JAST (Journal of Animal Science and Technology) TITLE PAGE

Upload this completed form to website with submission

ARTICLE INFORMATION	Fill in information in each box below
Article Type	Short Communication
Article Title (within 20 words without abbreviations)	Effect of *Sasa quelpaertensis* Nakai extract on gut microbiota and production performance in pigs
Running Title (within 10 words)	Effect of SQE on gut microbiota and productivity in pigs
Author	Jongan Lee[1], Hyeon-Ah Kim[1], Yong-Jun Kang[1], Yoo-Kyung Kim[1], and Moon-Cheol Shin[2,*]
Affiliation	[1]Subtropical Livestock Research Institute, National Institute of Animal Science, RDA, Jeju 63242, Republic of Korea [2]National Institute of Animal Science, RDA, Wanju 55365, Republic of Korea
ORCID (for more information, please visit https://orcid.org)	Jongan Lee (https://orcid.org/0000-0002-4761-1808) Hyeon-Ah Kim (https://orcid.org/0000-0002-4203-9857) Yong-Jun Kang (http://orcid.org//0000-0001-8949-7831) Yoo-Kyung Kim (https://orcid.org/0000-0002-6685-7467) Moon-Cheol Shin (https://orcid.org/0000-0001-6744-7155)
Competing interests	No potential conflict of interest relevant to this article was reported.
Funding sources State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available.	This work was supported by the "Cooperative Research Program for Agriculture Science & Technology Development (Project No. PJ01573901)" Rural Development Administration, Republic of Korea.
Acknowledgements	This study was supported by 2023 the RDA Fellowship Program of the National Institute of Animal Science, Rural Development Administration, Republic of Korea.
Availability of data and material	The 16S rRNA amplicon sequencing data deposited in the NCBI Sequence Read Archive (SRA) database with accession number PRJNA998697.
Authors' contributions Please specify the authors' role using this form.	Conceptualization: Lee J, Shin MC. Data curation: Kim HA, Kang YJ, Kim YJ. Formal analysis: Lee J, Kim HA. Methodology: Kim HA, Kang YJ, Kim YJ. Software: Lee J, Shin MC. Validation: Lee J, Shin MC. Investigation: Lee J, Shin MC. Writing - original draft: Lee J. Writing - review & editing: Lee J, Kim HA, Kang YJ, Kim YJ, Shin MC.
Ethics approval and consent to participate	All experimental procedures involving animals were approved by the Institutional Animal Care and Use Committee (IACUC) of the National Institute of Animal Science (NIAS) (number: NIAS20212189).

CORRESPONDING AUTHOR CONTACT INFORMATION

For the corresponding author (responsible for correspondence, proofreading, and reprints)	Fill in information in each box below
First name, middle initial, last name	Moon-Cheol Shin
Email address – this is where your proofs will be sent	shinemoon@korea.kr
Secondary Email address	herops2453@gmail.com

Address	National Institute of Animal Science, RDA, 500, Kongjwipatjwiro, iseomyeon, Wanju, Jeollabukdo, Republic of Korea
Cell phone number	+82-10-2727-8932
Office phone number	+82-63-238-7127
Fax number	+82-63-238-8148

6

Effect of *Sasa quelpaertensis* Nakai extract on gut microbiota and production performance in pigs

Jongan Lee[1], Hyeon-Ah Kim[1], Yong-Jun Kang[1], Yoo-Kyung Kim[1], and Moon-Cheol Shin[1,*]

[1]Subtropical Livestock Research Institute, National Institute of Animal Science, RDA, Jeju 63242, Republic of Korea.

[2] National Institute of Animal Science, RDA, Wanju 55365, Republic of Korea.

*Corresponding author:

Moon-Cheol Shin. Tel: +82-63-238-7127, Fax: +82-63-238-8148, E-mail: shinemoon@korea.kr

Abstract

Different dietary patterns affect the gut microbial compositions and diversity. Consistently, microbiome alterations are linked to digestion, immunity, and productivity. *Sasa quelpaertensis* Nakai (SQ) is a perennial bamboo species rich in proteins and fiber. Previous studies have confirmed the health benefits of SQ; however, the effects of SQ supplementation on gut microbiome and production performance are unclear. Herein, Landrace pigs were supplemented with SQ extract (SQE) and gut microbial compositions as opposed to the control group were assessed using 16S rRNA sequencing. Additionally, the influences of SQE supplementation on average daily gain (ADG) and backfat thickness (BF) were assessed after slaughter. In the SQE group, Firmicutes and Actinobacteria phyla increased significantly, whereas Bacteroidetes and Spirochaetes phyla markedly decreased ($p < 0.05$). The expression level of *Bifidobacterium* and *Lactobacillus* genera increased, whereas that of *Treponema*, *Prevotella*, and *Turicibacter* decreased ($p < 0.05$). The microbial richness was similar between groups; however, microbial diversity decreased in the SQE supplementation group. Additionally, the SQE supplementation in pigs resulted in a slight increase in ADG. In contrast, BF in the SQE group decreased notably ($p < 0.05$). These results underscore the significant influence of SQE supplementation on the gut microbiota and demonstrate the potential of SQ as a valuable feed resource for enhancing animal productivity.

