ARTICLE INFORMATION	Fill in information in each box below
Article Type	Research article
Article Title (within 20 words without abbreviations)	Effects of phytogenic feed additives in growing and finishing pigs under different stocking density
Running Title (within 10 words)	Effects of phytogenic feed additives in high stocking density
Author	Hyun Ah Cho <sup>1#</sup> , Min Ho Song <sup>2#</sup> Ji Hwan Lee <sup>3</sup> , Han Jin Oh <sup>1</sup> , Jae Woo An <sup>1</sup> , Se Yeon Chang <sup>1</sup> , Dong Cheol Song <sup>1</sup> , Seung Yeol Cho <sup>4</sup> , Dong Jun Kim <sup>4</sup> , Mi Suk Kim <sup>4</sup> , Hyeun Bum Kim <sup>5*</sup> , Jin Ho Cho <sup>1*</sup>
Affiliation	<sup>1</sup> Department of Animal Science, Chungbuk National University, Cheongju 28644, Republic of Korea
	<sup>2</sup> Division of Animal and Dairy Science, Chungnam National University, Daejeon 34134, Republic of Korea
	<sup>3</sup> Department of Poultry Science, University of Georgia (UGA), Athens, GA, United States
	<sup>4</sup> Eugene-Bio, Suwon 16675, Republic of Korea
	<sup>5</sup> Department of Animal Resources Science, Dankook University, Cheonan 31116, South Korea
ORCID (for more information, please visit	Hyun Ah Cho (https://orcid.org/0000-0003-3469-67154)
https://orcid.org)	Min Ho Song (https://orcid.org/0000-0002-4515-5212)
	Ji Hwan Lee (https://orcid.org/0000-0001-8161-4853)
	Han Jin Oh (https://orcid.org/0000-0002-3396-483X)
	Se Yeon Chang (https://orcid.org/0000-0002-5238-2982)
	Jae Woo An (https://orcid.org/0000-0002-5602-5499)
	Dong Cheol Song (https://orcid.org/0000-0002-5704-603X)
	Seung Yeol Cho (https://orcid.org/0000-0003-3853-1053)
	Dong Jun Kim (https://orcid.org/0000-0002-1420-0527)
	Mi Suk Kim (https://orcid.org/0000-0002-9177-8701)
	Hyeun Bum Kim (https://orcid.org/0000-0003-1366-6090)
	Jin Ho Cho (http://orcid.org/0000-0001-7151-0778)
Competing interests	No potential conflict of interest relevant to this article was reported.
Funding sources	This work was carried out with the support of "Cooperative Research
State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available.	Program for Agriculture Science & Technology Development (Project No. PJ01622001)" Rural Development administration, Korea.

Acknowledgements	
Availability of data and material	Upon reasonable request, the datasets of this study can be available
	from the corresponding author.
	#Uham Ab Oba Min Ha Oann and Kilburn has sentributed anyally to
Authors' contributions	" Hyun An Cho, Min Ho Song and Ji Hwan Lee contributed equally to
Please specify the authors' role using this form.	
	Conceptualization: Cho HA, Song MH, Kim HB, Cho JH
	Data curation: Cho HA, Song MH, Loo JH
	Formal analysis: Cho HA, Song MH, Lee JH
	Investigation: Oh HJ. Chang SY. An JW. Song DC. Cho SY. Kim DJ.
	Kim MS,
	Writing - original draft: Cho HA, Song MH,
	Writing roviou & editing: Che ILL Kim LP
	Whiting - review & editing. Cho 311, Kint hb
Ethics approval and consent to participate	This article does not require IRB/IACUC approval because there are
	no human and animal participants.
2	

#### CORRESPONDING AUTHOR CONTACT INFORMATION

For the corresponding author (responsible for correspondence, proofreading, and reprints)	Fill in information in each box below
First name, middle initial, last name	<sup>1</sup> Jin Ho Cho
	<sup>2</sup> Hyeun Bum Kim
Email address – this is where your proofs will be sent	<sup>1</sup> jinhcho@chungbuk.ac.kr
	<sup>2</sup> hbkim@dankook.ac.kr
Secondary Email address	
Address	<sup>1</sup> Department of Animal Science, Chungbuk National University, Cheongju 28644, Republic of Korea
	<sup>2</sup> Department of Animal Resources Science, Dankook University, Cheonan 31116, Republic of Korea
Cell phone number	<sup>1</sup> +82-10-8014-8580
	<sup>2</sup> +82-10-3724-3416
Office phone number	<sup>1</sup> +82-43-261-2544
	<sup>2</sup> +82-41-550-3652
Fax number	<sup>1</sup> +82-43-273-2240

### 5 Abstract

6 This study was to investigate effects of different phytogenic feed additives (PFA) in grower finishing 7 pigs with stressed by high stocking density. A total of 84 growing pigs [(Landrace × Yorkshire) × Duroc] 8 with initial body weight (BW) of  $28.23 \pm 0.21$ kg were used for 10 weeks (4 replicate pens with 3 pigs 9 per pen). The dietary treatment consisted of basal diets in animal welfare density (PC, Positive control), 10 basal diet in high stocking density (NC, negative control), NC + 0.04% bitter citrus extract (PT1), NC 11 + 0.01% microencapsulated blend of thymol&carvacrol (PT2), NC + 0.10% mixture of 40% bitter citrus extract and 10% microencapsulated blend of thymol and carvacrol (PT3), NC + 0.04% premixture of 12 grape seed and grape marc extract, green tea and hops (PT4), and NC + 0.10% fenugreek seed powder 13 (PT5). The reduction of space allowance significantly decreased (P < 0.05) growth performance 14 (average daily gain, average daily feed intake, feed efficiency) and nutrient digestibility (dry matter, 15 crude protein). Also, the fecal score of NC group increased (P < 0.05) compared with other groups. In 16 blood profiles, lymphocyte decreased (P < 0.05), and neutrophil, cortisol, TNF-  $\alpha$  increased (P < 0.05) 17 18 when pigs were in high stocking density. Basic behaviors (feed intake, standing, lying) were inactive (P < 0.05) and singularity behavior (biting) were increased (P < 0.05) under high stocking density. 19 However, PFA groups alleviated the negative effects such as reducing growth performance, nutrient 20 21 digestibility, increasing stress indicators in blood and animal behavior. In conclusion, PFA 22 groupsimproved the health of pigs with stressed by high stocking density and PT3 is the most effective.

23

24 Keywords: Pig, Robustness, Additive, Stress, Plant Extract, High Stocking Density

### Introduction

26 Recently, there has been increased interest in using natural and safe feed additives to enhance robustness 27 for pigs [1,2]. Phytogenic feed additives (PFA) are plant-derived compunds such as leaves, bark, seeds, roots, flowers, twigs, tree herbs, and fruits [3]. According to the European Council, PFAs can be 28 categorized as sensory and flavoring compounds and generally feels safe as substitutes for antibiotics 29 [3]. PFAs have been recognized as the latest feed additives and antibiotics alternatives for livestock 30 31 [4,5]. Previous studies have reported that PFA complex including sunflower, thyme, and garlic can 32 improve growth performance in monogastric animals [6-8]. Rahal et al [9] have also reported that dietary PFA supplementation has immunomodulatory effects such as immunoglobulin secretion, 33 cytokine, lymphocyte expression, phagocytosis, and histamine release. Essential oils such as thymol, 34 cavacrol, cymene, terpinene reduce the pathogenic microbial load, but also promote digestive enzymes 35 thereby affecting nutrient digestibility [10-12]. Other studies have shown dietary herbs (i.e., onion, 36 fenugreek seed, and anise seed) enhanced economical efficiency to farms by improving the growth and 37 38 health of mono-gastric animals [7,13,14]. High stocking density is the most significant caused by inducing stress during growing-finishing periods. Stress caused high stocking density can reduce feed 39 intake, thereby causing low body weight gain [15-17]. Also, this stress can increase aggressive and 40 negative social behavior such as fighting, feeder occupying, tail biting [18,19] and the incidence of 41 42 body lesions [20-22]. Supplementaion of Scutellaria baicalensis L. roots mitigated negative behavior 43 caused by heat stress in mono-gastric animals [23,24]. However, studies on the relationship between 44 high stocking density and PFA have not been reported. In addition, there are few studies searching for effective PFA against stress derived from high stocking density. Therefore, the objective of this study 45 46 was to explore effective PFA against environmental stress and the exact mechanism alleviated by PFA 47 in a stress situation for grower-finishing pigs.

### **Marterial and Methods**

The experimental protocol for this study was reviewed and approved by the Institutional Animal Care
and Use Committee of Chungbuk National University, Cheongju, Korea (approval CBNUA-1530-2101).

