

JAST (Journal of Animal Science and Technology) TITLE PAGE

Upload this completed form to website with submission

ARTICLE INFORMATION	Fill in information in each box below
Article Type	Genome Announcement
Article Title (within 20 words without abbreviations)	Complete genome sequence of bacteriocin-producing <i>Ligilactobacillus salivarius</i> B4311 isolated from fecal samples of broiler chicken with anti-listeria activity
Running Title (within 10 words)	Genome of <i>Ligilactobacillus salivarius</i> B4311, bacteriocin-producing strain
Author	Subin Han ^{1†} , Arxel G. Elnar ^{1†} , Chiwoong Lim ¹ , Geun-Bae Kim ¹
Affiliation	¹ Department of Animal Science and Technology, Chung-Ang University, Anseong 17546, Korea
ORCID (for more information, please visit https://orcid.org)	Subin Han (https://orcid.org/0000-0001-6658-2822) Arxel G. Elnar (https://orcid.org/0000-0002-2716-4924) Chiwoong Lim (https://orcid.org/0000-0002-6272-4464) Geun-Bae Kim (https://orcid.org/0000-0001-8531-1104)
Competing interests	No potential conflict of interest relevant to this article was reported.
Funding sources State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available.	This research was supported by the Chung-Ang University Graduate Research Scholarship (Academic Scholarship for College of Biotechnology and Natural Resources) in 2021.
Acknowledgements	Not applicable.
Availability of data and material	Upon reasonable request, the datasets of this study can be available from the corresponding author.
Authors' contributions Please specify the authors' role using this form.	Conceptualization: Kim GB Data curation: Han S, Elnar AG Formal analysis: Elnar AG, Kim GB Methodology: Elnar AG, Kim GB Software: Elnar AG, Chiwoong Lim, Kim GB Validation: Kim GB Investigation: Elnar AG Writing - original draft: Han S, Elnar AG Writing - review & editing: Han S, Elnar AG, Chiwoong Lim, Kim GB †Han S and Elnar AG contributed equally to this work.
Ethics approval and consent to participate	This article does not require IRB/IACUC approval because there are no human and animal participants.

2 CORRESPONDING AUTHOR CONTACT INFORMATION

For the corresponding author (responsible for correspondence, proofreading, and reprints)	Fill in information in each box below
First name, middle initial, last name	Geun-Bae Kim
Email address – this is where your proofs will be sent	kimgeun@cau.ac.kr
Secondary Email address	
Address	Department of Animal Science and Technology, Chung-Ang University, Anseong 17546, Korea
Cell phone number	
Office phone number	+82-31-670-3027
Fax number	+82-31-676-5986

3 **Abstract**

4 *Ligilactobacillus* is a genus of Gram-positive lactobacilli commonly found in the intestinal tracts of
5 vertebrates. It has been granted a Qualified Presumption of Safety (QPS) status from the European Food
6 Safety Authority (EFSA). One specific strain, *Ligilactobacillus salivarius* B4311, was isolated from
7 fecal samples of broiler chickens from a farm associated with Chung-Ang University (Anseong, Korea).
8 This strain was observed to have inhibitory effects against *Listeria monocytogenes*. In this paper, we
9 present the complete genome sequence of *Lig. salivarius* B4311. The whole genome of strain B4311
10 comprises 2,071,255 bp assembled into 3 contigs representing a chromosome, *repA*-type megaplasmid,
11 and small plasmid. The genome contains 1,963 protein-coding sequences, 22 rRNA genes, and 78 tRNA
12 genes, with a guanine + cytosine (GC) content of 33.1%. The megaplasmid of strain B4311 was found
13 to contain the bacteriocin gene cluster for salivaricin P, a two-peptide bacteriocin belonging to class IIb.