Keywords (3 to 6): Gut microbiota, diversity, *Sasa quelpaertensis* Nakai, production performance, pig

1. Introduction

Interactions between the microbiome and diet affect microbial colonization of the gastrointestinal tract in pigs. Specifically, dietary fiber supplementation in pigs has been associated with both negative and positive effects. A high fiber diet increases the expression level of *Lactobacillus* spp. in the digestive tract and enhances the volatile fatty acids (VFAs) production in the hindgut of weaned piglets [1]. VFA production is essential for regulating metabolism and contributes to animal health [2]. Insoluble fiber content reportedly induces changes in the gut microbiota, specifically by lessening the Firmicutes:Bacteroidetes (F/B) ratio and the expression level of *Lactobacillus* spp., which leads to increased susceptibility of pigs to colitis [3]. Additionally, different dietary protein sources affect the microbial composition of pigs. For example, cottonseed meal-based diets have been shown to increase the expression of *Lactobacillus* spp., which may improve gut health. In contrast, a fishmeal-based diet increases the abundance of *Escherichia* spp. and *Shigella* spp., which in turn increases the likelihood of diarrhea [4].

Sasa quelpaertensis Nakai (SQ) is a perennial bamboo species belonging to the grass family *Poaceae*, and is known for its high protein and fiber content. SQ is widespread, particularly on Mount Halla, Jeju Island, South Korea. A few researchers have assessed the influences of SQ as a potential ingredient in functional materials. SQ extract (SQE) has been shown to exert various health benefits, exhibiting antidiabetic, antioxidative, anti-inflammatory, and antiviral effects [5-7]. More recently, Lee et al. reported that, compared with rice straw, SQ has superior feed value as a roughage source for Hanwoo cattle when fed as part of a total mixed ration [8].

Despite the positive effects of SQ highlighted in previous research, the influences of SQ supplementation on the gut microbial communities of livestock remain unknown. Furthermore, the effects of SQ supplementation on important phenotypes related to livestock productivity, such as growth performance and carcass traits, have not been investigated. Therefore, in the present study, using SQE supplementation and control groups, and changes in the microbial composition and diversity in pigs were assessed using 16S rRNA amplicon sequencing. The influences of SQE supplementation on average daily gain (ADG) and backfat thickness (BF) were also assessed after slaughter. This study is, to our knowledge, the first to assess the influences of SQ supplementation on intestinal bacteria in pigs. We believe that these research findings will be helpful in demonstrating the potential of SQ as a feed resource and its ability to enhance animal productivity.

2. Materials and Methods

2.1. Animals and diet

All experiments involving animals presented in this research were approved by the Institutional Animal Care and Use Committee (IACUC) of the National Institute of Animal Science (NIAS) (approval number: NIAS20212189). A total of 14 Landrace pigs were used in the study, with seven pigs (five males and two females) in the control group and seven pigs (two males and five females) in the SQE supplementation group. The 14 Landrace pigs were raised in a standardized environment at the Subtropical Livestock Research Institute (Jeju, South Korea). The age of the pigs at the start of the experiment ranged from 151 to 160 d, with an average age of 154.8 d. The average ± standard deviation of body weight of the control group and SQE supplementation group were 80.57 ± 10.79 kg and 80.00 ± 11.28 kg, respectively. The experiment was conducted in the same rearing place for 41 d, wherein the control group was fed a commercial formula at a rate of 2.25 kg per pig twice a day, for a total of 4.5 kg per day. The SQE supplementation group received at same frequency and amount as the control group, with an additional 450 ml of SQE added to the commercial formula per feeding by top dressing. The chemical composition of the commercial formula and SQE are listed in Table 1.

2.2. SQE and production performance

The Jeju Plant Resources Institute (Jeju, South Korea) obtained permission from Jeju Island and collected SQ on Mount Halla. The collected SQ was washed and dried at 60 °C for 8-14 h. SQE was prepared as a mixture of dried SQ and water in a 1:20 ratio, which was then subjected to high-pressure extraction at 100 °C for 3 h. To calculate ADG, the initial body weight and the final body weight after 41 d of the experiment were measured in 14 Landrace pigs, and the weight differences were divided by the experimental period of 41 d. The 14 Landrace pigs were slaughtered within eight days after the end of the experiment to measure the BF in Jeju Livestock Cooperative (Jeju, South Korea). The BF information of 14 Landrace pigs was acquired from the Korea Institute for Animal Products Quality Evaluation (https://www.ekape.or.kr).