53

#### 54 Preparation of phytogenic feed additives

PFA1 is a bitter citrus extract (BioFlavex<sup>®</sup> GC, HTBA, Beniel, Spain) that is rich in 25-27% naringin 55 and 11-15% neohesperidin. PFA2 is a microencapsulated blend of thymol and carvacrol (AviPower<sup>®</sup> 2, 56 VetAgro SpA, Reggio, Emmilia, Italy) that contains 7% of thymol and 7% carcacrol. PFA3 is a mixture 57 of PFA1, PFA2 and excipient in ratio of 4:1:5. It contains 0.7% thymol, 0.7% carvacrol,  $10 \sim 10.8\%$ 58 naringin and  $4.4 \sim 6\%$  neohesperidin. PFA4 is a premixture of grape seed & grape marc extract, green 59 tea and hops (AntaOx<sup>®</sup>FlavoSyn, DR. Eckel GmbH, Niederzissen, Germany) containing more than 10% 60 of flavonoids. PFA5 is fenugreek seed powder containing 12% saponin (Fenugreek Seed Powder, P&D 61 62 Export, Jaguar, India). All PFAs materials were provided by EUGENE BIO Co., (Suwon, South Korea).

63

### 64 Animals, housing, and experimental design

65 A total of 84 crossbred LYD ([Landrace × Yorkshire] × Duroc) mixed-sex growing pigs at 10 weeks 66 of age (average body weight  $28.23 \pm 2.89$ kg) were used in a 10-week feeding trial. Pigs were allotted 67 to one of seven treatments in a completely randomized block design based on initial body weight (BW). 68 Treatments were as follow:PC (positive control; basal diet in animal welfare density), NC (negative control; basal diet in high stocking density), PT1 (basal diet with 0.05% PFA1 in high stocking density), 69 70 PT2 (basal diet with 0.04% PFA2 in high stocking density), PT3 (basal diet with 0.10% PFA3 in high 71 stocking density), PT4 (basal diet with 0.04% PFA4 in high stocking density), PT5 (basal diet with 0.05% 72 PFA5 in high stocking density). All pigs were housed in an environmentally controlled room. There are two types of room area. In growing pig periods, animal welfare stocking density is  $0.55m^2/pig$ , high 73 stocking density is 0.40  $m^2/pig$  and in finshing pig periods, animal welfare stocking density is 1.00 74  $m^2/pig$ , high stocking density is 0.60  $m^2/pig$ . Each pen consisted of a stainless steels self-feeder and 75

nipple drinker at one-side. There are 4 replicate pens with 3 pigs per pen during the experiment period.
Basal diet was mostly consisted with corn and soybean meal and were formulated to meet or exceed
National Research Council (2012) recommendations (Table 1). During the experimental period, each
pen was equipped with a self-feeder and nipple drinker to allow *ad libitum* access to feed and water.

80

#### 81 Sampling and measuremets

#### 82 *Growth performance*

To calculate average daily gain (ADG), pig's BW was individually measured at the 09:00 on an empty
stomach at start of grower (0 weeks), end of grower and start of finisher (4 weeks), end of the finisher
(10 weeks). Feed intake and wasted feed were recorded daily to calculate average daily intake (ADFI).
Feed efficiency (G:F) was calculated by ratio of body weight gain and feed intake.

87

#### 88 Nutrient digestibility

Apparent total tract digestibility (ATTD) of dry matter (DM) and nitrogen (N) were estimated using 89 0.2% of chromic oxide as an inert indicator (Fenton & Fenton, 1979). Crude proteins (CP) were 90 measured from the nitrogen. Pigs were fed diets mixed with chromic oxide on 4<sup>th</sup> week and 10<sup>th</sup> week. 91 Fresh fecal grab samples collected via rectal massage from each pig, and these samples were stored in 92 a freezer at -20°C until analyzed. All feed and fecal samples were analyzed for DM and N following 93 94 the procedures outlined by the AOAC (2005) methods. N was determined with a Kjeltec 2300 nitrogen 95 analyzer (Foss Tecator AB, Hoeganaes Sweden) and Chromium was analyzed via UV absorption 96 spectrophotometry (Shimadzu UV-1201, Shimadzu, Kyoto, Japan) following the method described by 97 Williams, David, & Iismaa (1962). The ATTD of DM and N were calculated with indirect ratio methods using the following formula: Coefficient of apparent total tract digestibility=  $\{1-[(Nf \times Cd)/(Nd \times Cf)]\}$ 98 99  $\times$  100. Where: Nf = nutrient concentration in faces (% DM), Nd= nutrient concentration in diet (% 100 DM), Cf = chromium concentration in faeces (% DM), Cd = chromium concentration in diets (% DM).

101

102 Fecal score

During experiment, each pig fecal score was measured by same person before daily feeding. The fecal was scored according to its moisture content and shape. Normal feces are 0-point, soft feces are 1-point, mild diarrhea are 2-point and severe diarrhea are 3-point (Marquardt et al., 1999). The score was calculated by averaging each group with the average value of the daily fecal score of each pig.

107

#### 108 *Blood sample*

For the serum profile, at each pen, one pig was randomly selected to collect blood samples through 109 venipuncture at the end of 4<sup>th</sup> week, and 10<sup>th</sup> week. At the time of collection, blood samples were 110 collected both whole blood and serum in nonheparinized tubes and vacuum tubes containing K<sub>3</sub>EDTA 111 (Becton Dickinson Vacutainer systems, Franklin Lake, NJ, U.S.A.), respectively. White blood cells 112 (WBC) and WBC including lymphocyte, neutrophil, basophil concentration in whole blood were 113 114 measured using an automatic blood analyzer (ADVIA 120, Bayer, NY, USA). After collection, serum samples were centrifuged 3,000g for 15 min at 4 °C. Samples were stored at -20°C in the refrigerator 115 116 until analysis. Serum cortisol levels were assessed using enzyme-linked immunosorbent assay kits (LDN GmbH & Co., Nordhorn, Germany) following to the manufacturer's protocol. Tumor necrotizing 117 factor-alpha (TNF-α) and interukine-6 (IL-6) concentration was analyzed with ELISA kit (Quantikine, 118 R&D systems, Minneapolis, MN, USA) and they were measured at 450 nm. 119

120

### 121 *Pig behavior*

Collection of each pig image data was recorded by using six-day/night infrared cameras (QNB-7080 122 123 RH, Hanwha, Seoul, Korea) installed 3m above each pen. A total of 28 pig behaviors were analyzed by 124 randomly selecting one pig from each pen. Observers collected data based on results of Yang et al. 125 (2018), and only one person made all observations and video analysis to see consistent results. The pig 126 behavior analysis was classified for the following criteria (A) Feed intake: the act of eating with the 127 head in the feed bin, or similar behavior. (B) Standing: the act of standing still with the forelimbs and hindlimbs extended perpendicular to the floor, or similar behavior. (C) Lying: the act of lying with the 128 whole body on the floor, lying with the head, front legs, hind legs and abdomen all touching the floor. 129

130 (D) Sitting: Two front legs are spread vertically to the floor, two rear legs and two hips are sitting on 131 the floor, like a dog sitting on the floor, or something like that. (E) Drinking water: the act of drinking water for 10 seconds by putting your mouth in a drinking nipple. (F) Posture transition (lying→standing) 132 A behavior that changes from lying down to standing, in which the two front legs are stretched first, 133 and the hind legs are naturally stretched out. (G): Posture transition (standing $\rightarrow$ lying): A behavior that 134 135 changes from a standing behavior to a lying behavior, in which the two front legs are bent to the floor first, and then the two hind legs are naturally folded and lying down. (H) Rooting: the act of repeating 136 similar behaviors, such as scratches, itching, or something on the nose and front legs. (I) Biting: The 137 act of biting another pig's ears, mouth, and tail with teeth and then biting again or doing similar things. 138

139

#### 140 Statistical analysis

141 All data were analyzed by one-way ANOVA using SPSS software (ver. 20.0; IBM, USA), and the

142 differences among treatments were examined by Tukey's multiple range test, which were considered to

143 be significant at P < 0.05, unless otherwise stated.

## **Results**

145 *Growth performance* 

There was no difference between treatment groups in the initial BW of pigs (Table 2). During the 146 growing period (0-4 weeks), PT3 group significantly increased (P < 0.05) ADG and G:F ratio than NC 147 148 group. During the finishing period (4-10 weeks), NC group significantly decreased (P < 0.05) ADG and ADFI than PC group. PFA groups ADG significantly higher (P < 0.05) than NC group. The PT3-149 PT4 group ADFI significantly higher (P < 0.05) than NC group. During entire experimental period (0-150 10 weeks), NC group significantly decreased (P < 0.05) ADG, ADFI and G:F ratio than PC group. PFA 151 152 groups significantly higher (P < 0.05) ADFI than NC group. The PT3-PT4 groups significantly 153 increased (P < 0.05) ADG and G:F ratio than NC group.

154

#### 155 Nutrient digestibility

During the growing period (0-4 weeks), the ATTD of DMsignificantly increased (P < 0.05) in PT1-PT3 groups compared PC group (Table 3). The ATTD of CP significantly decreased (P < 0.05) in NC group compared toPC group. However, PFA groups significantly increased (P < 0.05) CP digestibility than NC group. During the finishing period (4-10 weeks), PFA groups decreased (P < 0.05) ATTD of DM and CP compared to NC group. The PT3-PT4 groups CP digestibility numerically increased (P < 0.05) than other PFA groups.

