruselli PS, Menchaca A. Alternative programs for synchronizing  
324 and resynchronizing ovulation in beef cattle. *Theriogenology.* 2016;86(1):388-96. Epub  
325 2016/05/18. doi: 10.1016/j.theriogenology.2016.04.053. PubMed PMID: 27180326.
- 326 11. Benyei B, Komlosi I, Pecsí A, Pollott G, Marcos CH, de Oliveira Campos A, et al. The  
327 effect of internal and external factors on bovine embryo transfer results in a tropical  
328 environment. *Anim Reprod Sci.* 2006;93(3-4):268-79. Epub 2005/09/20. doi:  
329 10.1016/j.anireprosci.2005.07.012. PubMed PMID: 16169166.
- 330 12. Thomson SP, Holmes RJ, Landes PT, Allworth MB. Assessment and selection of the  
331 recipient cows' corpus luteum at the time of embryo transfer, and its influence on conception  
332 rate. *Aust Vet J.* 2021;99(7):288-92. Epub 2021/04/30. doi: 10.1111/avj.13068. PubMed  
333 PMID: 33913151.
- 334 13. Wang J, Li J, Wang F, Xiao J, Wang Y, Yang H, et al. Heat stress on calves and heifers: a  
335 review. *J Anim Sci Biotechnol.* 2020;11:79. Epub 2020/08/14. doi: 10.1186/s40104-020-  
336 00485-8. PubMed PMID: 32789013; PubMed Central PMCID: PMC7416401.
- 337 14. Chebel RC, Demetrio DG, Metzger J. Factors affecting success of embryo collection and  
338 transfer in large dairy herds. *Theriogenology.* 2008;69(1):98-106. Epub 2007/11/21. doi:  
339 10.1016/j.theriogenology.2007.09.008. PubMed PMID: 18023856.
- 340 15. Msahiko NISHIGAI, Hideo KAMOMAE, Tomomi TANAKA, KANEDA Y. The  
341 Relationship of Blood Progesterone and Estrogen Concentrations on the Day Before and the  
342 Day of Frozen-Thawed Embryo Transfer to Pregnancy Rate in Japanese Black Beef Cattle.  
343 *Journal of Reproduction and Development.* 2000;46(4):235-43. doi:  
344 doi.org/10.1262/jrd.46.235.
- 345 16. Cerri RL, Chebel RC, Rivera F, Narciso CD, Oliveira RA, Amstalden M, et al.  
346 Concentration of progesterone during the development of the ovulatory follicle: II. Ovarian  
347 and uterine responses. *J Dairy Sci.* 2011;94(7):3352-65. Epub 2011/06/28. doi:  
348 10.3168/jds.2010-3735. PubMed PMID: 21700021.
- 349 17. Looney CR, Nelson JS, Schneider HJ, Forrest DW. Improving fertility in beef cow recipients.  
350 *Theriogenology.* 2006;65(1):201-9. Epub 2005/11/18. doi:  
351 10.1016/j.theriogenology.2005.09.023. PubMed PMID: 16289261.
- 352 18. A.M. Gonella-Diazal GH, D. Montaña, D. Valbu. Corpus luteum diameter and embryo

353 developmental stage are associated with pregnancy rate : data analysis from 17,521 embryo  
354 transfers from a commercial in vitro bovine embryo production program. *Animal*  
355 *Reproduction*. 2013;10(2):106-11.

356 19. Henricks DM, Dickey JF, Hill JR, Johnston WE. Plasma estrogen and progesterone levels  
357 after mating, and during late pregnancy and postpartum in cows. *Endocrinology*.  
358 1972;90(5):1336-42. Epub 1972/05/01. doi: 10.1210/endo-90-5-1336. PubMed PMID:  
359 5012745.

360 20. de la Mata JJ, Nunez-Olivera R, Cuadro F, Bosolasco D, de Brun V, Meikle A, et al. Effects  
361 of extending the length of pro-oestrus in an oestradiol- and progesterone-based oestrus  
362 synchronisation program on ovarian function, uterine environment and pregnancy  
363 establishment in beef heifers. *Reprod Fertil Dev*. 2018;30(11):1541-52. Epub 2018/05/21.  
364 doi: 10.1071/RD17473. PubMed PMID: 29778102.