2.3. DNA extraction and 16S rRNA amplicon sequencing

After the experiment was completed, fecal samples were acquired from the 14 pigs and stored at −70 °C. Genomic DNA was extracted from a fecal sample (250 mg) using the QIAamp PowerFecal Pro DNA kit (Qiagen, Hilden, Germany). The quantity and quality of the extracted genomic DNA were measured using a NanoDrop ND-2000 spectrophotometer (Thermo Fisher Scientific, Waltham, MA, USA). The V3-V4 hypervariable segments of the 16S ribosomal RNA gene were amplified from the fecal genomic DNA using the universal primer sets Bakt_341F (CCTACGGGNGGCWGCAG) and Bakt_805R (GACTACHVGGGTATCTAATCC). The input DNA 2 ng was PCR amplified with 5x reaction buffer, 1 mM of dNTP mix, 500 nM each of the universal F/R PCR primer, and Herculase II fusion DNA polymerase (Agilent Technologies, Santa Clara, CA). Following purification, the 2 ul of 1st PCR product was PCR amplified for final library construction containing the index using Nextera XT Indexed Primer. The cycle condition for 2nd PCR was same as the 1st PCR condition except for 10 cycles. The PCR product was purified with AMPure beads. The final purified product is then quantified using qPCR according to the qPCR Quantification Protocol Guide (KAPA Library Quantification kits for Illumina Sequencing platforms) and qualified using the

TapeStation D1000 ScreenTape (Agilent Technologies, Waldbronn, Germany). The sequencing of amplified products was conducted using an Illumina MiSeq platform (Illumina, San Diego, CA, USA).

2.4. Taxonomic assignment and diversity analysis

The microbiome taxonomic profiling (MTP) of EzBioCloud (ChunLab Inc., South Korea) was utilized to perform microbial classification. Briefly, the sequencing reads were processed as follows: 1) paired reads were merged into a single read; 2) forward and reverse primer sequences were trimmed; 3) low-quality (< Q25) reads were filtered; 4) non-redundant reads were denoised and extracted; 5) chimeric reads were detected and removed; and 6) operational taxonomic units (OTUs) with similarity greater than 97% were selected. Microbial taxa were classified using the EzBioCloud 16S rRNA database (version PKSSU4.0) [9]. The taxonomic composition was normalized using the copy number of the 16S rRNA genes. Good's coverage for sequencing depth and diversity indices (Chao1, Shannon, and Simpson) of the microbial communities were estimated to identify species richness and diversity. Principal coordinate analysis (PCoA) method was employed to measure beta diversity using UniFrac distance matrices, including unclassified OTUs at the species level.

2.5. Statistical analysis

The Wilcoxon rank-sum test was employed to assess the difference in ADG, BF, and microbial diversity between groups. The results were considered statistically significant at $p < 0.05$. We used linear discriminant analysis effect size (LEfSe) method to measure the effect size of taxa, and defined taxa with a linear discriminant analysis (LDA) score > 4 and $p < 0.05$ as microbiota with differential expression between groups [10].

3. Results & Discussion

3.1 Alterations of the gut microbial composition and production performance in pigs

The total number of bases (bp), reads, GC (%), Q20 (%), and Q30 (%) were calculated to estimate the quality of the 16S rRNA amplicon sequencing (Table 2). To investigate how SQE supplementation affects the gut microbial environment, the relative proportion of the taxa in both groups was compared at the different taxonomic levels (Fig. 1A–1D). Firmicutes, Bacteroidetes, Spirochaetes, and Actinobacteria were identified as the most common phyla in both the control and SQE groups (Fig. 1A). LEfSe results indicated that the expression level of Firmicutes (LDA score = 4.69, $p = 0.008$) and Actinobacteria (LDA score = 4.44, $p = 0.018$) increased substantially, whereas Bacteroidetes (LDA score = 4.49, $p = 0.006$) and Spirochaetes (LDA score = 4.61, $p = 0.025$) decreased in the SQE group (Fig. 1E). The proportion of Firmicutes was 76.48% in the SQE supplementation group and 67.28% in the control group, while that of Bacteroidetes was 8.32% and 14.26%, respectively. The F/B ratio was increased in the SQE group (9.19) relative to the control group (4.71). Higher F/B ratios have been linked to increased energy absorption and accumulation in humans [11]. Additionally, obese pigs reportedly show elevated F/B ratios in their gut microbiota [12]. There were no notable differences observed in ADG between groups ($p = 0.52$), the ADG showed a tendency to increase in the SQE group (Fig. 2A). These results suggest that the increase in F/B ratios due to SQE supplementation may affect host energy metabolism and body weight gain. The SQE group (3.79%) exhibited a lower expression level of Spirochaetes in comparison to the control group (10.62%). Spirochaetes are intestinal pathogens that cause various diseases, including mucohemorrhagic colitis, typhlitis, and cholera [13]. SQE has been shown to be effective in reducing inflammation in intestinal epithelial cells [14], and recently, a correlation has been reported between an elevated expression of Spirochaetes and the occurrence of African Swine Fever [15]. These results suggest that SQE supplementation can potentially suppress inflammation and prevent disease by reducing the expression level of Spirochaetes. The abundance of Actinobacteria was elevated in the SQE group (11.28%) versus the control group (5.22%). Actinobacteria are actively involved in preserving gut homeostasis and development of the immune system [16]. Based on these results, maintaining the balance and stability of the microbial environment within the immune system can protect pigs from disease.