162

#### 163 *Fecal score*

During the growing period (0-4 weeks), NC group showed significantly higher (P < 0.05) fecal score than PC group (Table 4). However, PFA groups significantly decreased (P < 0.05) fecal score compared to NC group. During finishing period (4-10 weeks), the difference of diarrhea incidence was not observed among all treatment groups.

168

169 Blood profile

During the growing period (0-4 weeks), there were no significant difference (P > 0.05) on WBC,

171 Basophil, and IL-6 among treatment groups (Table 5). The NC group significantly decreased (P < 0.05) 172 lymphocyte and increased (P < 0.05) neutrophil, cortisol, and TNF-  $\alpha$  level in blood compared with PC group. However, PFA groups significantly alleviated (P < 0.05) these negative effects by stress with 173 stocking density and was similar with the level of PC group. During the finishing period (4-10 weeks), 174 there were no significant difference (P > 0.05) on WBC among treatment groups. NC group significantly 175 176 sdecreased (P < 0.05) lymphocyte and significantly increased (P < 0.05) neutrophil, cortisol, and TNF- $\alpha$  level in blood compared to PC group. However, PFA groups significantly increased (P < 0.05) 177 lymphocyte and significantly decreased (P < 0.05) neutrophil, cortisol, and TNF-  $\alpha$  compared with NC 178 179 group. PT3 group showed (P < 0.05) the lowest results in neutrophil, cortisol, and IL-6 among PFA 180 groups.

181

#### 182 Animal behavior

The effects of different PFA on animal behavior were shown in Table 6, Table 7, Figure 1. During the growing period (0-4 weeks), there are no significant difference (P > 0.05) in basic behavior and most of singularity behavior. The NC group had significantly higher (P < 0.05) biting frequency than PC group. However, PFA groups had significantly alleviated (P < 0.05) biting frequency compared with NC group. Among PFA groups, PT3 group showed the lowest biting frequency.

During the finishing period (4-10 weeks), NC group showed (P < 0.05) more lying time and less feed intake and standing time than PC group. Feed intake time significantly increased (P < 0.05) in PFA groups than NC group. Standing time significantly increased (P < 0.05) in PT2-PT5 group than NC group. Lying time significantly decreased (P < 0.05) in PFA groups than NC group. Especially, PT3-PT5 groups showed similar result with PC group. In singularity behavior, there are no significant difference (P > 0.05) in treatment groups. But NC group showed numerically high number of biting than other treatment groups.

### Discussion

196 *Growth performance* 

197 High stocking density can disturb the movement of animals due to limited feeding environment (space, feeders, and drinkers). Moreover, high stocking density can interfere with airflow and generate heat 198 199 energy [12]. It can result in difficulty in evacuating body temperature, poor air quality, reduced access 200 to feed and water, and poor performance of animals due to increased ammonia levels [28-30]. High heat 201 energy and poor air quality are known to cause heat stress and adverse effects on growth rate, feed 202 consumption, mortality, and health [31-33]. Similarly, our study showed that pigs under high stocking density (i.e., 0.40 m<sup>2</sup>/ growing pig, 0.60 m<sup>2</sup>/finishing pig) had reduced ADG and ADFI by 16.38% and 203 11.24%, respectively, than those under welfare density (i.e.,  $0.55 \text{ m}^2/\text{growing pig}$ ,  $1.0 \text{ m}^2/\text{finishing pig}$ ) 204 during the whole period (grower: 28-56 kg, finisher: 56-103 kg). Spicer and Aherne [34] have also 205 reported that daily gain and daily feed are reduced 8.47% and 13.15%, respectively, when group size is 206 decreased from 0.72 m<sup>2</sup>/pig to 0.35 m<sup>2</sup>/pig. Stress-induced heat and high stocking densities can reduce 207 208 growth performance by damaging cellular structure, increasing intracellular water imbalance, and increasing free radical concentration [35]. However, our study revealed that pigs under high stocking 209 density with supplementation of PFA showed improvement (i.e., BW decreased 11.61%) in growth 210 performance compared to those in the unsupplemented group. Many researchers reported that dietary 211 supplementation of PFA such as Korean pine extract, cinnamon, turmeric, essential oils, and rosemary 212 213 can improve growth performance with reducing stress response [35-38]. In our study, PFA 214 supplementation under our high stocking density showed no difference in ADFI between treatments in the growing period, but significantly increased with PFA supplementation in the finishing period This 215 216 is consistent with previous studies suggesting that PFA is effective for intake when supplied long-term 217 [39]. Moreover, PFA3 could improve the flavor of feed and increase the palatability of feed intake in pigs [40-43]. Therefore, using natural products with polyphenols (suitable structure for free radical 218 219 scavenging activity) can effectively alleviate stress caused by low space allowance and heat through 220 their antioxidant activity with improved low feed intake, thereby increasing growth performance.

195

#### 222 Nutrient digestibility

223 High stocking density can negatively affect nutrient digestibility and growth performance. During the whole experiment periods, nutrient digestibility (DM, CP) showed improvement in the treatment group 224 225 added with PFA than that in the control group without PFA under high stocking density. PFA can also 226 enhance nutrient digestibility and absorption [44,45]. It has been reported that the addition of essential 227 oils to monogastric animals can enhance the activity of trypsin, maltase and pancreatic amylase and 228 increases glucose absorption in the small intestine [46]. Therefore, the addition of PFA can stimulate 229 the secretion of mucus in the intestine, thereby reducing the adhesion of pathogens and stabilizing 230 intestinal microbial symbiosis [47]. It can be seen that improved digestive tract function is associated 231 with increased nutrient digestibility. It can also be said that the antibacterial action of PFA contributes 232 to the increase of nutrient digestibility. PFAs such as carvacrol, thymol, anetol, oregano, anise, and 233 citrus essential oil have antibacterial activity against intestinal microbes when ingested. Among them, phenolic substances are the most active compounds [48,49]. PT2 and PT3 have a phenolic structure in 234 235 our experiment. It was shown that the digestibility of DM and CP was higher than the high stocking density throughout the experiment period. Fiesel et al [50] reported an increase in nutrient digestibility 236 due to the antioxidant effect of polyphenols and an increased absorbable surface of the intestine. As the 237 experiment progressed, the digestibility deviation of DM and CP increased according to the presence 238 239 or absence of PFA in the feed under high stocking density. In this experiment, it was confirmed that the 240 digestibility was gradually improved when PFA was used, leading to improved performance of pigs. In 241 particular, it was found that the digestibility was significantly improved by flavonoids, a common 242 component of PT3-PT5 additives. A previous study has shown that flavonoids have DM and CP 243 synergistic effects [51]. Therefore, it can be concluded that the use of flavonoid additives can increase 244 the digestibility of nutrients, as it can improve nutrient availability by boosting immunity and 245 antibacterial action in pigs.

246

247 Fecal score

248 In high stocking density, the frequency of diarrhea was increased during the growing period, although

249 it showed no significant difference during the finishing period. Many studies have found that diarrhea 250 in pigs is more likely to be induced by stress [52-54]. Actually, the frequency of diarrhea is increased 251 in weaned pigs during stress [55]. Diarrhea has been found intermittently in growing pigs [56]. When pigs get stressed, their immunity is lower and pathogens in the intestine are activated. Intestinal 252 pathogens can suppress unnecessary energy loss such as reduced feed intake and G:F ratio known to 253 254 interfere with immune system activation. In addition, intestinal pathogens can inhibit homeostasis of 255 the epithelial barrier, causing secretory diarrhea due to intestinal damage through osmotic stress or 256 inflammatory diarrhea by increasing inflammatory cytokines. However, in our study, the frequency of 257 diarrhea was significantly reduced when PFAs were fed to pigs in a stressful situation. These results 258 indicate that PFA can improve fecal status by improving intestinal health, and further studies on fecal 259 microflora should be conducted. When pigs are fed with natural products reduces the frequency of 260 diarrhea due to stress as the natural product's antibacterial action improves intestinal health and increased digestibility [57,58]. Many researchers have checked diarrhea scores of weaning pigs, but not 261 262 those of growing to finishing pigs. In the present study, complete diarrhea was not found even in the growing period, although a lot of soft feces were observed for pigs under a high stocking density 263 condition. The difference between growing period and finishing period is that as pigs grow, their 264 immune system gets better, and their gut health improves. Therefore, we can confirm a meaningful 265 266 diarrhea score even in pigs during the growing period. Thus, it is necessary to check the status of feces.