365 21. Lopreiato V, Mezzetti M, Cattaneo L, Ferronato G, Minuti A, Trevisi E. Role of  
366 nutraceuticals during the transition period of dairy cows: a review. *J Anim Sci Biotechnol*.  
367 2020;11:96. Epub 2020/08/31. doi: 10.1186/s40104-020-00501-x. PubMed PMID:  
368 32864127; PubMed Central PMCID: PMC7450574.

369 22. Diskin MG, Waters SM, Parr MH, Kenny DA. Pregnancy losses in cattle: potential for  
370 improvement. *Reprod Fertil Dev*. 2016;28(1-2):83-93. Epub 2015/02/01. doi:  
371 10.1071/RD15366. PubMed PMID: 27062877.

372 23. Roper DA, Schrick FN, Edwards JL, Hopkins FM, Prado TM, Wilkerson JB, et al. Factors in  
373 cattle affecting embryo transfer pregnancies in recipient animals. *Anim Reprod Sci*.  
374 2018;199:79-83. Epub 2018/11/18. doi: 10.1016/j.anireprosci.2018.11.001. PubMed PMID:  
375 30442469.

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## Tables and Figures

378 Table 1. Changes in corpus luteum and coexistent follicle size before embryo transfer (n=145).

379 (a-b: Values with different letters, a and b, are significantly different at  $P < 0.001$ )

Group	E2/P4		2FTET		J-synch		Total	
	No. of Cow	%	No. of Cow	%	No. of Cow	%	No. of Cow	%
Only CL	4	10.0%	5	6.8%	2	6.3%	11	7.6% <sup>a</sup>
CL+MF <sup>§</sup>	9	22.5%	1	1.4%	2	6.3%	12	8.3% <sup>a</sup>
CL+LF <sup>§</sup>	22	55.0%	56	76.7%	23	71.9%	101	69.7% <sup>b</sup>
LF <sup>§</sup>	5	12.5%	9	12.3%	3	9.4%	17	11.7% <sup>a</sup>
X	0	0.0%	2	2.7%	2	6.3%	4	2.8% <sup>a</sup>
Total	40	100.0%	73	100.0%	32	100.0%	145	100.0%

380 <sup>§</sup> Medium (5–10 mm) and large (>10 mm) coexistent follicle with corpus luteum, <sup>a,b</sup> Corpus luteum and follicle size  
 381 was analyzed using the chi-square test. Statistical significance was set at  $p < 0.001$ .

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384 Table 2. Conception rate according to corpus luteum size, coexistent follicle size, and embryo transfer method  
 385 (n=96).

Group	E2/P4		2FTET		J-synch		No. of pregnant cow/Total	Pregnancy rates (%)
	No. of Cow	%	No. of Cow	%	No. of Cow	%		
Only CL	-*	-	1/5	20.0%	1/2	50.0%	2/7	28.6%
CL+MF <sup>§</sup>	-*	-	1/1	100.0%	0/2	0.0%	1/3	33.3%
CL+LF <sup>§</sup>	4/7*	57.1%	21/56	37.5%	12/23	52.2%	37/86	43.0%
Total	4/7*	57.1%	23/62	37.1%	13/27	48.1%	40/96	41.7%

386 <sup>§</sup> Medium (5–10 mm) and large (>10 mm) coexistent follicle with corpus luteum. A total of 21 cows belonging to  
 387 the large follicle and X groups did not undergo embryo transfer. \* A total of 28 cows were vaccinated against foot-  
 388 and-mouth disease three days after the embryo transfer and were therefore excluded from the experiment.