The proportion of *Bifidobacterium* (LDA score = 4.44, $p = 0.018$) and *Lactobacillus* (LDA score = 5.00, $p = 0.004$) genera was higher in the SQE group, whereas that of *Treponema* (LDA score = 4.58, $p = 0.006$), *Prevotella* (LDA score = 4.21, $p = 0.035$), and *Turicibacter* (LDA score = 4.15, $p = 0.025$) decreased in the SQE group (Fig. 1E). Lactic acid bacteria (LAB), including *Lactobacillus* and *Bifidobacterium* regulate the intestinal environment in pigs. LAB can inhibit or eliminate pathogenic agents in the digestive tract, improving the microbiome balance and preserving the intestinal barrier [17]. Interestingly, pigs with low BF have a higher abundance of *Lactobacillus reuteri* in the gut [18]. Furthermore, *Lactobacillus* spp. were associated with a decrease in fat mass in mice provided with a high-fat diet [19]. We observed a significant decrease in BF in the SQE group as opposed to the control group ($p < 0.05$) (Fig. 2B). A study has reported a positive connection between a high abundance of *Prevotella copri* and elevated levels of serum metabolites associated with obesity [20]. SQE supplementation can reduce the weight of fat tissue in the obesity mouse model and regulate abundances of key proteins participating in fat metabolism in rats provided with a high-fructose diet [21, 22]. Our results suggest that SQE supplementation in pigs can increase the expression level of *Lactobacillus*

and decrease the expression level of *Prevotella*, resulting in a reduction in BF.

Diet is a major factor that affects the gut microbial composition, and environmental conditions and age also affect the composition of the microbiome. In this study, we were limited to raising 14 pigs in the same rearing space during the experimental period in order to minimize the alterations of microbiome caused by environmental conditions. To minimize the changes in the microbiome caused by age differences, we excluded pigs that were more than two weeks apart in age from the experimental group and recruited pigs that were born around the same time. In this study, we performed a comparative analysis of the gut microbiome compositions in a small number of pigs. However, to increase the statistical power of the LEfSe method, we increased the LDA score threshold from the typical value of 2 to 4. A higher LDA score means that the gut microbiome is more likely to be present in one group at a higher relative abundance than in the other group. Although the sample size is small, we believe that results of microbial changes due to the additional feeding of SQE are acceptable because the LDA score threshold was increased. In addition, further studies on a larger number of pigs are needed to investigate the effects of SQE feeding on various growth performances such as feed intake and feed efficiency.

3.2 Comparison of microbial richness and diversity

To assess the distribution of taxa between groups, we conducted alpha diversity analysis, which estimated the microbial richness and diversity. Good's coverage index exceeded 99.4%, suggesting that the depth of 16S rRNA sequencing was sufficient to capture the fecal microbiota (Fig. 3A). The Chao1 index, a measure of species richness, showed similar observation values between groups ($p > 0.05$) (Fig. 3B). These findings indicate that SQE supplementation had a minimal impact on the count of different species present in the microbial communities. The diversity of species was evaluated using the Simpson and Shannon indices. The Shannon index decreased in the SQE group; however, the difference between observed values was marginal ($p = 0.053$) (Fig. 3C). In contrast, the Simpson index increased notably in the SQE group ($p < 0.05$) (Fig. 3D). Considering the diversity results, SQE supplementation affects the evenness of different species, leading to a less balanced composition of the microbiota. The PCoA plot of beta diversity evaluated by UniFrac distances showed dissimilarity in the microbial communities between the control and SQE groups (Fig. 3E).

Funding sources

This work was supported by the “Cooperative Research Program for Agriculture Science & Technology Development (Project No. PJ01573901)” Rural Development Administration, Republic of Korea.