267

#### 268 Blood profile

In the present study, there were no significant differences in blood profile between the entire experiment period WBC or growing period basophil and IL-6 of pigs between treatment groups. However, pigs fed with PFAs under a high stocking density condition had better blood results than those without addition of additives under a high stocking density condition. Pigs with high stocking density are subjected to critical psychological, social, and environmental stresses. High stocking density can also cause chronic severe stress that affects immunity and health [60]. Lymphocytes show various immunological responses including modulation of immune defense and immunoglobulin [61]. In our study, 276 lymphocytes were decreased during stress situation, but returned to PC levels when PFAs were added. 277 According to Dhabhar [62], in stressful situations, lymphocyte counts are decreased due to changes 278 induced by trafficking or redistribution of lymphocytes to other body compartments of glucocorticoids. 279 This result was similar to our study. In our study, the number of neutrophils was increased when pigs 280 were stressed. This number was then decreased after supplementation with PFA in our study. It has been 281 reported that stressful situations cause decreasing lymphocytes and increasing neutrophils in the blood 282 [63]. As a result, it was possible to confirm the indirect change caused by supplementation of PFA to 283 relieve stress. Cortisol, a steroid hormone, or glucocorticoid produced by the adrenal gland and released 284 in response to stress, is often used as a physiological marker to quantify animal stress [64]. It is well 285 known that cortisol can regulate intermediary metabolism, immunity, and growth [65,66]. A poor 286 welfare situation can cause animals to be extremely stressed. In this study, cortisol level was increased 287 under high stocking density compared with animal welfare density (space decreased in growing pig 27.27%, in finshing pig 40%). This result was agreement with the results of Jang et al. [67] that reported 288 289 decreasing space allowance (decreasing 28.13%) induced increasing cortisol level (2.3µg/dL to 4µg/dL). 290 However, PFAs supplementation alleviated high cortisol level in blood caused by high stocking density. Li et al [67] observed that flavonoids, which are physiologically active substances of PFA, down-291 regulated immune responses by mediated viruses and the T-cell, thereby reducing psychological stress. 292 293 This observation suggests that PFA mitigates the increased cortisol concentration by high stocking 294 density.

295 Pro-inflammatory cytokines such as TNF- $\alpha$  and IL-6 are potential outputs of the cellular immune system 296 and can indirectly reflect immune responses due to the activation of T-cells [68,69]. This study showed 297 that high stocking density increased pro-inflammatory cytokine level. These results suggest that the 298 environmental stress caused by a limited space allowance can induce a cellular immune response. When 299 stressed out, pro-inflammatory cytokines are secreted to promote cortisol secretion and suppress growth 300 hormone secretion [70,71]. Excessive pro-inflammatory cytokines can induce fever, inflammation, 301 tissue destruction [72], and in some cases, even shock and death (Dinarello, 2000). Thus, the immune 302 system is activated due to high stocking stress, which shifts nutrient distribution priorities from growth 303 to host defense [68,70,74].

In addition, TNF- $\alpha$  and IL-6 content is reduced through improved gut microbiota, antioxidant, and antiinflammatory effects, due to improved digestibility of nutrients, alleviating stress response, and strengthening immunity [75,76]. However, PFA was effective methods to alleviate negative effects of a high stocking density in our study. Other researchers also reported essential oil and herb extract reduced pro-inflammatory cytokines [77,78].

Therefore, PFA is effective in relieving stress, and PT3 group showed the highest effect among PFAs group. The reason the PT3 group outperformed the others was due to the construction of the PFA group. Flavonoids and terpenoids (carbacrol and thymol) may protect cells from the harmful effects of autoxidation.

313

#### 314 Animal behavior

A high stocking density equates to a reduced floor space allowance. Decreasing floor space allowance 315 per pig increases the frequency of contact, social tension, and aggression [79-82]. In addition, when 316 heat production per unit floor area is increased, heat stress will occur and induce oxidative stress [82,22]. 317 If this stress is not well managed in pigs, it can increase their susceptibility to stress and hence reduce 318 319 their immune and health status. Throughout our study, animal behavior at high stocking density 320 improved when fed with PFA. The biting frequency was increased in NC but decreased after PFA 321 treatment similar to PC. Among all treatment groups, PT3 group showed the lowest biting frequency. 322 Greene et al [35] has reported that biting as a representative form of aggressive behavior can occur in pigs under chronic stress. This is consistent with our study. When ingesting phenolic compounds as 323 324 components of PT3 group, it is possible to restore redox homeostasis and prevent oxidative stress by 325 improving the activity of antioxidant enzymes SOD, CAT, GPx, and GR [84]. Therefore, the effect of 326 adding PFA3 not only can help pigs cope with biting behavior caused by stress, but also can overcome it. During the finishing period, basic behaviors (eating, standing, lying down) were more active when 327 fed with PFA added in high stocking density. In addition, the feed intake increased during PFA feeding 328 329 in growth performance. Feed intake is an important indicator because it is related to body weight, ADG,

330 ADFI, and G: F ratio. Pigs with a high stocking density face difficulty in feeding due to competition in 331 the feeder. In this study, PC group showed less time than other treatments in feed intake. Therefore, the number of trips to the feeder is directly related to intake and can affect growth performance. Also, 332 333 standing and lying time were similar to NC group. They were more active than PC group. Especially, PT3-PT5 groups are more activated than others. Pearce and Paterson [85] have reported that observation 334 335 of the behavior of standing motionless in a narrow space is a behavior that pigs do to cope with stress at a high stocking density. As stress increased, the amount of physical activity decrease. It can be seen 336 337 that when the standing time decrease, the lying time increases at the same time. This indicates that there is a close relationship between basic behavior and growth performance. Through this experiment, it can 338 339 be seen that when pigs get stressed, their basic behaviors (standing, lying, and feeding) were affected at the same time. 340

# Conclusion

343	Dietary supplementation of PFA improves the growth performance, nutrient digestibility, immunity,
344	fecal score, and animal behavior in grower-finishing pigs. As a result, lymphocytes, neutrophils, cortisol,
345	IL-6, and TNF- $\alpha$ in the blood, bites, and basic behaviors were improved, indicating that stress was
346	reduced and strengthened. The diarrhea index improved because of getting healthier, which means less
347	damage to the intestines and increased digestibility. Due to these positive effects, growth performance
348	was improved, and it was found that PFA is an effective additive for stress due to high stocking density.
349	Among them, the most effective and additional advantages were found when using PFA3 (mixture of
350	PFA1 40%, PFA2 10% and excipient 50%) rather than using PFA1 (bitter citrus extract) and PFA2
351	(microencapsulated blend of thymol and carvacrol) separately.

## 352 **REFERENCE**

- Kholif, AE, Abdo MM, Anele UY, El-Sayed MM, Morsy TA. Saccharomyces cerevisiae does not work
   synergistically with exogenous enzymes to enhance feed utilization, ruminal fermentation and lactational
   performance of Nubian goats. *Livest Sci*, 2017; 206, 17-23. https://doi.org/10.1016/j.livsci.2017.10.002
- Kholif AE, Gouda GA, Galyean ML, Anele UY, Morsy TA. Extract of Moringa oleifera leaves increases milk production and enhances milk fatty acid profile of Nubian goats. *Agrofor Syst, 2019; 93*(5), 1877-1886. https://doi.org/10.1007/s10457-018-0292-9
- European Union. Council directive 2008/120/EC of 18 December 2008 laying down minimum standards for the protection of pigs (codified version). Off J Eur Union. 2009;L47:5-13.
- Ebeid HM., Mengwei L, Kholif AE, Hassan FU, Lijuan P, Xin L, Chengjian Y. Moringa oleifera oil modulates rumen microflora to mediate in vitro fermentation kinetics and methanogenesis in total mix rations. Curr Microbiol, 2020; 77, 1271-1282. https://doi.org/10.1007/s00284-020-01935-2
- 5. Elghalid OA, Kholif AE, El-Ashry GM, Matloup OH, Olafadehan OA, El-Raffa A M, Abd El-Hady AM.
  Oral supplementation of the diet of growing rabbits with a newly developed mixture of herbal plants and
  spices enriched with special extracts and essential oils affects their productive performance and immune
  status. *Livest. Sci*, 2020; 238, 104082. https://doi.org/10.1016/j.livsci.2020.104082
- Al-Gharabi HK., Al-Gharawi JK, Al-Sahlani AJ. EFFECT OF GARLIC (ALLIUM SATIVUM) AND
   ONION (ALLIUM CEPA) WATER EXTRACT ON SOME PRODUCTIVE TRAITS OF BROILERS. Plant
   Arch, 2019; 19(1), 565-569.
- Alhajj MS, Alhobaishi M, El Nabi AG, Al-Mufarrej SI. Immune responsiveness and performance of broiler chickens fed a diet supplemented with high levels of Chinese star anise fruit (Illicium verum Hook. F). J Anim Vet Adv, 2015; 14(2), 36-42. https://doi.org/10.36478/javaa.2015.36.42
- Khan RU, Nikousefat Z, Tufarelli V, Naz S, Javdani M, Laudadio V. Garlic (Allium sativum)
   supplementation in poultry diets: Effect on production and physiology. *Worlds Poult Sci J, 2012; 68*(3), 417 424. https://doi.org/10.1017/S0043933912000530
- Rahal A, Deb R, Latheef SK, Tiwari R, Verma AK, Kumar A, Dhama K. Immunomodulatory and therapeutic potentials of herbal, traditional/indigenous and ethnoveterinary medicines. Journal of biological sciences : JBS, 2012; 15(16), 754-774.
- Ahmed ST, Hossain ME, Kim GM, Hwang JA, Ji H, Yang CJ. Effects of resveratrol and essential oils on
   growth performance, immunity, digestibility and fecal microbial shedding in challenged piglets. Asian Australas J Anim Sci, 2013; 26(5), 683. https://doi.org/10.5713/ajas.2012.12683
- 11. Emami NK, Samie A, Rahmani HR, Ruiz-Feria CA. The effect of peppermint essential oil and fructooligosaccharides, as alternatives to virginiamycin, on growth performance, digestibility, gut morphology and immune response of male broilers. *Anim Feed Sci Technol, 2012; 175*(1-2), 57-64. https://doi.org/10.1016/j.anifeedsci.2012.04.001