14
15 **Keywords:** *Ligilactobacillus salivarius*, probiotics, bacteriocin, *Listeria monocytogenes*

18 **Main Text**

19 Bacteriocin production in lactic acid bacteria (LAB) has been consistently gaining attention owing to
20 its potential as a viable alternative to antibiotics. Bacteriocins are ribosomally-synthesized peptides
21 secreted by the producing strain. These peptides can have either a narrow or broad spectrum of activity,
22 which indirectly determines the niche of the producing strain. The production of bacteriocins is
23 generally viewed as a positive trait as it enables the producing strain to hinder potential competitors in
24 the immediate environment as well as inhibit potentially harmful microorganisms (1). The
25 proteinaceous nature of bacteriocins renders them suitable for human use as they can be inactivated by
26 digestive proteases. With the rapid development of antimicrobial drug resistance in microorganisms (2),
27 research efforts focused on developing alternative solutions must be prioritized.

28
29 Commonly associated with vertebrate hosts, *Ligilactobacillus* is a genus of lactic acid bacteria
30 composed of members that are homofermentative, non-motile, and urease-positive. Their ability to

31 survive in gastric acid conditions and their Qualified Presumption of Safety (QPS) status from the
32 European Food Safety Authority (EFSA) (3) make them popular choices for probiotics. Furthermore,
33 the production of various antimicrobial salivaricins among strains of *Lig. salivarius* is well accounted
34 for the development of probiotic strains (4). In the present study, we report the genome analysis of a
35 bacteriocin-producing *Ligilactobacillus salivarius* (formerly *Lactobacillus salivarius*) strain B4311,
36 which was isolated from fecal samples collected from broiler chickens.

37

38 Strain B4311 was routinely cultured in de Mann, Rogosa, Sharpe (MRS, BD Difco, USA) broth
39 supplemented with 0.05% L-cysteine, and incubated aerobically at 37°C for 24 h. Genomic DNA was
40 extracted using the MagAttract HWM DNA Kit (Qiagen, Germany) and quantified using Qubit ds DNA
41 HS assay kit (Invitrogen, USA) with the Epoch™ Spectrometer (BioTek). The genome was sequenced
42 using the Pacific Biosciences (PacBio, USA) Sequel II platform. *De novo* assembly of the sequence
43 reads was performed using the PacBio SMAR Analysis program (ver. 2.3.0). Functional annotation of
44 the genome was performed using PRODIGAL ver. 2.6.2 (5) software and compared against protein
45 databases (SwissProt, KEGG, SEED, EggNOG). Rapid annotation was employed using Subsystem
46 Technology (RAST) with default parameters (<https://rast.nmpdr.org/>). Transfer RNAs (tRNA) and non-
47 coding ribosomal RNAs (rRNA) were identified using tRNAscan-SE ver. 1.3.1 (6) and INFERNAL
48 ver. 1.1.3 (7), respectively.

49

50 The complete genome of *Lig. salivarius* B4311 is 2,071,255 bp which is assembled into three contigs:
51 a single chromosome (1,801,655 bp), one megaplasmid (247,930 bp), and a small plasmid (21,670 bp)
52 with a guanine + cytosine (GC) content of 33.1%. In addition, the genome contains 1,963 protein-coding
53 sequences, 22 rRNA genes, and 78 tRNA genes. The genome features and circular maps of strain B4311
54 are presented in Table 1 and Fig. 1, respectively. Antimicrobial resistance genes, specifically for
55 tetracycline and glycopeptides, were also detected via Resistance Gene Identifier (RGI,
56 <https://card.mcmaster.ca/home>). Among the 1,963 CDS, 1,241 were predicted with biological functions
57 associated with cell cycle (n = 23), cell wall and motility (n = 116), cellular response (n = 69), DNA
58 processing (n = 154), RNA processing (n = 119), protein processing (n = 202), defense mechanism (n

59 = 31), energy production (n = 63), and transport and metabolism (n = 464). Additionally, 61 putative
60 genes were detected with putative functions including stress response, DNA and RNA processing,
61 antibiotic resistance, periplasm signaling, acetylation, amino acid transport, and production of enzymes
62 including various hydrolases, methyltransferases, and transport proteins.