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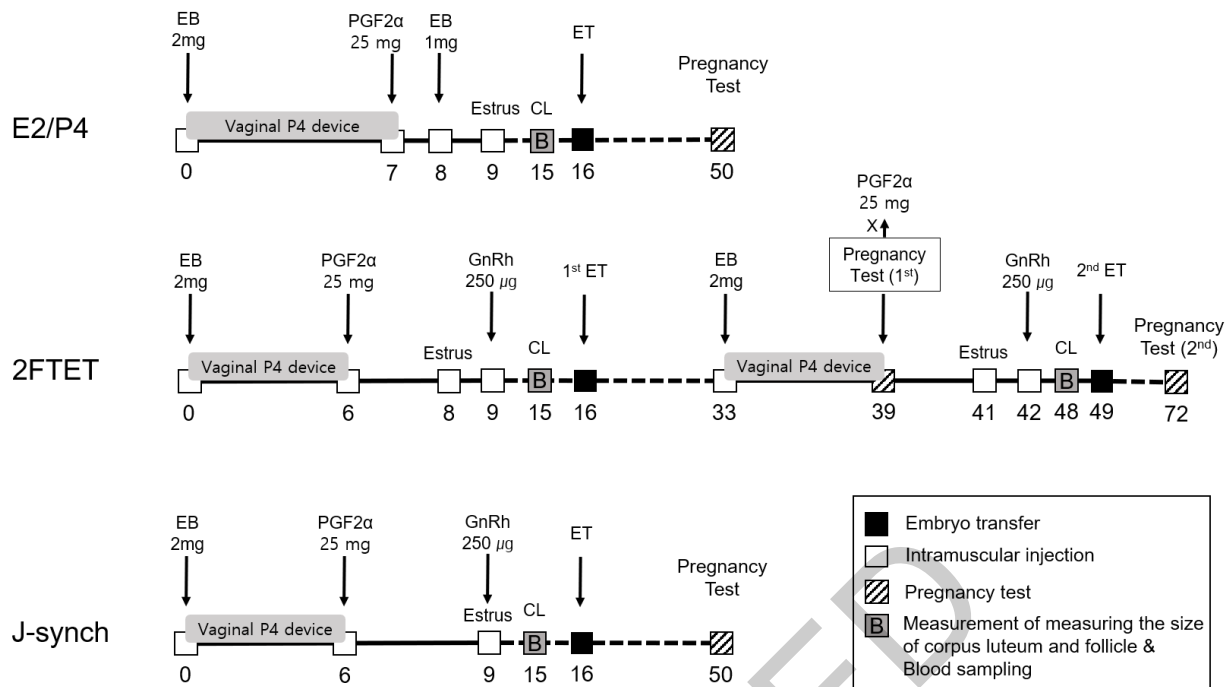
392 Table 3. Conception rate according to the position of the coexistent follicle (>10mm) prior to embryo transfer  
393 (n=86).

Group	No. of pregnant cow	Total	Pregnancy rates (%)
Same side CL/LF	13	42	31.0%
Other side CL/LF	24	44	54.5%
Total	37	86	43.0%

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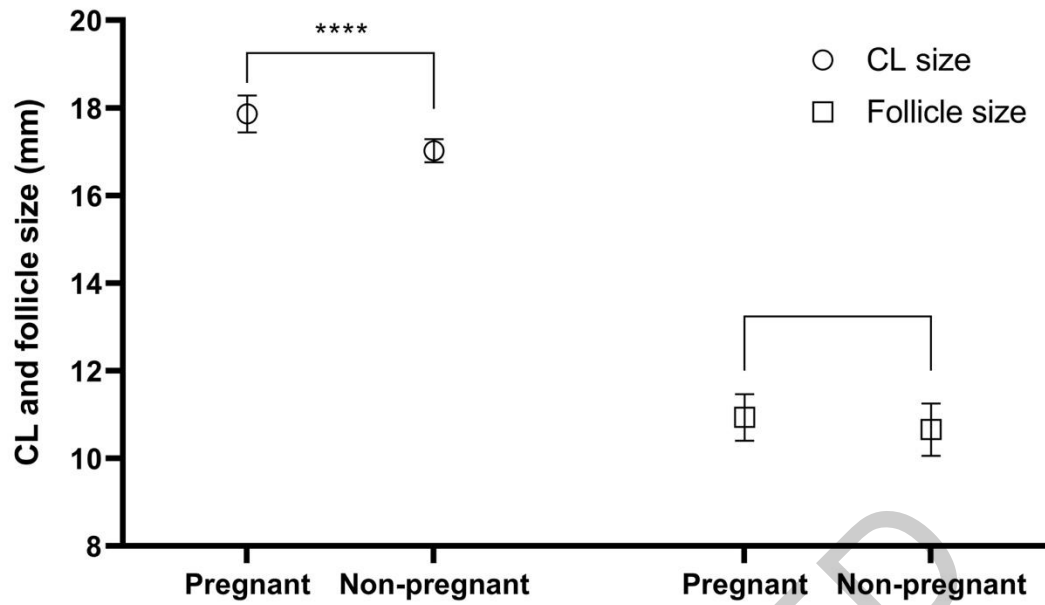
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**Figure 1. Synchronization method utilized in the experiment.** Black square boxes represent embryo transfers, white square boxes denote intramuscular injections, shaded square boxes indicate pregnancy tests, and "B" within a square box indicates measurement of corpus luteum and coexistent follicle to determine embryo transfer, with blood collection for further analysis.