- Zhang S, Jung JH, Kim HS, Kim BY, Kim IH. Influences of phytoncide supplementation on growth
   performance, nutrient digestibility, blood profiles, diarrhea scores and fecal microflora shedding in weaning
   pigs. Asian-Australas J Anim Sci, 2012; 25(9), 1309. https://doi.org/10.5713/ajas.2012.12170
- Aji SB, Ignatius K, Ado AY, Nuhu JB, Abdulkarim A, Aliyu U, Numan PT. Effect of feeding onion (Allium cepa) and garlic (Allium sativum) on some performance characteristics of broiler chickens. Res. J. Poult. Sci, 2011; 4, 22-27. https://doi.org/ 10.3923/rjpscience.2011.22.27
- Mamoun T, Mukhtar MA, Tabidi MH. Effect of fenugreek seed powder on the performance, carcass
   characteristics and some blood serum attributes. Adv. Res. Agri. Vet. Sci, 2014; 1(1), 6-11.
- Section 395
   Section 395
   Cho JH, Kim IH. Effect of stocking density on pig production. *Afr J Biotechnol*, 2011; 10(63), 13688-13692.
   https://doi.org/10.5897/AJB11.1691
- Hyun Y, Ellis M, Riskowski G, Johnson RW. Growth performance of pigs subjected to multiple concurrent environmental stressors. *J Anim Sci, 1998; 76*(3), 721-727. https://doi.org/10.2527/1998.763721x
- McGlone JJ, Salak JL, Lumpkin EA, Nicholson RI, Gibson M, Norman RL. Shipping stress and social status effects on pig performance, plasma cortisol, natural killer cell activity, and leukocyte numbers. J Anim Sci, 1993; 71(4), 888-896. https://doi.org/10.2527/1993.714888x
- Li Y, Wang C, Huang S, Liu Z, Wang H. Space allowance determination by considering its coeffect with toy provision on production performance, behavior and physiology for grouped growing pigs. *Livest. Sci*, 2021; 243, 104389. https://doi.org/ 10.1016/j.livsci.2020.104389
- 405 19. Cornale P, Macchi E, Miretti S, Renna M, Lussiana C, Perona G, Mimosi A. Effects of stocking density and 406 environmental enrichment on behavior and fecal corticosteroid levels of pigs under commercial farm 407 conditions. J Vet Behav, 2015; 10(6), 569-576. https://doi.org/10.1016/j.jveb.2015.05.002
- 408 20. Averos X, Brossard L, Dourmad JY, de Greef KH, Edge HL, Edwards SA, Meunier-Salaun MC. Quantitative assessment of the effects of space allowance, group size and floor characteristics on the lying behaviour of growing-finishing pigs. *Animal*, 2010; 4(5), 777–783. https://doi.org/10.1017/S1751731109991613
- 411 21. Hyun Y, Ellis M, Curtis SE, Johnson RW. Environmental temperature, space allowance, and regrouping:
   412 additive effects of multiple concurrent stressors in growing pigs. J Swine Health Prod. 2005;13:131-8.
- 413 22. Street BR, Gonyou HW. Effects of housing finishing pigs in two group sizes and at two floor space allocations
  414 on production, health, behavior, and physiological variables. J Anim Sci, 2008; 86(4), 982-991.
  415 https://doi.org/10.2527/jas.2007-0449
- 416 23. Casal-Plana N, Manteca X, Dalmau A, Fàbrega E. Influence of enrichment material and herbal compounds
  417 in the behaviour and performance of growing pigs. Appl Anim Behav Sci. 2017;195:38-43.
  418 https://doi.org/10.1016/j.applanim.2017.06.002
- 24. Zmrhal V, Lichovníková M, Hampel D. The Effect of Phytogenic Additive on Behavior During Mild Moderate Heat Stress in Broilers. Acta Univ Agric Fac Agron. 2018;

- 421 https://doi.org/10.11118/actaun201866040939
- 422 25. AOAC [Association of Official Analytical Collaboration] International. Official methods of analysis of
   423 AOAC International. 21st ed. Gaithersburg, MD: AOAC International; 2019.
- 424 26. Marquardt RR, Jin LZ, Kim JW, Fang L, Frohlich AA, Baidoo SK. Passive protective effect of egg-yolk
  425 antibodies against enterotoxigenic Escherichia coli K88+ infection in neonatal and early-weaned piglets.
  426 FEMS Immunol Med Microbiol. 1999;23:283-8. https://doi. org/10.1111/j.1574-695X.1999.tb01249.x
- Yang KY, Jeon JH, Kwon KS, Choi HC, Ha JJ, Kim JB, et al. Classification of behavior at the signs of
  parturition of sows by image information analysis. J Korea Acad Ind Soc. 2018;19:607-13.
  https://doi.org/10.5762/KAIS.2018.19.12.607
- Bilal RM, Hassan FU, Farag MR, Nasir TA, Ragni M, Mahgoub HA, et al. Thermal stress and high stocking
  densities in poultry farms: potential effects and mitigation strategies. J Therm Biol. 2021;99:102944.
  https://doi.org/10.1016/j.jtherbio.2021.102944
- 29. European Food Safety Authority (EFSA). Opinion of the Scientific Panel on Animal Health and Welfare 433 434 (AHAW) on a request from the Commission related to welfare of weaners and rearing pigs: effects of 435 EFSA different space allowances and floor. Journal, 2005; 3(10), 268, 436 https://doi.org/10.2903/j.efsa.2005.268
- 437 30. Feddes JJ, Emmanuel EJ, Zuidhoft MJ. Broiler performance, body weight variance, feed and water intake,
  438 and carcass quality at different stocking densities. Poult Sci. 2002;81:774-9.
  439 https://doi.org/10.1093/ps/81.6.774
- 440 31. Chegini S, Kiani A, Parizadian Kavan B, Rokni H. Effects of propolis and stocking density on growth
  441 performance, nutrient digestibility, and immune system of heat-stressed broilers. Ital J Anim Sci, 2019; 18(1),
  442 868-876. https://doi.org/10.1080/1828051X.2018.1483750
- 443 32. Estévez M. Oxidative damage to poultry: from farm to fork. Poult Sci. 2015;94:1368-78.
  444 https://doi.org/10.3382/ps/pev094
- 33. Sohail MU, Ijaz A., Yousaf MS, Ashraf K, Zaneb H, Aleem M, Rehman H. Alleviation of cyclic heat stress
  in broilers by dietary supplementation of mannan-oligosaccharide and Lactobacillus-based probiotic:
  Dynamics of cortisol, thyroid hormones, cholesterol, C-reactive protein, and humoral immunity. Poult
  Sci, 2010; 89(9), 1934-1938. https://doi.org/10.3382/ps.2010-00751
- 449 34. Spicer HM, Aherne FX. The effects of group size/stocking density on weanling pig performance and behavior. Appl Anim Behav Sci, 1987; 19(1-2), 89-98. https://doi.org/10.1016/0168-1591(87)90206-1

35. Greene ES, Cauble R, Kadhim H, de Almeida Mallmann B, Gu I, Lee SO, et al. Protective effects of the phytogenic feed additive "comfort" on growth performance via modulation of hypothalamic feeding- and drinking-related neuropeptides in cyclic heat-stressed broilers. Domest Anim Endocrinol. 2021;74:106487.
454 https://doi.org/10.1016/ j.domaniend.2020.106487

