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ARTICLE INFORMATION	Fill in information in each box below
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Article Title (within 20 words without abbreviations)	Complete genome sequence of <i>Enterococcus faecium</i> strain AK_C_05 with potential characteristics applicable in livestock industry
Running Title (within 10 words)	Complete genome sequence of <i>Enterococcus faecium</i> strain AK_C_05
Author	Hyunok Doo ^{1#} , Jin Ho Cho ^{2#} , Minho Song ^{3#} , Eun Sol Kim ¹ , Sheena Kim ¹ , Gi Beom Keum ¹ , Jinok Kwak ¹ , Srinivas Pandey ¹ , Sumin Ryu ¹ , Yejin Choi ¹ , Juyoun Kang ¹ , Hyeun Bum Kim ^{1*} , and Ju-Hoon Lee ^{4*}
Affiliation	1 Department of Animal Resources Science, Dankook University, Cheonan 31116, Korea 2 Division of Food and Animal Science, Chungbuk National University, Cheongju 28644, Korea 3 Division of Animal and Dairy Science, Chungnam National University, Daejeon 34134, Korea 4 Department of Food Animal Biotechnology, Department of Agricultural Biotechnology, Center for Food and Bioconvergence, Seoul National University, Seoul 08826, Korea
ORCID (for more information, please visit https://orcid.org)	Hyunok Doo: https://orcid.org/0000-0003-4329-4128 Jin Ho Cho: https://orcid.org/0000-0001-7151-0778 Minho Song: https://orcid.org/0000-0002-4515-5212 Eun Sol Kim: https://orcid.org/0000-0001-8801-421X Sheena Kim: https://orcid.org/0000-0002-5410-1347 Gi Beom Keum: https://orcid.org/0000-0001-6006-9577 Jinok Kwak: https://orcid.org/0000-0003-1217-3569 Srinivas Pandey: https://orcid.org/0000-0002-6947-3469 Sumin Ryu: https://orcid.org/0000-0002-1569-3394 Yejin Choi: https://orcid.org/0000-0002-7434-299X Juyoun Kang: https://orcid.org/0000-0002-3974-2832 Hyeun Bum Kim: https://orcid.org/0000-0003-1366-6090 Ju-Hoon Lee: https://orcid.org/0000-0003-0405-7621
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Authors' contributions Please specify the authors' role using this form.	Conceptualization: Doo H, Kim HB, Lee JH Data curation: Keum GB, Choi Y, Kang J Formal analysis: Kim ES, Kim S, Keum GB, Ryu S Methodology: Cho JH, Song M Validation: Kim S, Kwak J, Pandey S Writing - original draft: Doo H, Cho JH, Song M Writing - review & editing: Doo H, Cho JH, Song M, Kim ES, Kim S, Keum GB, Kwak J, Pandey S, Ryu S, Choi Y, Kang J, Kim HB, Lee JH
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5 **CORRESPONDING AUTHOR CONTACT INFORMATION**

For the corresponding author (responsible for correspondence, proofreading, and reprints)	Fill in information in each box below
First name, middle initial, last name	Hyeun Bum Kim
Email address – this is where your proofs will be sent	hbkim@dankook.ac.kr
Secondary Email address	
Address	Department of Animal Resources Science, Dankook University, Cheonan 31116, Korea
Cell phone number	+82-10-3724-3416
Office phone number	+82-41-550-3653
Fax number	+82-41-565-2940

6

7 **CORRESPONDING AUTHOR CONTACT INFORMATION**

For the corresponding author (responsible for correspondence, proofreading, and reprints)	Fill in information in each box below
First name, middle initial, last name	Ju-Hoon Lee
Email address – this is where your proofs will be sent	juhlee@snu.ac.kr
Secondary Email address	
Address	Department of Food Animal Biotechnology, Department of Agricultural Biotechnology, Center for Food and Bioconvergence, Seoul National University, Seoul 08826, Korea
Cell phone number	+82-10- 9678-5529
Office phone number	+82-2-880-4854
Fax number	+82-2-875-5095

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11 Complete genome sequence of *Enterococcus faecium* strain AK_C_05 with potential characteristics applicable in
12 livestock industry

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14 Hyunok Doo^{1#}, Jin Ho Cho^{2#}, Minho Song^{3#}, Eun Sol Kim¹, Sheena Kim¹, Gi Beom Keum¹, Jinok Kwak¹, Srinivas
15 Pandey¹, Sumin Ryu¹, Yejin Choi¹, Juyoun Kang¹, Hyeun Bum Kim^{1*} and Ju-Hoon Lee^{4*}

16

17

18 ¹ Department of Animal Resources Science, Dankook University, Cheonan 31116, Korea

19 ² Division of Food and Animal Science, Chungbuk National University, Cheongju 28644, Korea