63

64 *In silico* analysis of the B4311 genome using BAGEL4 online program (<http://bagel4.molgenrug.nl/>)
65 revealed the presence of a bacteriocin gene cluster for salivaricin P, a family of two-peptide bacteriocins
66 belonging to class IIb. This bacteriocin family was originally discovered from *Lig. salivarius* DPC6005
67 (6) and is commonly produced among strains of *Lig. salivarius* isolated from animals intestines (8). The
68 salivaricin P gene cluster is located in the *repA*-type megaplasmid. Although the presence of
69 megaplasms is considered a typical feature of *Lig. salivarius*, variations exist among megaplasmid-
70 encoded traits, including contingency metabolism genes (i.e., assimilation of sugars) and the presence
71 or absence of bacteriocin genes, which provides a competitive advantage.

72

73 The genetic architecture of the bacteriocin gene cluster (Fig. 2) revealed the presence of two open
74 reading frames (ORFs) encoding the salivaricin P chain A and chain B. The two peptide chains share a
75 homologous sequence. Located downstream of the genes for the bacteriocin peptides are two ORFs
76 encoding a histidine kinase and *AbpR*, which function as regulator proteins (9). These are followed by
77 *AbpIM* which encodes an immunity protein. These five ORFs are flanked by two *comC* genes, which
78 have been reported as competence-stimulating peptide precursors in streptococci (10). At the 3' end of
79 the gene cluster, two export proteins, *LanT* and *HlyD* were detected, encoding *AbpT* and *AbpD*
80 bacteriocin export accessory proteins, respectively. Several ORFs encoding bacteriocin core peptides
81 (i.e., lactacin F and plantaricins) were also detected. However, the similarity of these genes with the
82 reference was poor, suggesting that the translated peptides might be inactive.

83

84 Production of active bacteriocins was demonstrated by spot-on-lawn assay against *Listeria*
85 *monocytogenes* ATCC 19114 and ATCC 19115 strains. The cell-free supernatant of strain B4311
86 successfully inhibited the growth of *Lis. monocytogenes* (unpublished data), a common foodborne

87 pathogen associated with raw and unpasteurized milk and the causative agent of listeriosis. The genomic
88 information presented in this study confirms the ability of strain B4311 to elaborate bioactive peptides,
89 which can have valuable applications in the food and animal industries.

90

91

92 **Nucleotide sequence accession number**

93 The sequence obtained in this Whole Genome Shotgun project has been deposited in
94 DDBJ/ENA/GenBank under the accession number CP117983-CP117985. The BioProject accession
95 number PRJNA932943 is and the Biosample accession number is SAMN33215311.

96

97 **Acknowledgments**

98 This research was supported by the Chung-Ang University Graduate Research Scholarship (Academic
99 Scholarship for College of Biotechnology and Natural Resources) in 2021.

100

101 **Availability of data and material**

102 Upon a reasonable request, the datasets of this study can be requested from the corresponding author.

103

104

105 **References**

- 106 1. Dobson A, Cotter PD, Ross RP, Hill C. Bacteriocin production: a probiotic trait? Appl
107 Environ Microbiol. 2012;78(1):1-6.
- 108 2. Darbandi A, Asadi A, Mahdizade Ari M, Ohadi E, Talebi M, Halaj Zadeh M, et al.
109 Bacteriocins: Properties and potential use as antimicrobials. J Clin Lab Anal.
110 2022;36(1):e24093.
- 111 3. Hazards EPoB, Koutsoumanis K, Allende A, Alvarez-Ordenez A, Bolton D, Bover-Cid S,
112 et al. Update of the list of QPS-recommended biological agents intentionally added to food
113 or feed as notified to EFSA 12: suitability of taxonomic units notified to EFSA until March
114 2020. EFSA J. 2020;18(7):e06174.
- 115 4. Dec M, Stepien-Pysniak D, Puchalski A, Hauschild T, Pietras-Ozga D, Ignaciuk S, et al.
116 Biodiversity of *Ligilactobacillus salivarius* Strains from Poultry and Domestic Pigeons.
117 Animals (Basel). 2021;11(4).
- 118 5. Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. Prodigal: prokaryotic
119 gene recognition and translation initiation site identification. BMC Bioinformatics.
120 2010;11:119.
- 121 6. Barrett E, Hayes M, O'Connor P, Gardiner G, Fitzgerald GF, Stanton C, et al. Salivaricin P,
122 one of a family of two-component antilisterial bacteriocins produced by intestinal isolates of
123 *Lactobacillus salivarius*. Appl Environ Microbiol. 2007;73(11):3719-23.
- 124 7. Nawrocki EP, Eddy SR. Infernal 1.1: 100-fold faster RNA homology searches.
125 Bioinformatics. 2013;29(22):2933-5.
- 126 8. Messaoudi S, Manai M, Kergourlay G, Prevost H, Connil N, Chobert JM, et al. *Lactobacillus*
127 *salivarius*: bacteriocin and probiotic activity. Food Microbiol. 2013;36(2):296-304.
- 128 9. Barbour A, Wescombe P, Smith L. Evolution of Lantibiotic Salivaricins: New Weapons to
129 Fight Infectious Diseases. Trends Microbiol. 2020;28(7):578-93.
- 130 10. L.S. H, R. H, P. G. Natural Competence in the Genus *Streptococcus*: Evidence that
131 Streptococci Can Change Pherotype by Interspecies Recombinational Exchanges. Journal of
132 Bacteriology. 1997;179(21):6589-94.