- 455 36. Li HL, Zhao PY, Lei Y, Hossain MM, Kang J, Kim IH. Dietary phytoncide supplementation improved growth
  456 performance and meat quality of finishing pigs. Asian-Australas J Anim Sci, 2016; 29(9), 1314.
  457 https://doi.org/10.5713/ajas.15.0309
- 458 37. Hashemzadeh F, Rafeie F, Hadipour A, Rezadoust MH. Supplementing a phytogenic-rich herbal mixture to heat-stressed lambs: Growth performance, carcass yield, and muscle and liver antioxidant status. Small
  460 Rumin Res, 2022; 206, 106596. https://doi.org/10.1016/j.smallrumres.2021.106596
- 38. Yan L, Meng QW, Kim IH. Effect of an herb extract mixture on growth performance, nutrient digestibility,
  blood characteristics, and fecal microbial shedding in weanling pigs. *Livest Sci*, 2012; 145(1-3), 189-195.
  https://doi.org/10.1016/j.livsci.2012.02.001
- Bartoš P, Dolan A, Smutný L, Šístková M, Celjak I, Šoch M, et al. Effects of phytogenic feed additives on growth performance and on ammonia and greenhouse gases emissions in growing-finishing pigs. Anim Feed Sci Technol. 2016;212:143-8. https://doi.org/10.1016/j.anifeedsci.2015.11.003
- 40. Mucha W, Witkowska D. The applicability of essential oils in different stages of production of animal-based foods. Molecules, 2010; 26(13), 3798. https://doi.org/10.3390/molecules26133798
- 469 41. Czech A, Kowalczuk E, Grela E. The effect of a herbal extract used in pig fattening on the animals. Ann Univ Mariae Curie Sklodowska Sect EE Zootech, 2009; 27(2). https://doi.org/10.2478/v10083-009-0009-7
- 471 42. Wenk C. Herbs and botanicals as feed additives in monogastric animals. Asian-Australas J Anim Sci, 2003;
  472 16(2), 282-289. https://doi.org/10.5713/ajas.2003.282
- 473 43. FrAnKIČ T, Voljč M, Salobir J, Rezar V. Use of herbs and spices and their extracts in animal nutrition. Acta Agric Slov, 2009; 94(2), 95-102.
- 44. Amad AA, Männer K, Wendler KR, Neumann K, Zentek J. Effects of a phytogenic feed additive on growth
  performance and ileal nutrient digestibility in broiler chickens. Poult Sci, 2011; 90(12), 2811-2816.
  https://doi.org/10.3382/ps.2011-01515
- 478 45. Hafeez A, Männer K, Schieder C, Zentek J. Effect of supplementation of phytogenic feed additives
  479 (powdered vs. encapsulated) on performance and nutrient digestibility in broiler chickens. Poult Sci,
  480 2016; 95(3), 622-629. https://doi.org/10.3382/ps/pev368
- 481 46. Lee KW, Everts H, Kappert HJ, Frehner M, Losa R, Beynen AC. Effects of dietary essential oil components
  482 on growth performance, digestive enzymes and lipid metabolism in female broiler chickens. Br Poult
  483 Sci, 2003; 44(3), 450-457. https://doi.org/10.1080/0007166031000085508
- 484 47. Jamroz D, Wertelecki T, Houszka M, Kamel C. Influence of diet type on the inclusion of plant origin active substances on morphological and histochemical characteristics of the stomach and jejunum walls in chicken. J Anim Physiol Anim Nutr, 2006; 90(5-6), 255-268. https://doi.org/10.1111/j.1439-0396.2005.00603.x
- 488 48. Giannenas I, Bonos E, Christaki E, Florou-Paneri P. Essential oils and their applications in animal nutrition.

- 489 Med Aromat Plants, 2013; 2(140), 2167-0412.
- 49. Panghal M, Kausha V, Yadav JP. In vitro antimicrobial activity of ten medicinal plants against clinical isolates of oral cancer cases. Ann Clin Microbiol Antimicrob, 2011; 10(1), 1-11. https://doi.org/10.1186/1476-0711-10-21
- 50. Fiesel A, Gessner DK, Most E, Eder K. Effects of dietary polyphenol-rich plant products from grape or hop on pro-inflammatory gene expression in the intestine, nutrient digestibility and faecal microbiota of weaned pigs. *BMC Vet Res, 2014; 10*(1), 1-11. https://doi.org/10.1186/s12917-014-0196-5
- 496 51. Cui K, Wang Q, Wang S, Diao Q., & Zhang, N. (2019). The facilitating effect of tartary buckwheat flavonoids
  497 and Lactobacillus plantarum on the growth performance, nutrient digestibility, antioxidant capacity, and fecal
  498 microbiota of weaned piglets. *Animals*, 9(11), 986. https://doi.org/10.3390/ani9110986
- 52. Oh HJ, Park YJ, Cho JH, Song MH, Gu BH, Yun W, Kim MH. Changes in diarrhea score, nutrient digestibility,
   zinc utilization, intestinal immune profiles, and fecal microbiome in weaned piglets by different forms of
   zinc. Animals, 2021; 11(5), 1356. https://doi.org/10.3390/ani11051356
- 502 53. Yang KM, Jiang ZY, Zheng CT, Wang L, Yang XF. Effect of Lactobacillus plantarum on diarrhea and intestinal barrier function of young piglets challenged with enterotoxigenic Escherichia coli K88. J Anim Sci, 2014; 92(4), 1496-1503. https://doi.org/10.2527/jas.2013-6619
- 54. Zhao Y, Weaver AC, Fellner V, Payne RL, Kim SW. Amino acid fortified diets for weanling pigs replacing
  fish meal and whey protein concentrate: Effects on growth, immune status, and gut health. J Anim Sci
  Biotechnol, 2014; 5(1), 1-10. https://doi.org/10.1186/2049-1891-5-57
- 508 55. Moeser AJ, Blikslager AT. Mechanisms of porcine diarrheal disease. J Am Vet Med Assoc. 2007;231:56-67. https://doi.org/10.2460/javma.231.1.56
- 56. Panah FM, Lauridsen C, Højberg O, Nielsen TS. Etiology of Colitis-Complex Diarrhea in Growing Pigs: A
   Review. Animals, 2021; 11(7), 2151. https://doi.org/10.3390/ani11072151
- 57. Cho JH, Chen YJ, Min BJ, Kim HJ, Kwon OS, Shon KS, Asamer A. Effects of essential oils supplementation
   on growth performance, IgG concentration and fecal noxious gas concentration of weaned pigs. *Asian- Australas J Anim Sci, 2005; 19*(1), 80-85. https://doi.org/10.5713/ajas.2006.80
- 515 58. Caprarulo V, Turin L, Hejna M, Reggi S, Dell'Anno M, Riccaboni P, Rossi L. Protective Effect of Phytogenic
  516 based Additives in Enterotoxigenic Escherichia Coli Challenged Piglets, 2022;
  517 https://doi.org/10.21203/rs.3.rs-1207181/v1
- 518 59. Griffin JFT. Stress and immunity: a unifying concept. *Vet Immunol Immunopathol, 1989; 20*(3), 263-312.
   519 https://doi.org/10.1016/0165-2427(89)90005-6
- 60. Campbell TW. Clinical pathology. Reptile Medicine and Surgery (ed. D.R. Mader), W.B. Saunders Company,
   Philadelphia, 1996; pp. 248-257

- 522 61. Dhabhar FS. Stress-induced augmentation of immune function—the role of stress hormones, leukocyte
  523 trafficking, and cytokines. *Brain Behav Immun, 2002; 16*(6), 785-798. https://doi.org/10.1016/S0889524 1591(02)00036-3
- 525 62. Demir S, Atli A, Bulut M, İbiloğlu AO, Güneş M, Kaya MC, Sır A. Neutrophil–lymphocyte ratio in patients
   526 with major depressive disorder undergoing no pharmacological therapy. Neuropsychiatr Dis Treat, 2015; 11,
   527 2253. https://doi.org/ 10.2147/NDT.S89470
- Warriss PD, Brown SN, Edwards JE, Knowles TG. Effect of lairage time on levels of stress and meat quality
   in pigs. *Animal science*, 1998; 66(1), 255-261. https://doi.org/10.1017/S1357729800009036
- 64. Oyarzún R, Paredes R, Saravia J, Morera FJ, Muñoz JLP, Ruiz-Jarabo I, Vargas-Chacoff L. Stocking density
  affects the growth performance, intermediary metabolism, osmoregulation, and response to stress in
  Patagonian blennie Eleginops maclovinus. Aquaculture, 2020; 515, 734565.
  https://doi.org/10.1016/j.aquaculture.2019.734565
- 65. Chrousos GP, Kino T. Glucocorticoid action networks and complex psychiatric and/or somatic disorders. *Stress*, 2007; 10(2), 213-219. https://doi.org/10.1080/10253890701292119
- 536 66. Jang JC, Jin XH, Hong JS, Kism YY. Effects of different space allowances on growth performance, blood
  537 profile and pork quality in a grow-to-finish production system. Asian-Australas J Anim Sci, 2017; 30(12),
  538 1796. <u>https://doi.org/10.5713/ajas.17.0076</u>
- 539 67. Li, H. L., Zhao, P. Y., Lei, Y., Hossain, M. M., Kang, J., and Kim, I. H. Dietary phytoncide supplementation improved growth performance and meat quality of finishing pigs. Asian-Australasian Journal of Animal Sciences, 2016; 29(9), 1314. https://doi.org/10.5713/ajas.15.0309
- 542 68. Colditz IG. Effects of the immune system on metabolism: implications for production and disease resistance
   543 in livestock. Livest Prod Sci, 2002; 75(3), 257-268. https://doi.org/10.1016/S0301-6226(01)00320-7
- Fossum C. Cytokines as markers for infections and their effect on growth performance and well-being in the
   pig. Domest Anim Endocrinol, *1998*; *15*(5), 439-444. https://doi.org/10.1016/s0739-7240(98)80001-5
- Fan J, Molina PE, Gelato MC, Lang CH. Differential tissue regulation of insulin-like growth factor-I content
  and binding proteins after endotoxin. *Endocrinology*, 1994; 134(4), 1685-1692.
  https://doi.org/10.1210/en.134.4.1685
- 549 71. Johnson OL, Jaworowicz W, Cleland JL, Bailey L, Charnis M, Duenas E, Putney SD. The stabilization and
  550 encapsulation of human growth hormone into biodegradable microspheres. *Pharm Res, 1997; 14*(6), 730551 735. https://doi.org/10.1023/A:1012142204132
- Murtaugh MP, Baarsch MJ, Zhou Y, Scamurra RW, Lin G. Inflammatory cytokines in animal health and disease. *Vet Immunol Immunopathol, 1996; 54*(1-4), 45-55. https://doi.org/10.1016/s0165-2427(96)05698-x
- 554 73. Dinarello CA. Proinflammatory cytokines. Chest, 2000; 118(2), 503-508.
   555 https://doi.org/10.1378/chest.118.2.503