Acknowledgments

This study was supported by 2023 the RDA Fellowship Program of the National Institute of Animal Science, Rural Development Administration, Republic of Korea.

References

1. Bikker P, Dirkzwager A, Fledderus J, Trevisi P, Le Huërou-Luron I, Lallès JP, et al. The effect of dietary protein and fermentable carbohydrates levels on growth performance and intestinal characteristics in newly weaned piglets. Journal of animal science. 2006;84(12):3337-45.https://doi.org/10.2527/jas.2006-076

2. Koh A, De Vadder F, Kovatcheva-Datchary P, Bäckhed F. From dietary fiber to host physiology: short-chain fatty acids as key bacterial metabolites. Cell. 2016;165(6):1332-45.https://doi.org/10.1016/j.cell.2016.05.041

3. Burrough ER, Arruda BL, Patience JF, Plummer PJ. Alterations in the colonic microbiota of pigs associated with feeding distillers dried grains with solubles. PLoS One. 2015;10(11):e0141337.https://doi.org/10.1371/journal.pone.0141337

4. Cao K, Zhang H, Han H, Song Y, Bai X, Sun H. Effect of dietary protein sources on the small intestine microbiome of weaned piglets based on high-throughput sequencing. Letters in Applied Microbiology. 2016;62(5):392-8.https://doi.org/10.1111/lam.12559

5. Yoon S-A, Kang S-I, Shin H-S, Ko H-C, Kim S-J. Anti-diabetic potential of a Sasa quelpaertensis Nakai extract in L6 skeletal muscle cells. Food Science and Biotechnology. 2014;23:1335-9.https://doi.org/10.1007/s10068-014-0183-4

6. Kim S-J, Hwang J-H, Shin H-S, Jang M-G, Ko H-C, Kang S-I. Antioxidant and anti-inflammatory activities of sasa quelpaertensis leaf extracts. Phytochemicals as Nutraceuticals-Global Approaches to Their Role in Nutrition and Health: IntechOpen; 2012.https://doi.org/10.5772/26874

7. Kang H, Lee C. Sasa quelpaertensis Nakai extract suppresses porcine reproductive and respiratory syndrome virus replication and modulates virus-induced cytokine production. Archives of virology. 2015;160:1977-88.https://doi.org/10.1007/s00705-015-2469-0

8. Lee S, Baek YC, Lee M, Jeon S, Bang HT, Seo S. Evaluating feed value of native Jeju bamboo (Sasa quelpaertensis Nakai) for beef cattle. Animal Bioscience. 2023;36(2):238.https://doi.org/10.5713/ab.22.0160

9. Yoon S-H, Ha S-M, Kwon S, Lim J, Kim Y, Seo H, et al. Introducing EzBioCloud: a taxonomically united database of 16S rRNA gene sequences and whole-genome assemblies. International journal of systematic and evolutionary microbiology. 2017;67(5):1613.https://doi.org/10.1099/ijsem.0.001755

10. Segata N, Izard J, Waldron L, Gevers D, Miropolsky L, Garrett WS, et al. Metagenomic biomarker discovery and explanation. Genome biology. 2011;12:1-18.https://doi.org/10.1186/gb-2011-12-6-r60

11. Turnbaugh PJ, Hamady M, Yatsunenko T, Cantarel BL, Duncan A, Ley RE, et al. A core gut microbiome in obese and lean twins. nature. 2009;457(7228):480-4.https://doi.org/10.1038/nature07540

12. Guo X, Xia X, Tang R, Wang K. Real-time PCR quantification of the predominant bacterial divisions in the distal gut of Meishan and Landrace pigs. Anaerobe. 2008;14(4):224-8.https://doi.org/10.1016/j.anaerobe.2008.04.001

13. Hampson DJ, Ahmed N. Spirochaetes as intestinal pathogens: lessons from a Brachyspira genome. Gut pathogens. 2009;1(1):1-3.https://doi.org/10.1186/1757-4749-1-10

14. Kim K-M, Kim Y-S, Lim JY, Min SJ, Ko H-C, Kim S-J, et al. Intestinal anti-inflammatory activity of Sasa quelpaertensis leaf extract by suppressing lipopolysaccharide-stimulated inflammatory mediators in intestinal epithelial Caco-2 cells co-cultured with RAW 264.7 macrophage cells. Nutrition Research and Practice. 2015;9(1):3-10.https://doi.org/10.4162/nrp.2015.9.1.3

15. Ko Y-S, Tark D, Moon S-H, Kim D-M, Lee TG, Bae D-Y, et al. Alteration of the Gut Microbiota in Pigs Infected with African Swine Fever Virus. Veterinary Sciences. 2023;10(5):360.https://doi.org/10.3390/vetsci10050360