20 ³ Division of Animal and Dairy Science, Chungnam National University, Daejeon 34134, Korea

21 ⁴ Department of Food Animal Biotechnology, Department of Agricultural Biotechnology, Center for Food and
22 Bioconvergence, Seoul National University, Seoul 08826, Korea

23

24

25 # Equal contributors

26

27 * Corresponding authors

28 Hyeun Bum Kim

29 Department of Animal Resources Science, Dankook University, Cheonan 31116, Korea

30 Tel: +82-41-550-3653

31 Email: hbkim@dankook.ac.kr

32

33 Ju-Hoon Lee

34 Department of agricultural biotechnology, Seoul National University, Seoul 08826, Korea

35 Tel: +82-2-880-4854

36 Email: juhlee@snu.ac.kr

37

38 **(Unstructured) Abstract (up to 350 words)**

39 The *Enterococcus faecium* (*E. faecium*) strain AK_C_05 was isolated from cheonggukjang, the Korean
40 traditional food, collected from a local market in South Korea. In this report, we presented the complete genome
41 sequence of *E. faecium* strain AK_C_05. The genome of *E. faecium* strain AK_C_05 genome consisted of one circular
42 chromosome (2,691,319 bp) with a guanine + cytosine (GC) content of 38.3% and one circular plasmid (177,732 bp)
43 with a guanine + cytosine (GC) content of 35.48%. The Annotation results revealed 2,827 protein-coding sequences
44 (CDSs), 18 rRNAs, and 68 tRNA genes. It possesses genes, which encodes enzymes such as alpha-galactosidase (EC
45 3.2.1.22), beta-glucosidase (EC 3.2.1.21) and alpha-L-arabinofuranosidase (EC 3.2.1.55) enabling efficient utilization
46 of carbohydrates. Based on Clusters of Orthologous Groups analysis, *E. faecium* strain AK_C_05 showed
47 specialization in carbohydrate transport and metabolism indicating the ability to generate energy using a variety of
48 carbohydrates.

49 **Keywords (3 to 6):**

50 *Enterococcus faecium*, Livestock, Carbohydrates

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The main text

52
53 The Enterococci bacteria belong to lactic acid bacteria (LAB) group, which can be found in fermented foods[1].
54 Especially, *Enterococcus faecium* is also utilized as probiotics, which could enhance the microbial balance in
55 animals[2]. Despite of safety concerns regarding its use as probiotics, recent research has explored the use of
56 *Enterococcus faecium* as a feed additive for livestock to enhance growth performance[1, 3].

57 In the present study, the *E. faecium* strain AK_C_05 was isolated from homemade cheonggukjang, the Korean
58 traditional food, collected from a local market in Cheonan (36.802917° N, 127.149796° E), Chungcheongnam-do,
59 South Korea. Then, the whole genome sequencing was performed to understand the genomic characteristics of *E.*
60 *faecium* strain AK_C_05 as a potential probiotic in the livestock industry. The *E. faecium* strain AK_C_05 was
61 cultivated in Enterococcosel broth (MBcell, Seoul, South Korea) at 37°C for 24 hours. Genomic DNA was extracted
62 from the cultured *E. faecium* pellet using CTAB DNA extraction method. The complete genome of the *E. faecium*
63 AK_C_05 was sequenced using the Oxford Nanopore Technologies MinION platform at eGnome (Seoul, South
64 Korea). Briefly, library preparation was performed using Native barcoding Sequencing Kit (SQK_NBD114.24, V14)
65 following the manufacturer's instructions (Oxford Nanopore Technologies, Oxford, UK). The prepared library was
66 loaded into the MinION MK1b sequencing device (Oxford Nanopore) equipped with a MinION flow cell (MIN114,
67 R10.4.1, Oxford Nanopore). The Oxford Nanopore sequencing produced 79,247 of long reads, resulting in a total of
68 572,297,864 base pairs. De novo assemble was performed using a Flye assembler v2.9.2, followed by polishing using
69 the Homopolish polisher v0.4.1. The quality of genome assembly was assessed using Quality Assessment Tool for
70 Genome Assemblies (QUAST) v5.2.0[4]. The quantitative assessment of the genome completeness was conducted
71 using the Benchmarking Universal Single-Copy Orthologs (BUSCO) v5.4.6[5]. To annotate and predict the protein
72 coding genes, rRNA, and tRNA genes of *E. faecium strain AK_C_05*, the Rapid Annotation using Subsystem
73 Technology (RAST) v2.0 tool was utilized[6]. The functional categorization of all predicted protein coding genes was
74 performed using the Clusters of Orthologous Groups (COGs)-based EggNOG-mapper v2.0[7]. Furthermore, the
75 presence of virulence factors and antibiotic resistance in *E. faecium strain AK_C_05* was predicted using the BLASTn
76 method, with reference to the Virulence Factor Database (VFDB) and the Comprehensive Antibiotic Resistance
77 Database (CARD)[8, 9].