- Kim KH, Kim KS, Kim JE, Kim DW, Seol KH, Lee SH, Kim YH. The effect of optimal space allowance on growth performance and physiological responses of pigs at different stages of growth. *Animal, 2017; 11*(3), 478-485. https://doi.org/10.1017/S1751731116001841
- Costa LB, Luciano FB, Miyada VS, Gois FD. Herbal extracts and organic acids as natural feed additives in pig diets. *S Afr J Anim*, 2013; 43(2), 181-193. https://doi.org/10.4314/sajas.v43i2.9
- 561 76. Dorman HD, Deans SG. Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *J Appl Microbiol, 2000; 88*(2), 308-316
- 563 77. Lee DY, Li H, Lim HJ, Lee HJ, Jeon R, Ryu JH. Anti-inflammatory activity of sulfur-containing compounds
   564 from garlic. *J Med Food*, 2012; 15(11), 992-999. https://doi.org/10.1089/jmf.2012.2275
- Firgozliev V, Mansbridge SC, Rose SP, Lillehoj HS, Bravo D. Immune modulation, growth performance, and nutrient retention in broiler chickens fed a blend of phytogenic feed additives. *Poult Sci, 2019; 98*(9), 3443-3449. https://doi.org/10.3382/ps/pey472
- 568 79. Bryant MJ, Ewbank R. Some effects of stocking rate and group size upon agonistic behaviour in groups of growing pigs. *British Veterinary Journal*, 1972; 128(2), 64-70. https://doi.org/10.1016/S0007-1935(17)37133-6
- 80. Hemsworth PH, Rice M, Nash J, Giri K, Butler KL, Tilbrook AJ, Morrison RS. (2013). Effects of group size and floor space allowance on grouped sows: Aggression, stress, skin injuries, and reproductive performance. J Anim Sci, 91(10), 4953-4964. https://doi.org/10.2527/jas.2012-5807
- 81. Nannoni E, Martelli G, Rubini G, Sardi L. Effects of increased space allowance on animal welfare, meat and ham quality of heavy pigs slaughtered at 160Kg. PloS One, 2019; 14(2), e0212417. https://doi.org/10.1371/journal.pone.0212417
- 577 82. Turner SP, Ewen M, Rooke JA, Edwards SA. The effect of space allowance on performance, aggression and
  578 immune competence of growing pigs housed on straw deep-litter at different group sizes. Livest Prod Sci.
  579 2000;66:47-55. https://doi.org/10.1016/S0301-6226(00)00159-7
- Menchetti L, Nanni Costa L, Zappaterra M, Padalino B. Effects of Reduced Space Allowance and Heat Stress
  on Behavior and Eye Temperature in Unweaned Lambs: A Pilot Study. Animals, 2021; 11(12), 3464.
  https://doi.org/10.3390/ani11123464
- 583 84. Zhang H, Tsao R. Dietary polyphenols, oxidative stress and antioxidant and anti-inflammatory effects. Curr
   584 Opin Food, 2016; 8, 33-42. https://doi.org/10.1016/j.cofs.2016.02.002
- Pearce GP, Paterson AM. The effect of space restriction and provision of toys during rearing on the behaviour,
  productivity and physiology of male pigs. Appl Anim Behav Sci. 1993;36:11-28.
  https://doi.org/10.1016/0168-1591(93)90095-7

Items	Grower 0-4w	Finisher 4-10w
Ingredients (%)		
Corn	65.10	72.38
Soybean meal	23.90	17.40
Wheat bran	7.00	6.00
Soybean oil	1.00	1.00
L-Lysine	0.10	0.28
DL-Methionine	0.04	0.04
L-T-hreonine	0.03	0.03
Dicalcium phosphate	1.00	1.00
Limestone	1.20	1.25
Salt	0.50	0.50
Vitamin premix <sup>a</sup>	0.08	0.08
Mineral premix <sup>b</sup>	0.05	0.05
Calculated composition		
ME (kcal/kg)	3276	3284
Crude protein (%)	18.00	15.50
Lysine (%)	1.01	0.97
Methionine (%)	0.33	0.29
Calcium (%)	0.78	0.76
Phosphorus (%)	0.62	0.58

Table 1. Ingredients and chemical composition of the basal experimental diets (as fed basis).

Note: ME, metabolizable energy.

<sup>a</sup>Provided per kilogram of complete diet: 20 000 IU of vitamin A, 4000 IU of vitaminD<sub>3</sub>, 80 IU of vitamin E, 16mg of vitamin K<sub>3</sub>, 4 mg of thiamine, 20mg of riboflavin, 6 mg of pyridoxine, 0.08 mg of vitamin B<sub>12</sub>, 120 mg of niacin, 50 mg of Ca-Pantothenate, 2 mg of folic acid, 0.08 mg of biotin.

<sup>b</sup>Provided per kilogram of complete diet: 12.5 mg of manganese, 179 mg of zinc, 140 mg of copper, 0.5 mg of iodine, 0.4 mg of selenium.

Items	PC	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value
BW, kg									
initial	28.00	27.53	27.64	28.97	28.62	28.48	27.88	0.309	0.868
4w	56.30 <sup>ab</sup>	53.23 <sup>b</sup>	53.77 <sup>b</sup>	57.43 <sup>ab</sup>	59.35ª	57.04 <sup>ab</sup>	56.15 <sup>ab</sup>	0.592	0.083
final	110.63ª	96.72°	97.23°	101.37 <sup>bc</sup>	109.43ª	106.41 <sup>ab</sup>	102.39 <sup>bc</sup>	0.767	< 0.001
0-4w									
ADG, kg	0.98 <sup>ab</sup>	0.89 <sup>b</sup>	0.90 <sup>b</sup>	0.98 <sup>ab</sup>	1.06 <sup>a</sup>	0.98 <sup>ab</sup>	0.98 <sup>ab</sup>	0.014	<0.001
ADFI,kg	1.98 <sup>bc</sup>	2.03 <sup>abc</sup>	1.94°	2.01 <sup>abc</sup>	2.06 <sup>ab</sup>	2.08 <sup>a</sup>	2.03 <sup>ab</sup>	0.012	< 0.001
G:F	0.49 <sup>ab</sup>	0.44 <sup>b</sup>	0.46 <sup>ab</sup>	0.49 <sup>ab</sup>	0.51ª	$0.47^{ab}$	0.48 <sup>ab</sup>	0.006	< 0.001
4-10w							$, \bigvee$		
ADG, kg	1.26ª	1.01°	1.01°	1.02°	1.16 <sup>ab</sup>	1.15 <sup>b</sup>	1.08 <sup>bc</sup>	0.013	< 0.001
ADFI, kg	2.92ª	2.54°	2.82 <sup>ab</sup>	2.82 <sup>ab</sup>	2.88 <sup>ab</sup>	2.79 <sup>b</sup>	2.77 <sup>b</sup>	0.016	< 0.001
G:F	0.43ª	$0.40^{\rm abc}$	0.36°	0.36°	0.41 <sup>ab</sup>	$0.41^{ab}$	0.39 <sup>bc</sup>	0.004	< 0.001
Overall period									
ADG, kg	1.16ª	0.97°	0.98°	1.02 <sup>de</sup>	1.14 <sup>ab</sup>	1.1 <sup>bc</sup>	1.05 <sup>cd</sup>	0.009	<0.001
ADFI, kg	2.58ª	2.29°	2.41 <sup>b</sup>	2.43 <sup>b</sup>	2.46 <sup>b</sup>	2.43 <sup>b</sup>	2.41 <sup>b</sup>	0.012	<0.001
G:F	0.45 <sup>ab</sup>	0.43 <sup>cd</sup>	0.41 <sup>d</sup>	0.42 <sup>cd</sup>	0.46 <sup>a</sup>	0.45 <sup>ab</sup>	0.44 <sup>bc</sup>	0.004	< 0.001

Table 2. Effects of different phytogenic feed additives on growth performance in growing-finishing pigs with stressed by stocking density

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; BW, body weight; ADG, average daily gain; ADFI, average daily feed intake; G:F, feed efficiency; SEM, standard error of means. Each value is the mean value of 4 replicates. <sup>a-e</sup>Means within column with different superscripts differ significantly (p < 0.05).