16. Binda C, Lopetuso LR, Rizzatti G, Gibiino G, Cennamo V, Gasbarrini A. Actinobacteria: a relevant minority for the maintenance of gut homeostasis. Digestive and Liver Disease. 2018;50(5):421-8.https://doi.org/10.1016/j.dld.2018.02.012

17. Yang F, Hou C, Zeng X, Qiao S. The use of lactic acid bacteria as a probiotic in swine diets. Pathogens. 2015;4(1):34-45.https://doi.org/10.3390/pathogens4010034

18. Cao Y, Wang F, Wang H, Wu S, Bao W. Exploring a Possible Link between the Fecal Microbiota and the Production Performance of Pigs. Veterinary Sciences. 2022;9(10):527.https://doi.org/10.3390/vetsci9100527

19. Song W, Song C, Li L, Wang T, Hu J, Zhu L, et al. Lactobacillus alleviated obesity induced by high-fat diet in mice. Journal of Food Science. 2021;86(12):5439-51. https://doi.org/10.1111/1750-3841.15971

20. Chen C, Fang S, Wei H, He M, Fu H, Xiong X, et al. Prevotella copri increases fat accumulation in pigs fed with formula diets. Microbiome. 2021;9(1):1-21.https://doi.org/10.1186/s40168-021-01110-0

21. Kang S-I, Shin H-S, Kim H-M, Hong Y-S, Yoon S-A, Kang S-W, et al. Anti-obesity properties of a Sasa quelpaertensis extract in high-fat diet-induced obese mice. Bioscience, biotechnology, and biochemistry. 2012;76(4):755-61.https://doi.org/10.1271/bbb.110868

22. Park JY, Jang MG, Oh JM, Ko HC, Hur S-P, Kim J-W, et al. Sasa quelpaertensis leaf extract ameliorates dyslipidemia, insulin resistance, and hepatic lipid accumulation in high-fructose-diet-fed rats. Nutrients. 2020;12(12):3762.https://doi.org/10.3390/nu12123762

Tables and Figures

Table 1. Chemical composition of the commercial formula and SQE.

Commercial formula	Quantity
Crude Protein (%)	13.11
Crude Fiber (%)	2.74
Moisture (%)	12.16
NDF (%)	9.75
ADF (%)	3.14
Crude Ash (%)	4.33
Crude Fat (%)	5.53
GE (Kcal/g)	4.05
DE (Kcal/g)	3.30
SQE	**Quantity**
Moisture (%)	99.7
Carbohydrate (%)	0.1
Protein (%)	0.1
Dietary Fiber (%)	0.1
Na (%)	0.0115
Ca (%)	0.0019
Fe (%)	0.0001
K (%)	0.0159
GE (Kcal/g)	0.01

NDF, neutral detergent fiber; ADF, acid detergent fiber; GE, gross energy; DE, digestible energy.

Table 2. Assembly statistics of 16S rRNA amplicon sequencing, Q20 (%): The ratio of bases with Phred quality score of 20 or higher; Q30 (%): The ratio of bases with Phred quality score of 30 or higher.

Group	Sample ID	Total bases (bp)	The number of total reads	GC (%)	Q20 (%)	Q30 (%)
Control	L22-188	44,776,760	148,760	53.9	91.2	82.3
	L22-193	44,127,804	146,604	53.4	91.8	83.0
	L22-199	44,124,794	146,594	54.0	91.2	82.2
	L22-200	41,687,898	138,498	53.8	89.1	79.3
	L22-201	48,285,216	160,416	54.0	91.0	81.9
	L22-207	46,370,856	154,056	53.5	91.6	82.7
	L22-208	43,538,446	144,646	54.0	91.7	82.9
SQE	L22-186	42,644,476	141,676	53.9	91.5	82.6
	L22-190	43,236,242	143,642	53.8	91.5	82.8
	L22-192	47,592,314	158,114	53.8	91.5	82.7
	L22-198	35,927,962	119,362	54.2	90.0	80.2
	L22-202	41,680,674	138,474	53.6	89.7	80.2
	L22-209	40,436,340	134,340	53.5	92.3	83.9
	L22-226	40,438,748	134,348	54.0	91.9	83.2