133

Legends of Tables and Figures

Table 1. Genome features of *Ligilactobacillus salivarius* B4311.

Fig. 1. Circular map of *Ligilactobacillus salivarius* B4311 genome. G, guanine; C, cytosine; CDS, coding sequences. Circles represent the following characteristics from the outermost circle to the center: (1) contig information, (2) coding sequences on forward strand, (3) coding sequences on reverse strand, (4) transfer RNAs (tRNAs) and ribosomal RNAs (rRNAs), (5) GC skew, and (6) GC ratio. G, guanine; C, cytosine.

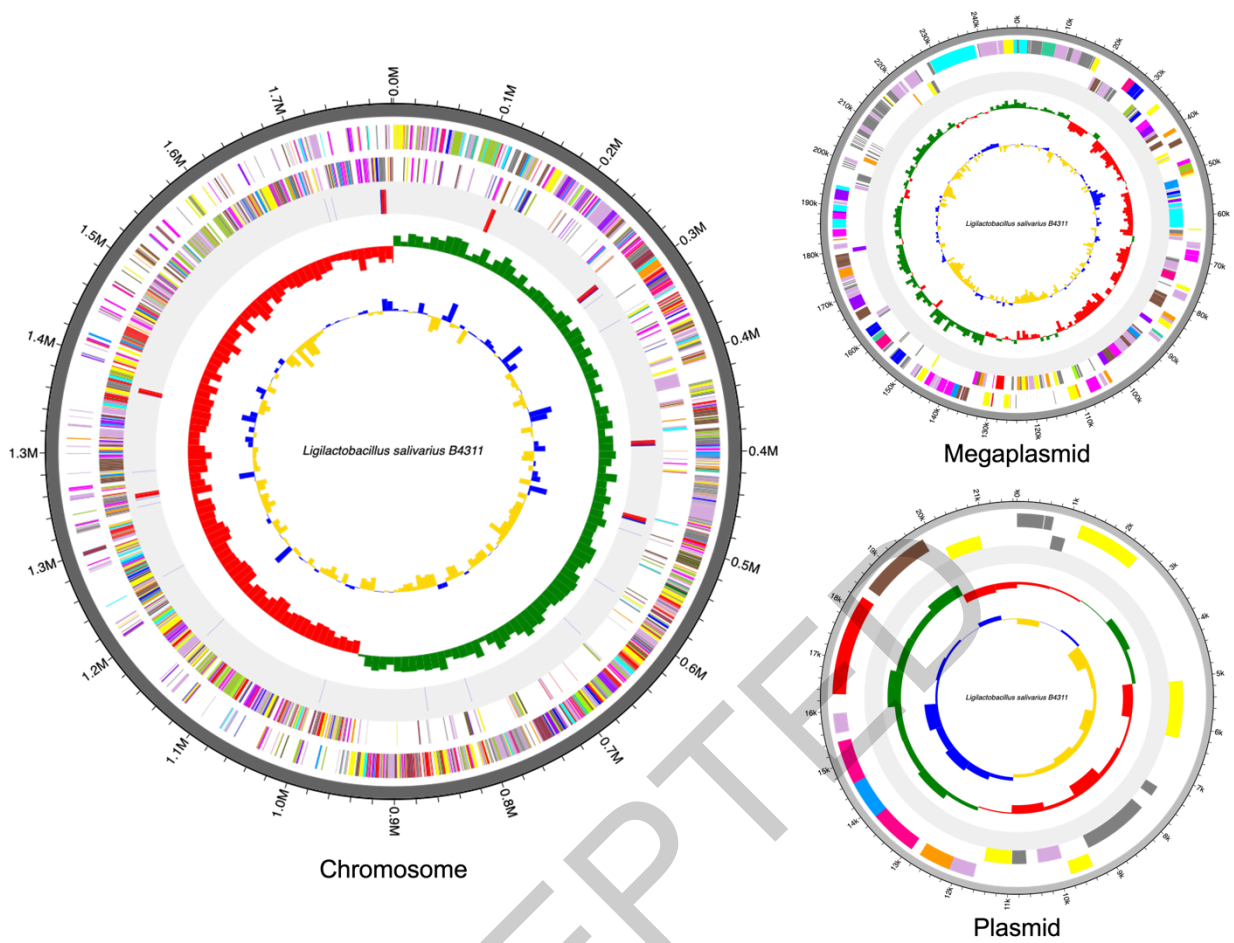
Fig. 2. Predicted bacteriocin gene cluster in *Ligilactobacillus salivarius* B4311 genome showing two ORFs for salivaricin P core peptides. ORF, open reading frame.