Items	PC	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value
4 week									
DM, %	85.55ª	84.02 <sup>b</sup>	85.72ª	85.71ª	86.14 <sup>a</sup>	85.43 <sup>ab</sup>	85.46 <sup>ab</sup>	0.143	0.03
СР, %	73.35ª	69.45 <sup>b</sup>	73.35 <sup>a</sup>	73.58 <sup>a</sup>	74.47ª	73.16 <sup>a</sup>	72.88 <sup>a</sup>	0.271	0.01
10 week									
DM, %	85.85ª	83.17 <sup>b</sup>	85.84ª	85.75ª	86.93ª	86.15ª	86.11ª	0.179	0.01
СР, %	67.98 <sup>bc</sup>	64.75°	70.58 <sup>ab</sup>	70.65 <sup>ab</sup>	72.87ª	71.37ª	71.22 <sup>ab</sup>	0.398	0.01

Table 3. Effects of different phytogenic feed additives on nutrient digestibility in growing-finishing pigs with stressed by stocking density

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; DM, dry matter; CP, crude protein; SEM, standard error of means. Each value is the mean value of 4 replicates. <sup>a-c</sup>Means within column with different superscripts differ significantly (p < 0.05).

Table 4. Effects of	f different phytogenic	feed additives o	n fecal score in	growing pigs	with stressed by	stocking
density						

Items	PC	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value
4 week									
Fecal score <sup>1</sup>	0.26 <sup>b</sup>	0.76 <sup>a</sup>	0.33 <sup>b</sup>	0.31 <sup>b</sup>	0.30 <sup>b</sup>	0.29 <sup>b</sup>	0.28 <sup>b</sup>	0.017	0.02

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; SEM, standard error of means. Each value is the mean value of 4 replicates. <sup>1</sup>Fecal score was determined as follow : 0, normal feces; 1, soft feces; 2, mild diarrhea; 3, severe diarrhea. <sup>a-b</sup>Means within column with different superscripts differ significantly (p < 0.05).



Items	РС	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value
4 week									
WBC, $10^3/\mu$ l	23.03	23.47	23.18	23.21	22.72	22.99	23.04	0.155	0.93
Lymphocyte, %	45.31ª	36.9 <sup>b</sup>	46.77 <sup>b</sup>	47.47 <sup>ab</sup>	46.37 <sup>b</sup>	41.70 <sup>a</sup>	47.80 <sup>a</sup>	0.506	0.01
Neutrophil, %	39.43 <sup>bc</sup>	48.07ª	39.63 <sup>bc</sup>	38.53 <sup>bc</sup>	36.97°	39.83 <sup>b</sup>	40.03 <sup>b</sup>	0.436	0.03
Basophil, %	0.65	0.70	0.67	0.70	0.63	0.70	0.63	0.027	0.98
Cortisol, ug/dL	1.82°	3.47 <sup>a</sup>	2.60 <sup>b</sup>	2.19 <sup>bc</sup>	1.92°	2.74 <sup>b</sup>	2.78 <sup>b</sup>	0.069	0.01
TNF-α, pg/mL	61.90 <sup>b</sup>	73.13ª	62.63 <sup>b</sup>	62.77 <sup>b</sup>	61.93 <sup>b</sup>	62.67 <sup>b</sup>	62.40 <sup>b</sup>	0.506	0.01
IL-6, pg/mL	72.58	72.20	72.50	72.00	72.37	72.13	72.33	0.254	0.99
10 week									
WBC, 10 <sup>3</sup> /µl	17.74	17.76	17.85	17.64	17.76	17.63	17.72	0.148	0.99
Lymphocyte, %	43.40 <sup>a</sup>	35.90 <sup>b</sup>	45.40 <sup>a</sup>	46.57 <sup>a</sup>	44.37ª	45.63ª	46.87ª	0.492	0.01
Neutrophil, %	42.08 <sup>b</sup>	52.37ª	41.20 <sup>b</sup>	40.83 <sup>b</sup>	42.03 <sup>b</sup>	42.20 <sup>b</sup>	44.13 <sup>b</sup>	0.499	0.01
Basophil, %	0.68 <sup>ab</sup>	0.70 <sup>a</sup>	0.70ª	0.63 <sup>abc</sup>	0.50 <sup>bc</sup>	0.47°	0.47°	0.019	0.01
Cortisol, ug/dL	0.72 <sup>b</sup>	2.40 <sup>a</sup>	0.61 <sup>be</sup>	0.64 <sup>bc</sup>	0.51°	0.67 <sup>b</sup>	$0.61^{bc}$	0.070	0.01
TNF-α, pg/mL	86.70 <sup>b</sup>	99.83ª	85.73 <sup>b</sup>	85.80 <sup>b</sup>	84.90 <sup>b</sup>	85.70 <sup>b</sup>	85.83 <sup>b</sup>	0.596	0.01
IL-6, pg/mL	80.80 <sup>ab</sup>	81.77ª	80.37 <sup>ab</sup>	80.70 <sup>ab</sup>	80.30 <sup>b</sup>	80.43 <sup>ab</sup>	80.53 <sup>ab</sup>	0.132	0.04

Table 5. Effects of different phytogenic feed additives on blood profile in growing-finishing pigs with stressed by stocking density

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; WBC, white blood cell; TNF-  $\alpha$ , tumor necrosis factor- $\alpha$ ; IL-6, InterLeukin-6; SEM, standard error of means. Each value is the mean value of 4 replicates. <sup>a-c</sup>Means within column with different superscripts differ significantly (p < 0.05).

	PC	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value		
Basic behavior (min/hour)											
Feed intake	4.03	4.01	4.03	4.02	4.10	4.10	4.05	0.009	0.02		
Standing	7.05	7.11	6.99	7.12	7.01	6.98	7.13	0.030	0.74		
Lying	44.58	44.16	44.39	44.51	44.09	44.66	44.29	0.106	0.78		
Sitting	4.34	4.72	4.59	4.35	4.80	4.26	4.53	0.087	0.64		
Singularity behavio	or (count/	'hour)									
Drink water	5.04	5.19	5.15	5.10	5.11	5.14	5.12	0.015	0.23		
Rooting	1.08	1.10	1.11	1.04	1.12	1.06	1.03	0.014	0.46		
Posture transition (lying-sitting)	3.54	3.49	3.50	3.48	3.44	3.44	3.51	0.020	0.86		
Posture transition (sitting-lying)	3.53	3.48	3.50	3.48	3.43	3.45	3.50	0.015	0.64		
Biting	0.18 <sup>b</sup>	0.23ª	0.21 <sup>ab</sup>	0.18 <sup>b</sup>	0.15°	0.17 <sup>b</sup>	0.18 <sup>b</sup>	0.05	< 0.001		

Table 6. Effects of different phytogenic feed additives on behavior changes in growing pigs with stressed by stocking density

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; SEM, standard error of means. Each value is the mean value of 4 replicates. <sup>a-b</sup>Means within column with different superscripts differ significantly (p < 0.05).

	PC	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value	
Basic behavior (min/hour)										
Feed intake	4.63 <sup>a</sup>	4.37 <sup>b</sup>	4.67 <sup>a</sup>	4.59ª	4.61ª	4.67 <sup>a</sup>	4.61 <sup>a</sup>	0.020	< 0.001	
Standing	6.77ª	6.32°	6.38°	6.44 <sup>bc</sup>	6.71ª	6.56 <sup>ab</sup>	6.61 <sup>ab</sup>	0.031	< 0.001	
Lying	44.88°	45.51ª	45.21 <sup>b</sup>	45.18 <sup>b</sup>	44.98°	44.99°	44.96 <sup>bc</sup>	0.041	< 0.001	
Sitting	3.72	3.80	3.74	3.79	3.70	3.78	3.82	0.012	0.75	
Singularity beha	vior (coun	t/hour)								
Drink water	5.34	5.28	5.27	5.30	5.44	5.38	5.29	0.026	0.59	
Rooting	1.12	1.08	1.11	1.09	1.21	1.19	1.15	0.021	0.58	
Posture transition (lying-sitting)	3.78	3.43	3.49	3.58	3.71	3.68	3.69	0.035	0.60	
Posture transition (sitting-lying)	3.77	3.41	3.50	3.60	3.70	3.67	3.68	0.045	0.37	
Biting	0.16	0.23	0.18	0.15	0.17	0.16	0.18	0.008	0.20	

Table 7. Effects of different phytogenic feed additives on behavior changes in finishing pigs with stressed by stocking density

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; SEM, standard error of means. Each value is the mean value of 4 replicates. <sup>a-</sup> <sup>e</sup>Means within column with different superscripts differ significantly (p < 0.05).

595



Abbreviation: A, Feed intake; B, Standing; C, Lying; D, Sitting; E, Drink water; F, Posture transition (lying  $\rightarrow$  standing); G, Posture transition (standing  $\rightarrow$  lying); H, Rooting; I, Biting.