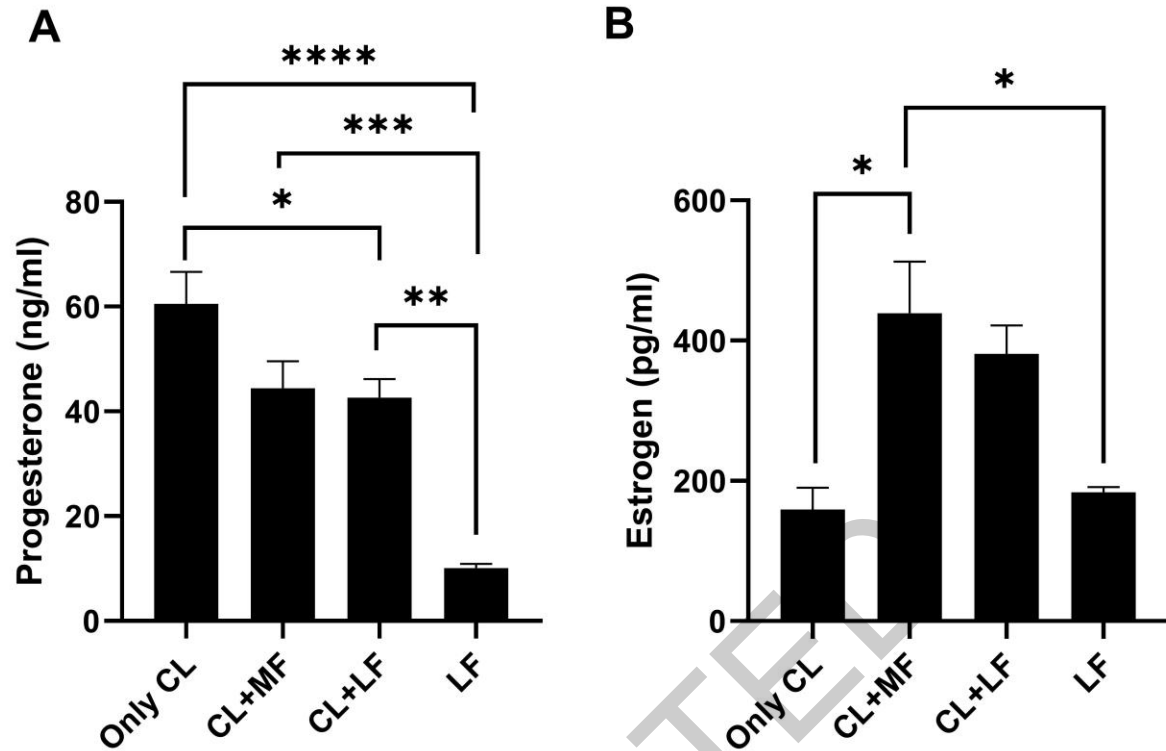
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404 **Figure 2. Size of corpus luteum and coexistent follicle according to pregnancy (n=96).** Differences in the size

405 (mean  $\pm$  SEM) of the corpus luteum and follicle were analyzed via two-way analysis of variance (ANOVA)

406 (Tukey's multiple comparisons test). \*\*\*\* Significance level  $p < 0.001$ .