135 **Table 1.** Genome features of *Ligilactobacillus salivarius* B4311.

Attribute	Value			
	Chromosome	Megaplasmid	Plasmid	Total
Size (bp)	1,801,655	247,930	21,670	2,071,255
GC content (%)	33.24	32.25	33.68	33.1
No. of contigs	1	1	1	3
Total genes	1,768	273	22	2,063
Protein-coding gene	1,668	273	22	1,963
tRNA	78	-	-	78
rRNA	22	-	-	22

136
137

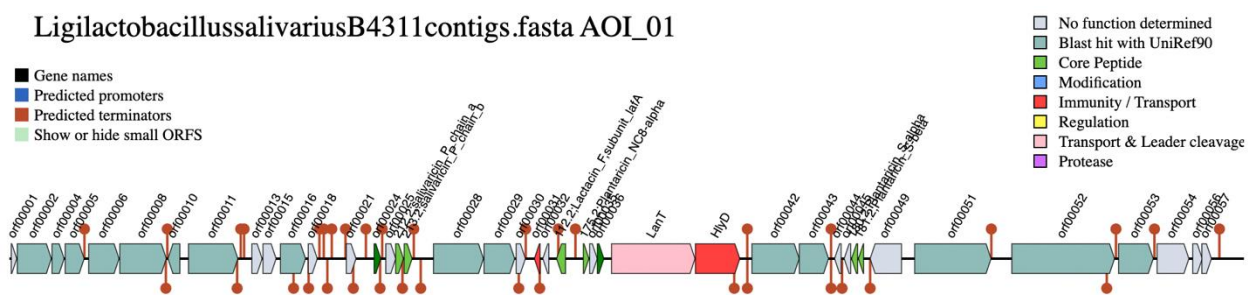
ACCEPTED



138
 139
 140
 141
 142
 143
 144
 145

Fig. 1. Circular map of *Ligilactobacillus salivarius* B4311 genome. G, guanine; C, cytosine; CDS, coding sequences. Circles represent the following characteristics from the outermost circle to the center: (1) contig information, (2) coding sequences on forward strand, (3) coding sequences on reverse strand, (4) transfer RNAs (tRNAs) and ribosomal RNAs (rRNAs), (5) GC skew, and (6) GC ratio. G, guanine; C, cytosine.

Ligilactobacillus salivarius B4311 contigs.fasta AOI_01



147

148

149 **Fig. 2.** Predicted bacteriocin gene cluster in *Ligilactobacillus salivarius* B4311 genome showing two

150 ORFs for salivaricin P core peptides. ORF, open reading frame.

151

ACCEPTED