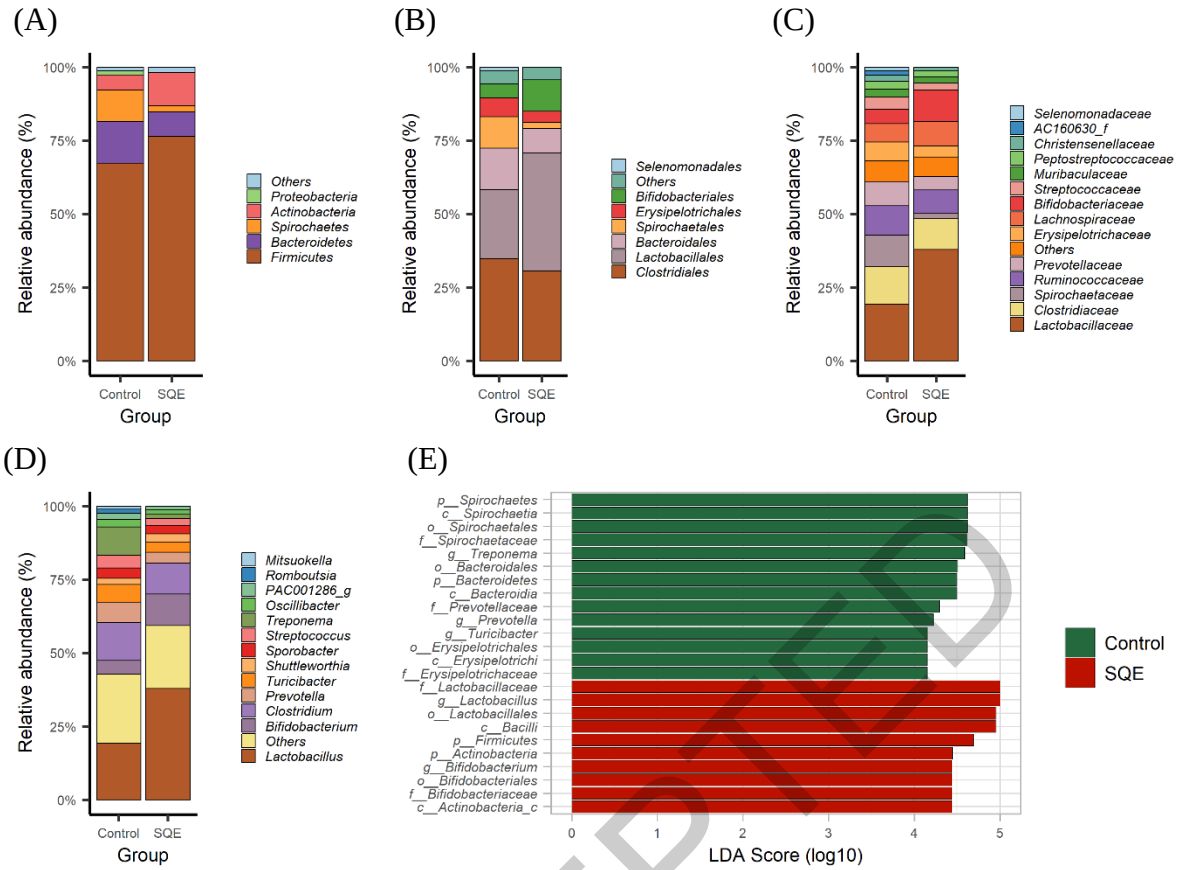

Figure 1. Relative abundances and linear discriminant analysis effect size (LEfSe) of gut microbiota between the control and SQE groups. Distribution of gut microbiota at the (A) phylum, (B) order, (C) family, and (D) genus levels. (E) Taxonomic levels from the phylum to the genus (LDA score > 4, $p < 0.05$). Horizontal bars represent the effect size for each taxon. LDA, linear discriminant analysis; SQE, *Sasa quelpaertensis* Nakai extract.

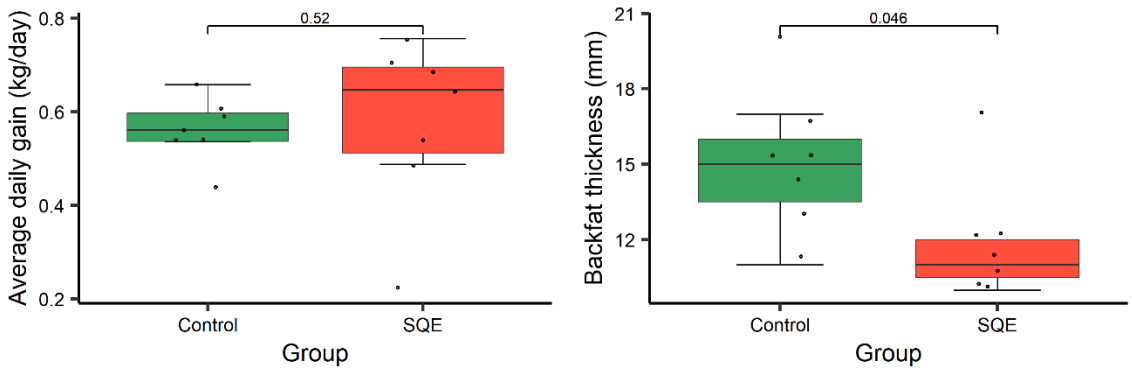

Figure 2. Average daily gain (ADG) and backfat thickness (BF) between the control and SQE groups. Boxplot of (A) ADG and (B) BF. Boxes represent the interquartile range (IQR) between the 25th and 75th percentiles, whereas the horizontal line within the box indicates the median value. The whiskers refer to the two lines extending from the box, spanning from the minimum value to the lower quartile. The *p*-value as assessed using the Wilcoxon rank-sum test is indicated above the boxplot. SQE, *Sasa quelpaertensis* Nakai extract.