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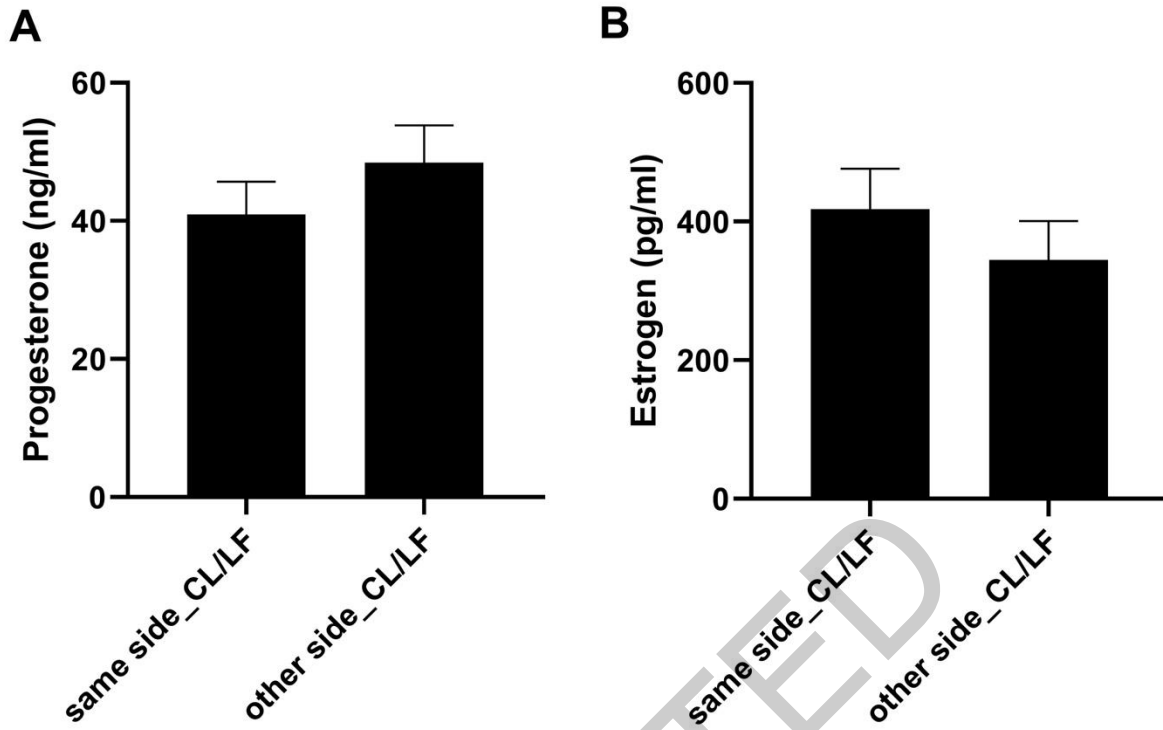
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**Figure 3. Plasma concentration of progesterone and estrogen 1 day before embryo transfer (n=96).** The gray bar represents the plasma concentration of progesterone and estrogen categorized into four groups based on corpus luteum and follicle size. Differences in plasma concentration (mean  $\pm$  SEM) of progesterone and estrogen were analyzed using two-way analysis of variance (ANOVA) (Tukey's multiple comparisons test). \*\*\*\* Significance level  $p < 0.001$ . \*\*\* Significance level  $p < 0.005$ . \*\* Significance level  $p < 0.01$ . \* Significance level  $p < 0.05$ .



417  
 418 **Figure 4. Plasma concentration of progesterone and estrogen a day before embryo transfer according to the**  
 419 **location of corpus luteum and coexistent follicle (n=86).** The gray bars represent plasma progesterone and  
 420 estrogen concentrations categorized into two groups based on the location of the corpus luteum and coexistent large  
 421 follicle.

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