78 The complete genome of the *E. faecium strain AK_C_05* contain one circular chromosome (2,691,319 bp)
79 with a guanine + cytosine (GC) content of 38.3% and one circular plasmid (177,732 bp) with a guanine + cytosine
80 (GC) content of 35.48%. A total of 2,827 predicted protein-coding sequence, 18 rRNA genes, and 68 tRNA genes

81 were identified in *E. faecium* strain AK_C_05. The most abundant COGs category, excluding Function unknown [S],
82 was Carbohydrate transport and metabolism [G], which accounted for 235 genes, representing 10.4% of the total genes
83 identified. The genome feature and map of *E. faecium* strain AK_C_05 were presented in Table 1, Figure 1A and 1B.

84 Based on its specific focus on carbohydrate transport and metabolism, *E. faecium* strain AK_C_05 possesses
85 genes and enzymes, such as alpha-galactosidase (EC 3.2.1.22), beta-glucosidase (EC 3.2.1.21) and alpha-L-
86 arabinofuranosidase (EC 3.2.1.55), that enable efficient utilization of carbohydrates and the capacity to derive energy
87 from diverse carbohydrate substrates. This characteristic makes *E. faecium* strain AK_C_05 a potential candidate for
88 application in the livestock industry. The complete genome of *E. faecium* strain AK_C_05 has indicated the presence
89 of the antibiotic resistance gene *aac* (6')-Ii in the chromosome and not in the plasmid, confirming that there is no
90 potential for transmission of the resistance gene to other microorganisms. In the plasmid of *E. faecium* strain AK_C_05,
91 the *filA* gene was detected, while no other virulence factors were identified. Interestingly, the *filA* gene's ability to
92 facilitate adhesion to the cell wall is regarded as a beneficial trait for probiotics[10]. Overall, our results indicate that
93 *E. faecium* AK_C_05 could be a promising functional probiotic for improving growth performance in the livestock
94 industry.

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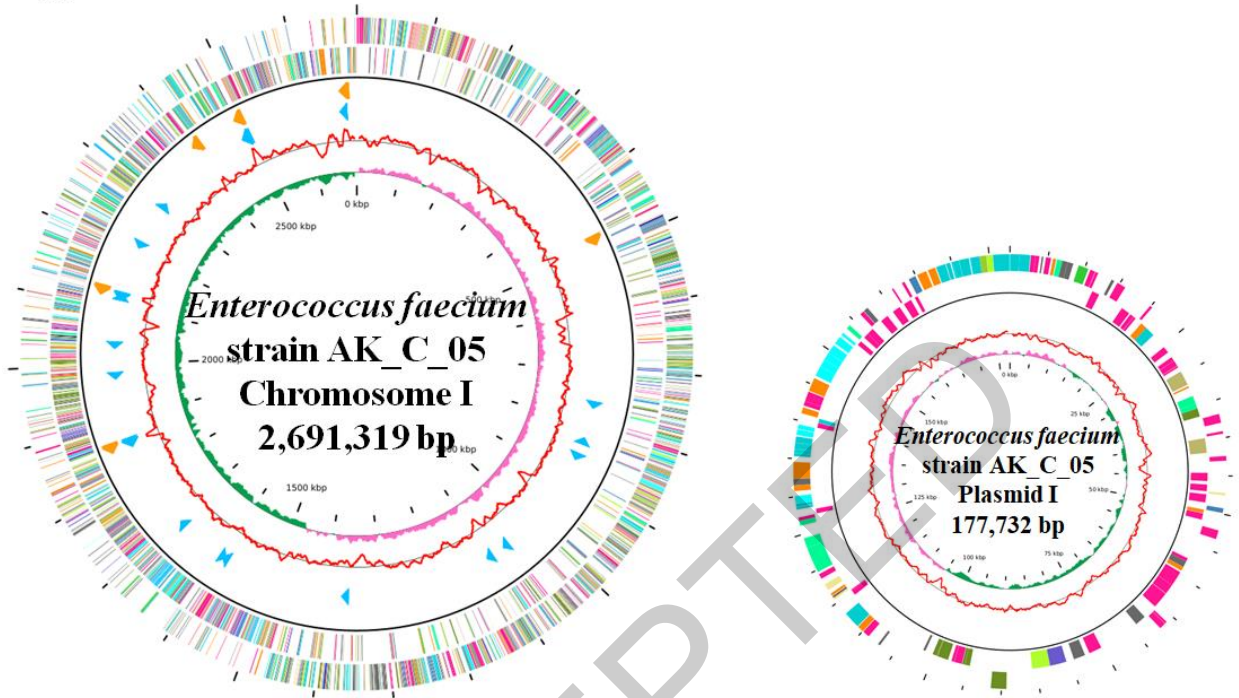
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Tables and Figures