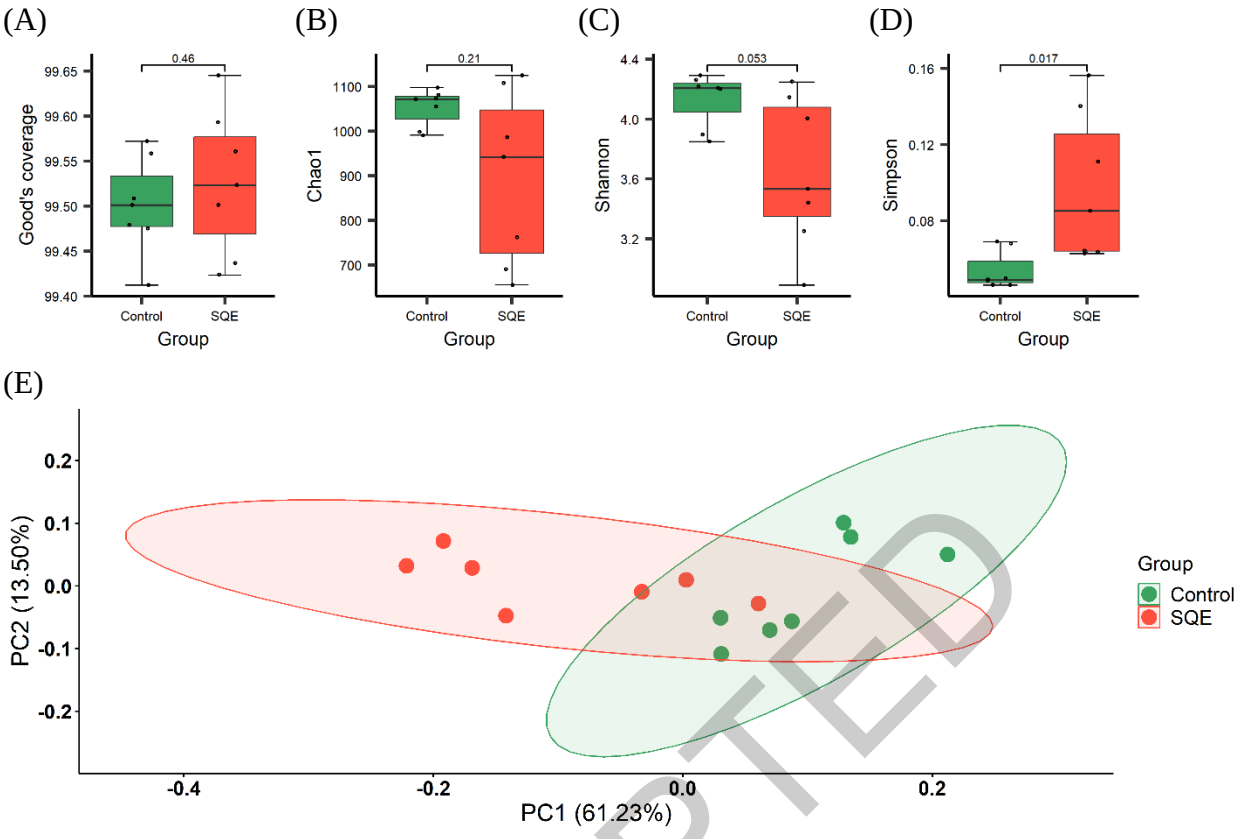

Figure 3. Alpha and beta diversity of gut microbiota between the control and SQE groups. Boxplot of (A) Good's coverage, (B) Chao1 index, (C) Shannon index, and (D) Simpson index. Boxes represent the IQR between the 25th and 75th percentiles, whereas the horizontal line within the box indicates the median value. The whiskers refer to the two lines extending from the box, spanning from the minimum value to the lower quartile. The *p*-value as assessed using the Wilcoxon rank-sum test is indicated above the boxplot. (E) Plot of principal coordinate analysis (PCoA). PCoA was performed at a species level with UniFrac distances, including unclassified operational taxonomic units. SQE, *Sasa quelpaertensis* Nakai extract.