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A



Type	Contig size (bp)	GC-content (%)	Protein-coding genes	tRNA	rRNA
Chromosome	2,691,319	38.3	2,629	68	18
Plasmid	177,732	35.48	198	-	-

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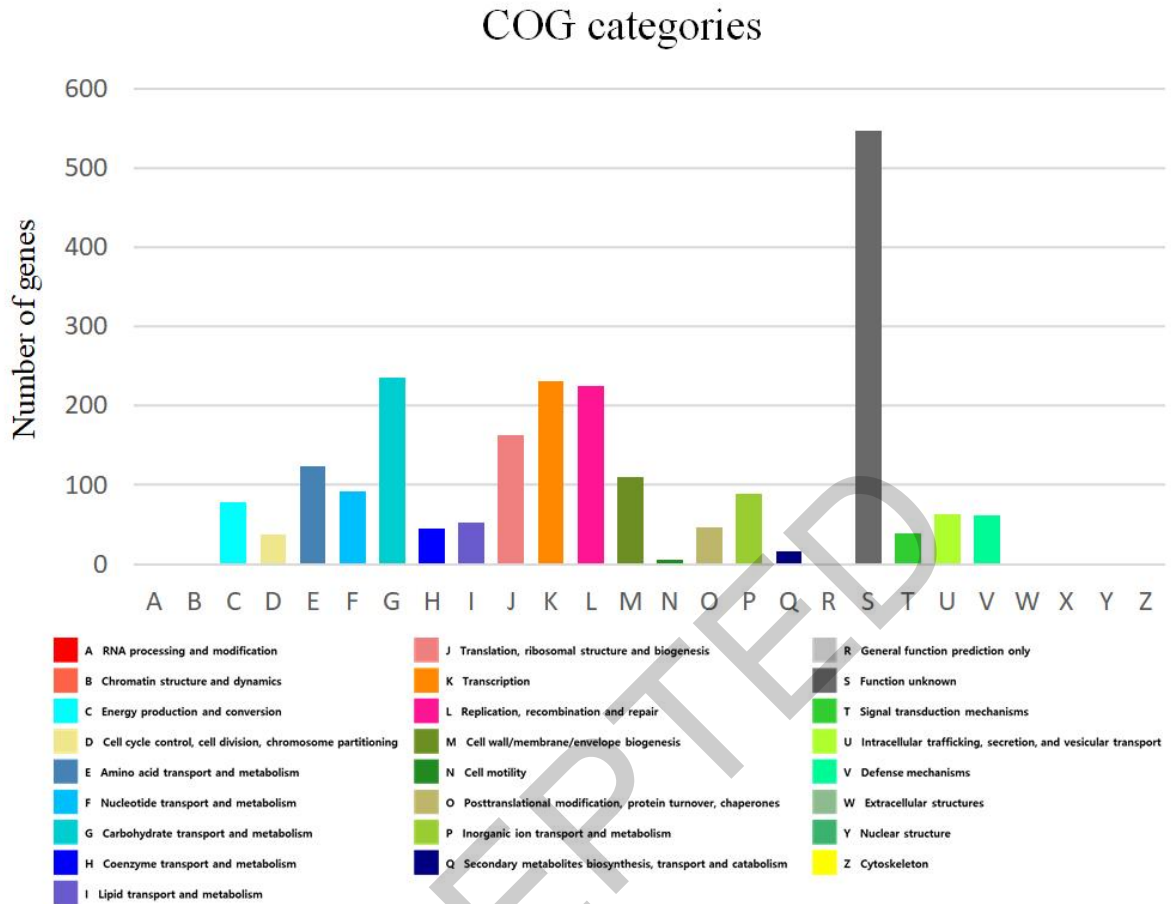
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Figure 1. Genome map of *Enterococcus faecium* strain AK_C_05 and the functional categorization of predicted protein coding genes. The outer ring represents the positions of all annotated gene coding regions (ORFs), while the inner ring in red indicates the guanine + cytosine (GC) content. Peaks in pink and green indicate GC skew. The orange and sky-blue arrows represent rRNA and tRNA operons, respectively. The annotated ORFs are color-coded based on their Clusters of Orthologous Groups (COG) assignments in Figure 1A. The COG functional categories of the predicted protein coding genes are represented in Figure 1B.

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137 **Table 1. Genome features of *Enterococcus faecium* strain AK_C_05**

Property	Term	
	Chromosome	Plasmid
Contig length (bp)	2,691,319 bp	177,732 bp
No. of contig	1 (chromosome)	1 (plasmid)
Guanine + cytosine (G + C)	38.3	35.48
Protein-coding genes	2,629	198
rRNA genes	18	-
tRNA genes	68	-
Genbank Accession No.	CP128995	CP128994

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